

Rezente Pollenspektren der Planalto-Region der Serra
Geral, Rio Grande do Sul/ Brasilien: Rekonstruktion der
rezenten Vegetation – Eine Fallstudie

Dissertation

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1. Berichterstatter:	Prof. Dr. Dr. hc. V. Mosbrugger
2. Berichterstatter:	Prof. Dr. em. H. Walther



Recent palynological spectra from the Planalto region
of the Serra Geral, Rio Grande do Sul/ Brazil:
Reconstruction of contemporary vegetation –
a case study



Thesis for obtaining the doctoral degree in natural sciences

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presented by

certified biologist Kerstin Kriegel

Eidesstattliche Erklärung

Hiermit erkläre ich, **Kerstin Kriegel**, geboren am 12.01.1977, daß ich die vorliegende Arbeit ohne unerlaubte Hilfe und nur mit den in der Arbeit angegebenen Hilfsmitteln angefertigt habe.

Deweiteren bestätige ich, daß bisher von mir noch kein Promotionsversuch an einer anderen Universität unternommen wurde.

Kerstin Kriegel

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Abstract

The relationships between vegetation and recent palynomorphs are examined along a areal of 8000 km² in the state of Rio Grande do Sul, South Brazil. This areal is located between Caxias do Sul (NW) and Cambará do Sul (NE), and Canela/Gramado (SW) and the CPCN Pró-Mata (Centro de Pesquisas e Conservação da Natureza Pró-Mata, SE). Local plant communities of semi-deciduous and deciduous forests, *Araucaria* forests, Capões, gallery forests, Capoeiras, grasslands (Campos) and anthropogenic affected areas were sampled for palynological analysis and plant identification. The distribution of these communities can be related to the equivalent specific regional environmental parameters such as topography, temperature and moisture. The palynomorphs of eighty-five surface-soil samples were classified using TILIA and raster interpolation method ArcGIS 8/ArcMap for visualisation of counted pollen and spore data, cluster analysis and were ordinated using Principal Component Analysis (PCA). Results indicate that the plant communities sampled have distinct pollen and spore assemblages: *Araucaria* forests are characterised by *Dicksonia sellowiana*; semi-deciduous and deciduous forests are characterised by Cyatheaceae; Capões are characterised by *Podocarpus sp.*, Poaceae, Euphorbiaceae, *Ilex sp.* and *Drymis brasiliensis*, gallery forests are also characterised by *Podocarpus sp.*, Poaceae, Euphorbiaceae and *Ilex sp.* but in lower amounts, and grasslands are characterised by Poaceae, Anthocerotaceae, Cyperaceae and Asteraceae. While Capoeiras are characterised by pollen and spores of the previous (original) plant communities, anthropogenic affected areas are characterised by pollen and spores of actual plant communities. The raster interpolation method shows that two spore types (*Dicksonia sellowiana* and Cyatheaceae) are sufficient to distinguish *Araucaria* forests, semi-deciduous and deciduous forests. Cluster analysis and PCA revealed that palynological spectra permit to interpret the distribution of vegetation. Applied on fossil assemblages, they allow an interpretation of the vegetation history.

Zusammenfassung

In der vorliegenden Arbeit waren die Beziehungen zwischen Vegetation und der rezenten Palynomorphae in Oberflächenbodenproben Gegenstand der Untersuchungen. Als Untersuchungsgebiet wurde ein 8000 km² großes Areal im Bundesstaat Rio Grande do Sul, Brasilien ausgewählt, das sich zwischen den Ortschaften Caxias do Sul (NW) und Cambará do Sul (NO) sowie Canela/Gramado (SW) und dem Pró-Mata – Schutzareal (CPCN, Centro de Pesquisas e Conservação da Natureza Pró-Mata, SO) erstreckte. In dieser Region lassen sich 7 pflanzensoziologische Einheiten unterscheiden – subtropisch Halbimmergrüne Feuchtwälder und Laubabwerfende Trockenwälder, *Araukarien*-Wälder, inselartige *Araukarien*wälder (Capões), Galleriewälder, verbuschte Grasländer (Capoeiras), offene Grasländer (Campos) und stark anthropogen überprägte Flächen. Aus den Oberböden der in dem Untersuchungsgebiet anzutreffenden Vegetationstypen wurden die Pollen- und Sporengesellschaften isoliert und sowohl qualitativ als auch quantitativ analysiert. Außerdem wurde für jede Fläche eine bodengestützte Vegetationskartierung nach der Methode von BRAUN-BLANQUET durchgeführt. Die Verteilung dieser pflanzensoziologischen Einheiten ist stark an die abiotischen Parameter der Region, wie Topographie, Temperatur und Feuchtigkeit, gekoppelt. Insgesamt wurden 85 Oberflächenbodenproben quantitativ und qualitativ palynologisch ausgewertet. Die Ergebnisse wurden mit Hilfe der TILIA-Software und der Raster-Interpolations-Methode von ArcGIS 8/ArcMap visualisiert. Außerdem wurden die Ergebnisse aus der palynologischen Untersuchung einer multivariaten statistischen Auswertung – Clusteranalyse, Hauptkomponentenanalyse (Principal Component Analysis, PCA) - unterzogen. Die Ergebnisse zeigen, daß jede der unterschiedenen pflanzensoziologischen Einheiten ein spezifisches palynologisches Spektrum aufweist: *Araukarien*wälder können mit Hilfe der Sporen des Baumfarnes *Dicksonia sellowiana* charakterisiert werden; in den halbimmergrünen und laubabwerfenden Wäldern sind Sporen einer zweiten Baumfarn-Familie, die der Cyatheaceae, dominant; *Podocarpus sp.*, Poaceae, Euphorbiaceae, *Ilex sp.* und *Drymis brasiliensis* sind typisch für Capões; Galleriewälder werden ebenfalls von den Pollen von *Podocarpus sp.*, Poaceae, Euphorbiaceae und *Ilex sp.* dominiert jedoch zu einem

deutlich geringeren Anteil als in Capões; und für Grasländer sind die Pollen der Poaceae, Anthocerotaceae, Cyperaceae und Asteraceae charakteristisch. Während in den Oberflächenbodenproben der Capoeira-Flächen vor allem Palynomorphae der vormals anzutreffenden Vegetation nachweisbar sind, werden die stark anthropogen überprägten Flächen durch Pollen und Sporen der aktuellen Vegetation charakterisiert. Die angewendete Raster-Interpolations-Methode hat deutlich gezeigt, daß zwei Sporen-Typen (*Dicksonia sellowiana* und Cyatheaceae) ausreichend sind, um *Araukarienwälder*, halbimmergrüne und laubabwerfende Wälder palynologisch voneinander zu unterscheiden. Die Clusteranalyse und die PCA haben verdeutlicht, daß palynologische Spektren eine Interpretation der Vegetationsverteilung gestatten. Wird diese Methode auf fossile Proben angewendet, erlaubt sie eine Interpretation der Vegetation in Raum und Zeit.

Resumo

No presente trabalho, foram analisadas as relações entre vegetação recente e palinomorfos de solo, em uma área de 8.000 Km² no Estado do Rio Grande do Sul, Região Sul do Brasil, localizada entre Caxias do Sul (limite Noroeste – NO), Cambará do Sul (limite Nordeste – NE), Canela/Gramado (limite Sudoeste – SO) e o Centro de Pesquisas e Conservação da Natureza Pró-Mata (CPCN Pró-Mata, limite Sudeste – SE). Para o desenvolvimento do estudo foram coletadas amostras de solo para análise palinomórfica e identificadas as plantas ocorrentes em cada ponto. Na área de estudo ocorrem as seguintes Comunidades Vegetais, cuja distribuição pode ser relacionada com fatores ambientais como topografia, temperatura e umidade: Floresta Estacional Semidecidual; Floresta Estacional Decidual, Mata de Araucária (Floresta Ombrófila Mista); Capões; Mata de Galeria; Capoeira; Campo e Áreas de Cultivo. Os palinomorfos de 85 amostras de solo foram classificados com a utilização do Programa TILIA e o *Método de interpolação* matricial (*Raster Interpolation Method, ArcGIS 8/ArcMap*), possibilitando a visualização dos dados palinomórficos, os quais posteriormente, foram analisados por *Cluster* e ordenados por *Principal Component Analysis (PCA)*. Os resultados indicam que as diferentes Comunidades Vegetais amostradas possuem associações distintas de polens e esporos. A Mata de Araucária é caracterizada pelo domínio de esporos de *Dicksonia sellowiana*; as Florestas Estacional Semidecidual e Estacional Decidual pelo de esporos de Cyatheaceae; Capões pelo de polens de *Podocarpus* sp., Poaceae, Euphorbiaceae, *Ilex* sp. e *Drymis brasiliensis*; Mata de Galeria pelo de polens de *Podocarpus* sp., Poaceae, Euphorbiaceae e *Ilex* sp., todavia em quantidades consideravelmente menores que a anterior; e Campos pelo de esporos de Anthocerotaceae e polens de Poaceae, Cyperaceae e Asteraceae. De forma bastante diferenciada, as Capoeiras são caracterizadas por palinomorfos das Comunidades Vegetais que se desenvolviam nesses locais anteriormente. As Áreas de Cultivo são caracterizadas por palinomorfos das Comunidades Vegetais circundantes atuais. O *Método de interpolação* mostra que dois tipos de esporos (*Dicksonia sellowiana* e Cyatheaceae) são suficientes para distinguir Mata de Araucária, Floresta Estacional Semidecidual e Floresta Estacional Decidual. *Cluster Analysis* e *PCA* revelam que os espectros polínicos permitem inferir as Comunidades

Vegetais. Esses métodos também, quando aplicados em associações palinomórficas fósseis, permitem estabelecer a composição da vegetação em tempos remotos.

1. Introduction

Brazil offers an interesting investigation area with respect to its bountiful and impressive flora and fauna. Since the beginnings of the 19th century, many expeditions were undertaken to research Brazil. The scientifically interested Earl KLEMENS WENZEL LOTHAR NEPOMUK VON METTERNICH (1773–1859), foreign secretary of Austria around 1800, organised an expedition to Brazil from 1817 to 1821. Noteworthy scientists like the botanist HEINRICH WILHELM SCHOTT (1794-1865), the botanist and mineralogist JOHANN BAPTIST EMANUEL POHL (1782-1834) and the zoologist JOHANN NATTERER (1787-1834) accompanied him. Since the 19th century the vegetation of South Brazil has been meticulously explored. The first scientist who studied the vegetation of Rio Grande do Sul was AUGUSTE DE SAINT-HILAIRE (1779-1853). Numerous scientists followed. LINDMAN (1906), KLEIN (1960, 1961, 1964, 1984), RAMBO (1945, 1956) and VELOSO & KLEIN (1957, 1959, 1961, 1968) are being mentioned exemplary.

During the 20th century the original forest vegetation of Rio Grande do Sul has been nearly completely cleared. Thus, the knowledge about the original vegetation is fragmentary and a reconstruction is difficult (LEITE 2002, OVINGTON 1983). Today, the vegetation of Rio Grande do Sul is strongly affected by agro-forestal use. Therefore, the environmental agency of Brazil (IBAMA) placed the forests of Rio Grande do Sul in large part under protection by the Brazilian legislation since 1998.

The identification of vegetation proves itself as being complicated as a result of the density of vegetation and the tree size. The height of trees renders the collection of leave and blossom material impossible for plant identification. Mapping of vegetation according to BRAUN-BLANQUET (1964), as applied in Europe, is hence difficult to realize. Therefore, recently the plant covering is identified by remote sensing. But this method holds errors regard to the resolution, therefore mapping of vegetation in the field is indispensable.

Since the 20th century the correlation between contemporary vegetation and palynological spectra and the history of vegetation of South America, especially of Brazil, has received growing attention. GRAF (1992) undertook the effort to reconstruct the palaeoenvironment of the Andes based on palynological analysis. BEHLING (e.g. BEHLING 1993, 2001, 2002, 2003, BEHLING & NEGRELLE 2001, BEHLING et al. 2000, 2001, 2004) analysed meticulously the palaeoenvironment of Brazil. He extracted and analysed palynomorphs from cores of peat, lakes and other organic deposits. Determined pollen and spores of the deposits were used to provide better insight into previous vegetation. Grassland ecosystems (their composition, biodiversity and distribution) and advices of human impacts on the vegetation received special attention because different opinions exist. LINDMANN (1906) has already noticed that the forest-grassland-pattern of Rio Grande do Sul does not present the climax. According to his opinion the climate would generate a more widespread forest. Some ecologists (e.g. DE QUADROS & PILLAR 2002,) explain the grassland-forest-mosaic of South Brazil as a result of human impacts. Unlike this position, others explain this phenomenon as a result of climatic changes especially during Late Quaternary times (BEHLING 1995, 2001, LORSCHBITTER 1988). Knowledge of palaeograssland distribution and composition may help to improve models of past vegetation and climate in these tropical and subtropical regions.

The present thesis may represent a possibility to simplify the reconstruction method. This work is attached to the thesis of EBNER (2005), who developed and tested the method in the biological reserve “Pró-Mata” (near São Francisco de Paula/ RS, Brazil). The method is based on a palynological analysis of topsoil (analysis of “recent” pollen and spores) in relationship with the contemporary vegetation. Local plant communities of *Araucaria* forests, semi-deciduous and deciduous forests, islands of *Araucaria* forest (Capões), gallery forests, scrubland (Capoeiras), grassland (Campo) and anthropogenic affected area were sampled for pollen analysis and the contemporary vegetation was identified according to a simplified BRAUN-BLANQUET-technique. A special pollen spectrum for each explored vegetation type was aimed to be detected. These spectra may eventually be utilised by future scientists in order to reconstruct palaeoenvironment. In this work emphases were laid on the following 5 questions.

Objectives

1. Are the different vegetation types of the plateau of Rio Grande do Sul (Planalto) and the slopes of the Serra Geral distinguishable from each other by pollen spectra extracted from topsoil and if so, to what extent?
2. If the first objective can be answered with YES, are discriminatory pollen/ spore type detected characterising a certain vegetation type?
3. Is EBNER`s (2005) proposition, as postulated on the basis of his research in the Pró-Mata area, applicable to a larger area in the region of the Planalto and Serra Geral?
4. How detailed does a topsoil pollen spectrum represent the contemporary plant covering?
5. To what extent is a reconstruction of the original plant formations and their distribution in former time (under application of the drilling method) feasible?

2. Material and methods

2.1. Study area

2.1.1 Geographical position

The present study was carried out in the state of Rio Grande do Sul, South Brazil. The study site was defined between the longitudinal values of 50°50'33.60" and 51°09'53.94" and latitudinal values 29°18'59.40" and 29°31'10.70", including the municipalities Cambará do Sul, Canela, Caxias do Sul, Gramado, Jaquirana, Linha Nova, Nova Petropolis and São Francisco de Paula (Fig. 1). The limits of the study site corresponded to Caxias do Sul at northwest, Cambará do Sul at northeast, Canela/Gramado at southwest and the biological reserve "Pró-Mata" (Centro de Pesquisas e Conservação da Natureza Pró-Mata) at southeast. To consider the regional variability of the different vegetation types, the observed area was divided from East to West in three sub-areas (Fig. 1, red lines).

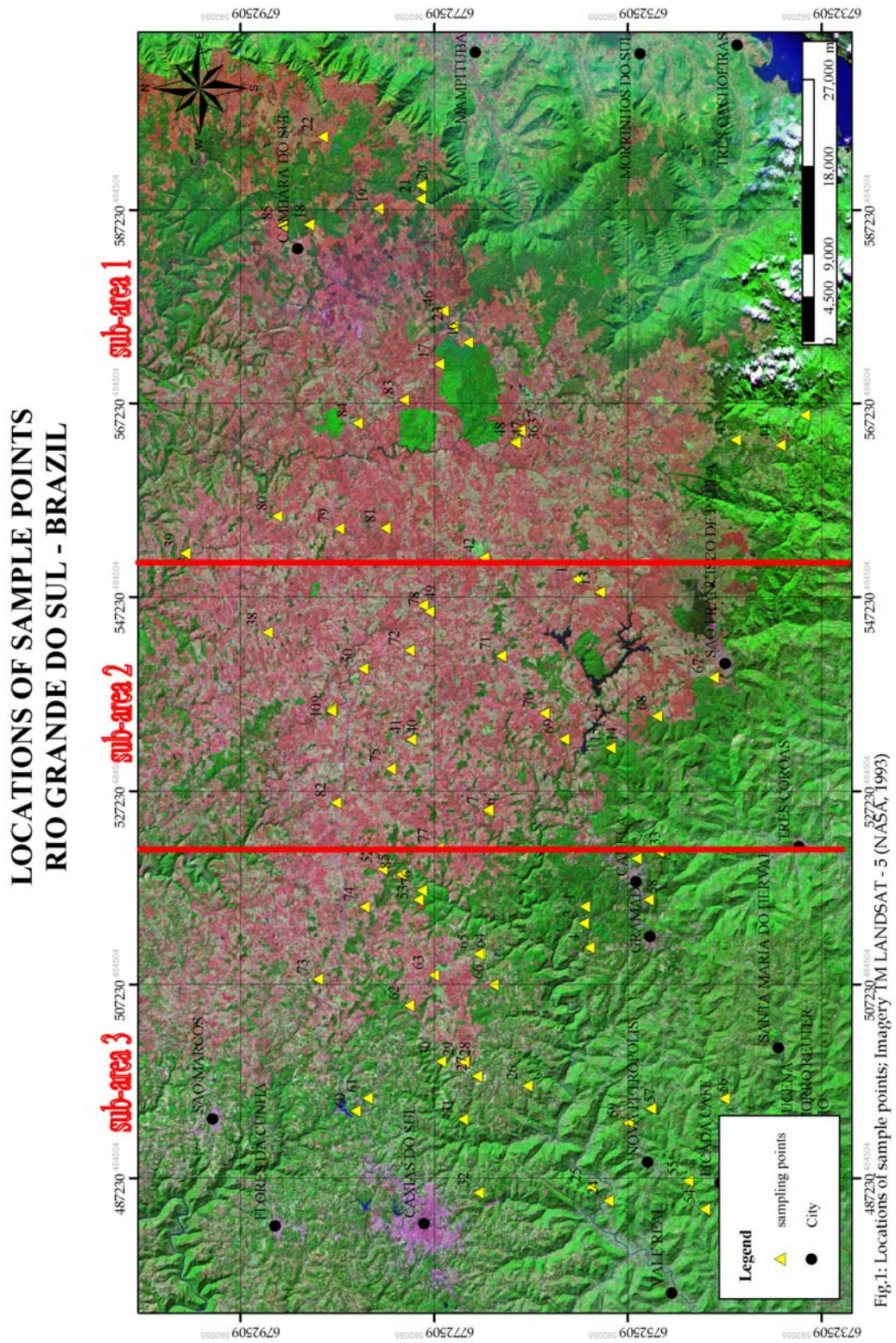


Fig. 1: Locations of sample points; Imagery TM LANDSAT – 5 (NASA, 1993)

2.1.2 Landscape, geomorphology and soil

The study area was located at the top of the South Brazilian plateau, *Planalto* (HUECK 1966) and its slopes. The cleft and steep rugged slopes of the Planalto (Fig. 2) form a distinct geomorphological unit, the so-called **Serra Geral**. The Planalto is formed by ancient sedimentary rocks mainly *sandstone* from the Botucatu–formation (Jurassic/Cretaceous) and is covered by *trap basalt* (IBGE 1986, LEINZ 1949, ROISEMBERG 1990).



Fig. 2: *Cleft slopes of Serra Geral; transition from Araucaria forest (top) to Mata Atlântica, photo: M. EBNER*

The ground is subject to profound weathering. These activities result in elevated acidification, a slow base saturation and cation exchange capacity. Laterites and limonitic surface crusting are common phenomena not only in grassland but also in humid forests (SCHULTZ 2000). The pedogenesis is mainly influenced by the humid subtropical climate with high precipitation rates.

According to SCHULTZ (2000), the soils in the studied region belong to the Acrisol-zone. According to Brazilian Soil Classification (SIBCS, EMBRAPA 1999) and STRECK et al. (2002) five soil types are found in the area (Fig. 3):

- 1) **Argisol** occurs predominantly in Rio Grande do Sul and corresponds to Podsol in the Russian Systematics or to Ultisol in US Soil Taxonomy (1975). Argisols are moderately profound, well drained soils. Their main characteristic is the presence of a textural B-horizon. The B-horizon is always more clayish than the horizons above and below it. A-Bt-C is a typical horizon sequence, where Bt represents the textural B-horizon.
- 2) **Alisol** originates from rhyolite, occurs in the region Caxias do Sul and shows an elevated content of aluminium. Usually, Alisol is moderately profound.
- 3) **Cambisol** occurs in areas with high rates of rainfall and low temperatures. Cambisols are shallow and well drained soils still including fragments of the parent material in their upper horizons. Cambisol is typical for the regions of the Planalto and Caxias do Sul. Their horizon sequence is A-Bi-C, where Bi represents the B incipient horizon. These soils are similar to Tropepts in Soil Taxonomy (1975).
- 4) **Chernossol** is a shallow or profound and less coloured soil. It contains at least 18% iron, as well as high concentrations of calcium and magnesium.
- 5) **Neosol (Litholic Neosol)** is a “young” soil that contains many primary minerals and high amounts of silt, also in the surface horizon. It is present on accentuated relief in the regions of Caxias do Sul and the Planalto. Litholic Neosol was previously classified as Litholic Soil. Like Cambisol, Litholic is widely distributed in Brazil and is shallow and sometimes stony.

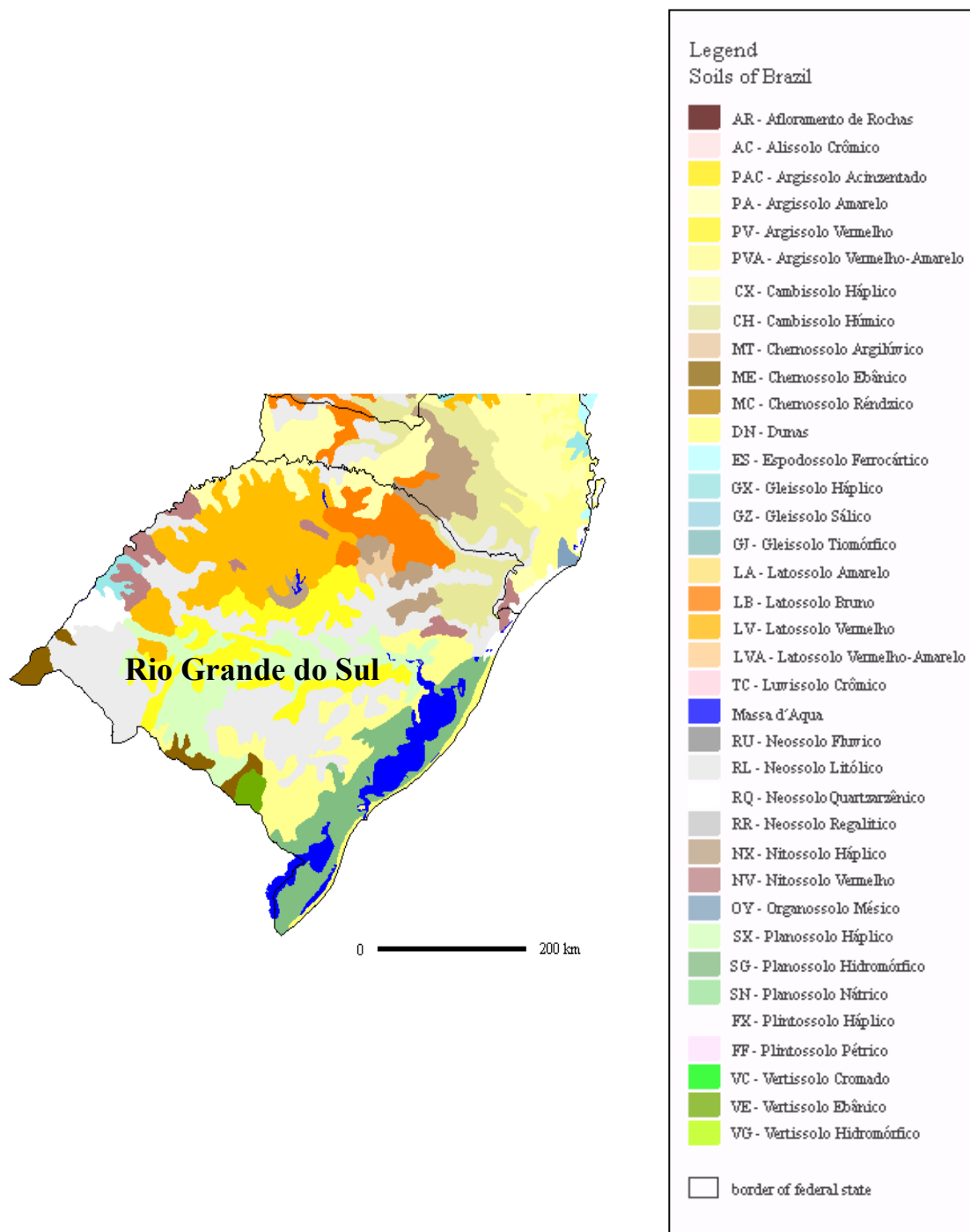


Fig. 3: Soils of Rio Grande do Sul according to the Brazilian Soil Classification (EMBRAPA 1999, IBGE 2005)

2.1.3 Climate

The humid subtropical climate predominates in South Brazil (SCHULTZ 2000). It varies from humid to moderate climate and lacks dry periods (FÄHSER 1985). Precipitation is influenced by tropical cyclones, zenith rainfall and maritime trade winds on the one hand and sea current (Brasil stream) on the other hand. These factors interact upon the climate of South Brazil (IBGE 1986, GOLTE 1993). The study area was located on the eastern side of the South American continent therefore the permanent high pressure determines the climate with maximum rainfall in summer (LAUER & RAFIQPOOR 2002). The climate causes a longitudinal humidity-vegetation-gradient from the East to the West, with decreasing precipitation from the coast westward. While the climate, at lower regions of Rio Grande do Sul and the slopes of Serra Geral, corresponds to type Cfa in the classification of KÖPPEN (1923), the Planalto is characterised by the temperate climate type Cfb1 (MORENO 1961, NIMER 1989) and Bslph γ (LAUER & RAFIQPOOR 2002). Reference values from São Francisco de Paula can be applied to describe the climate of the explored area. In São Francisco de Paula the average annual temperature is 14.5°C, July being the coldest month (10.3°C); January the warmest (18.8°C). São Francisco de Paula features an average precipitation of 2252mm annually. The precipitations are dispersed evenly over the year (Fig. 4). Polar air masses are responsible for pleasant temperatures all over the year. Frost and snow are rare climatic phenomena (GOLTE 1993, HUECK 1966, MORENO 1961). Due to the subtropical climate, various vegetation types are able to grow within a small regional scale. Rainforests occur beside semi-deciduous and deciduous forests as well as savanna and grassland.

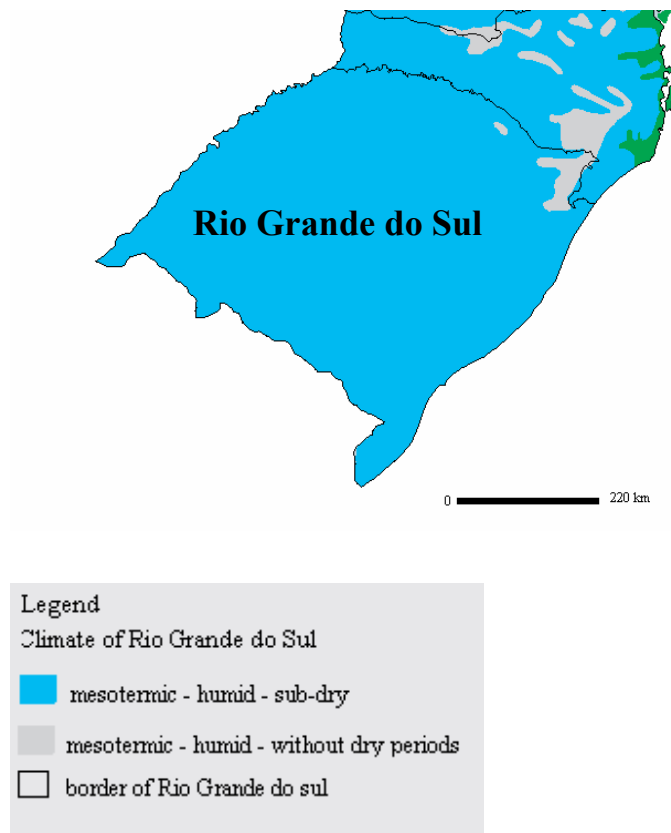


Fig. 4: *Climate of Rio Grande do Sul according to IBGE (2005)*

2.1.4 Vegetation in the study region

Rio Grande do Sul belongs to the ecozone of the Evergreen Subtropics (SCHULTZ 2000). The Evergreen Subtropics are located between the latitudes 25° and 35°, and are solely located on the east side of continents.

Rio Grande do Sul is characterised by an interesting mosaic of grassland and forest vegetation. A transition of *Araucaria* forest, broad-leaved evergreen forest (in the Anglophone literature also named warm-temperate, e.g. OVERTON 1983) and semi-deciduous forest predominates the northern territory of the state. The southern territory is dominated by grassland (HUECK & SEIBERT, 1972, KLEIN, 1975 and TEIXEIRA et al., 1986).

The total territory of the state of Rio Grande do Sul includes 132.103 km² (46.73%) of grassland and 49.557 km² (17.53%) of forest (SEMA 2001). The forest ecosystems have been strongly cleared during the last century. Today, the vegetation of Rio Grande do Sul is strongly affected by agro-forestal use. Thus, the knowledge about the original vegetation is only fragmentary and a reconstruction is difficult (HUECK 1956, 1966, OVINGTON 1983).

The modern potential natural vegetation cover of the study site is composed of voluptuous broad-leaved evergreen forest which is located on the coastal eastern slopes of the Serra Geral. These slopes are characterised by high amounts of precipitation during all season. As a result of lower amounts of precipitation westwards, this forest type turn into a semi-deciduous forest and then into a deciduous forest. The Planalto is predominantly covered by subtropical grassland, which in Brazil is called 'campos'. Patches of these species-rich grasslands form a mosaic with *Araucaria* forests (Fig. 5).



Fig. 5: Mosaic of grassland (Campo) and islands of *Araucaria* forest (Capões)

Different methods were applied to classify the vegetation of the Neotropics, especially of South Brazil (DRUDE 1890, RAMBO 1956, LINDMANN 1906, HUECK and SEIBERT 1972). Recent publications refer to a nomenclature proposed by VELOSO & GÓES-FILHO (1982) which was further modified and applied by JOLY et al. (1999), IBGE (1986) and others. In this system the vegetation of South Brazil is divided into phyto-ecological regions. A phyto-ecological region is characterised by an original, well-defined floristic composition, by uniform climate and similar geomorphology, whereas the geology may be different (IBGE 1986).

In the present study, seven different vegetation types were considered as follow (Fig. 6, 7):

1. **Semi-deciduous forest (Floresta Estacional Semidecidual)**
2. **Deciduous forest (Floresta Estacional Decidual)**
3. ***Araucaria* forest (Floresta Ombrófila Mista, Mata de Araucária)**
4. **Islands of *Araucaria* forest (Capões, Matas insulares)**
5. **Gallery forest (Mata de Galeria).**
6. **Grassland (Campo and Capoeira)**
7. **Anthropogenic disturbed area**

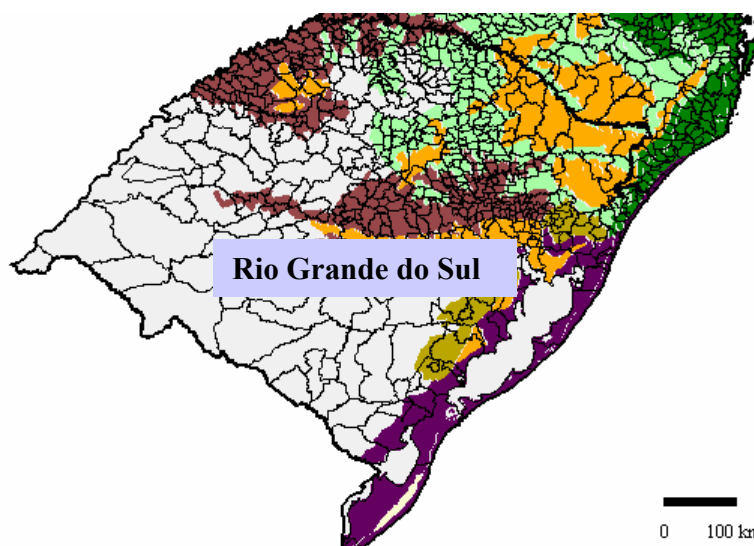


Fig. 6: *Vegetation of Rio Grande do Sul (SOS Mata Atlântica, 2003, slightly modified, legend: below Fig. 7)*

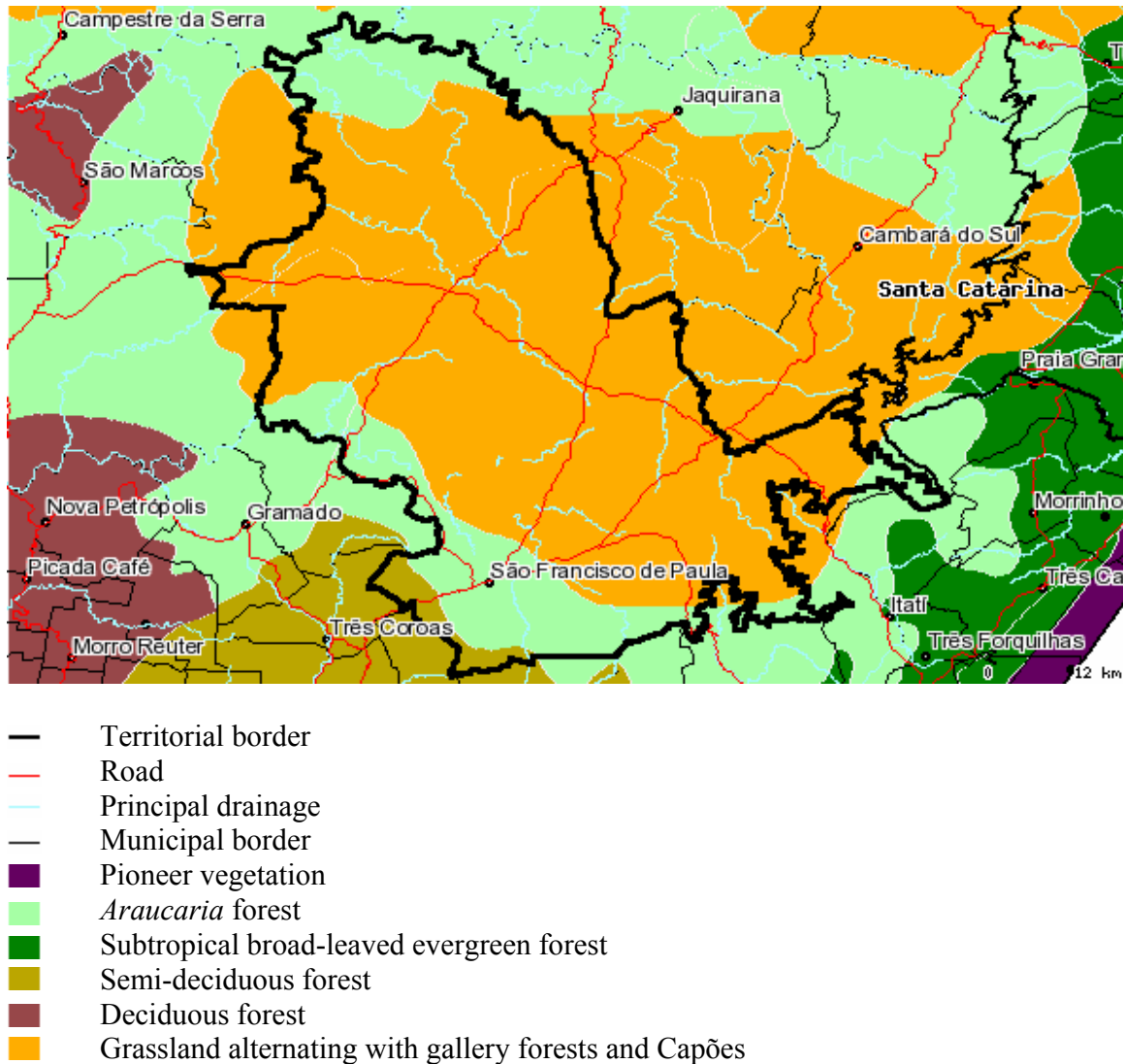


Fig. 7: Overview of the vegetation of the study area (SOS Mata Atlântica, 2003, slightly modified)

1. Semi-deciduous forest (Floresta Estacional Semidecidual)

This forest type is found exclusively on the Serra Geral slopes at low altitudes. Southward it is limited by pasture and to the north by *Araucaria* forest and the grassland of the Planalto. Within this geographical range the semi-deciduous forest may vary considerably. While the forest in the valley of the river Maquiné assumes a more subtropical character

like the Atlantic rainforest, the forests of the Canela region and parts of the river Cai valley present a blend between a semi-deciduous forest and a deciduous forest.



Fig. 8: *Subtropical forest in the valley of the river Cai*

This forest type resembles tropical forest regarding the floristic composition and the occurrence of lianas. A feature of the semi-deciduous forest is a clear understorey structure. The upper tree storey contains strong trees forming dense canopy. This storey is mainly composed by species of the genera of Lauraceae *Ocotea* and *Nectandra*, *Cabralea canjerana* (Meliaceae), *Cedrela fissilis* (Meliaceae) and *Luehea divaricata* (Tiliaceae). The medium storey generally contains *Actinostemon concolor* (Euphorbiaceae), *Sorocea bonplandii* (Moraceae) as well as *Euterpe edulis* (Palmito, Arecaceae) whose “hearts” are an appreciated delicacy. The subjacent shrub storey contains high shrubs and small trees. *Trichilia elegans* (Meliaceae), *Mollinedia* (Monimiaceae), *Rudgea* (Rubiaceae), *Psychotria*

(Rubiaceae), *Piper* (Piperaceae) and *Celtis* (Ulmaceae) are the dominant species. Lianas render the forest its typical subtropical character. Species of *Mikania* (Asteraceae), *Aristolochia* (Aristolochiaceae, Fig. 9), *Smilax* (Smilacaceae) and several species of the families Bignoniaceae and Leguminosae appear frequently. Another species is the “stairs of monkey” (*Bauhinia langsdorffiana*) named after its zigzag growth, belonging to the Caesalpinoidae.

This high plant diversity was already referred by KLEIN (1964, 1984) and VELOSO & KLEIN (1957, 1959). They described meticulously the composition of several formations of this forest type and underlined this aspect with numerous species.



Fig. 9: *Aristolochia* sp. (Aristolochiaceae)

Tree ferns like *Alsophila* spp. and *Nephelea* spp. (Cyatheaceae) appear on humid ground (Fig. 10 and 11).



Fig. 10: Forest with tree ferns



Fig. 11: Stem of a tree fern

Epiphytes are significant components of semi-deciduous forests. Most of the epiphytes belong to the orchids, e.g. species of the genera *Cattleya* and *Oncidium*. Further on, various cactus species, e.g. of *Rhipsalis* and bromelias like *Tillandsia* and *Vriesea* are abundant. The prevalence of bryophytes and lichens is obvious.

2. Deciduous forest (Floresta Estacional Decidual)

Upcountry semi-deciduous forest merges into deciduous forest. The River Caí valley in the regions of Nova Petropolis, Picada Café and Morro Reuter is interesting because it is characterised by a mélange of semi-deciduous and deciduous forests (Fig. 7). Controversial studies exist about the distinct `limits` between these forest types.

The floristic inventories of semi-deciduous and deciduous forests are similar to subtropical broad-leaved forests but they also include deciduous floristic elements (IBGE 1986). While 20-50% of the plants are deciduous in semi-deciduous forests, deciduous forests include more than 50% deciduous plants.

3. *Araucaria* forest (Floresta Ombrófila Mista, Mata de Araucária)

The *Araucaria* forest (Fig. 12) is formed by the *Araucaria angustifolia* (pinheiro-do-paraná), which due to its habit is the main characteristic tree of the Planalto. According to HUECK (1952) and REITZ & KLEIN (1966) the *Araucaria* forest is the predominant forest formation on the Planalto of Rio Grande do Sul state. The *Araucaria* forest occupies regions at an altitude between 500m in the West and 1000m in the East. The *Araucaria* forests are interlocked by grassland and the landscape therefore presents the typical mosaic pattern.



Fig. 12: *Araucaria* forest

The composition of the storey in *Araucaria* forest shows a specific pattern. The upper storey consists of *Araucaria angustifolia* in association with some species of Lauraceae as *Cryptocarya aschersoniana*, *Ocotea pulchella* and *Ocotea puberula* (LÖTSCHERT 1981, IBGE 1986). The lower tree storey is dominated by trees of *Drymis brasiliensis* (Winteraceae), *Schinus polygamus* (Anacardiaceae), *Cupania vernalis* (Sapindaceae), *Lamanonia speciosa* (Cunnoniaceae), *Berberis laurina* (Berberidaceae) and *Podocarpus lambertii* (Podocarpaceae) (HUECK 1956, RAMBO 1956). Myrtaceae occur in an extremely high diversity of species, some of which differ slightly from each other and are therefore difficult to distinguish. Typical genera of the lower tree storey and the shrub storey are *Myrceugenia*, *Myrcia*, *Eugenia* and *Calyptranthes*. Epiphytes, like *Tillandsia usneoides* (Bromeliaceae), play an important role because they provide the unique character of a virgin forest. Some of the understorey plants belong to the *andin–subantarctic* elements (RAMBO 1951) (Fig. 13, 14).



Fig. 13: *Drymis brasiliensis* (Winteraceae)



Fig. 14: *Fuchsia regia* (Onagraceae)



Fig. 15: *Sophronites coccinea* (Orchideaceae)

The huge, conspicuous tree fern *Dicksonia sellowiana* (Dicksoniaceae) occurs occasionally on humid ground.

The *Araucaria* forests of the Planalto in the region between the River Taquari and the spring of the river Sinos are different. They are higher and more sumptuous than the *Araucaria* forest on the border of “Aparados da Serra” and contain more elements of the broad-leaved evergreen forest (RAMBO 1956). *Ilex paraquariensis* (Aquifoliaceae), *Drymis brasiliensis* (Winteraceae), *Lithraea brasiliensis* (Tiliaceae), *Sebastiania commersoniana* (Euphorbiaceae), *Zanthoxylum rhoifolium* (Rutaceae), *Prunus sellowii* (Rosaceae), different species of *Roupala* (Proteaceae), *Matayba elaeagnoides* (Sapindaceae), *Campomanesia xanthocarpa* (Myrtaceae), *Eugenia uniflora* (Myrtaceae) and different species of the genera *Nectandra* and *Ocotea* (both Lauraceae) could be listed as examples for the high diversity in these *Araucaria* forests.

4. Islands of *Araucaria* forest (Capões, Matas insulares)

Capões (Fig. 16 and 17) are small forest islands, not larger than four to five hectares and are characterised by common elements of subtropical broad-leaved evergreen forest but in a lower amount and volume. They are also part of the Planalto. In the majority of cases they are located where water reaches the surface.



Fig. 16: Capão, completely surrounded by grassland



Fig. 17: Interior view of a Capão; trees are trapped by *Tillandsia usneoides* (Bromeliaceae), lower surface of the leaves of *Drymis brasiliensis* (Winteraceae, white arrow)

5. Gallery forest (Mata de Galeria)

Gallery forests (Fig. 18) resemble Capões but unlike them they join rivers and accompany them downstream. The vegetation is relatively open and poor in species.

Lithraea brasiliensis (Anacardiaceae), *Matayba elaeagnoides* (Sapindaceae), *Calyptranthes concinna* and *Siphoneugenia reitzii* (both Myrtaceae), *Casearia decandra* (Flacourtiaceae), *Daphnopsis racemosa* (Thymeleaceae), *Actinostemon concolor* (Euphorbiaceae), *Sorocea bonplandii* (Moraceae) and *Zanthoxylum* (Rutaceae) are typical species of Capões and gallery forests.



Fig. 18: Gallery forest following a river (arrow)

6. Grassland (Campo)

Campo alternates with *Araucaria* forests and Capões in the territory of the Planalto. The grasslands are often riddled with *Sphagnum* moss patches (Fig. 19) particularly in contact with Capões. Prior to post-Columbian settlement more than half of the territory (60% = 161.460 km²) of Rio Grande do Sul was covered by grasslands (FELDENS 1989). Today, the modern grasslands and also the forest biomes are strongly influenced by human activities. The origin of these grasslands could still not be explained sufficiently (HUECK 1952, LÖTSCHERT 1981, RÖNICK 1981, IBGE 1986, GOLTE 1993, MAIER 1998, BEHLING 2002). It has been proved that the grassland vegetation does not correspond to the actual humid and moderate warm climate, which normally would favour more bountiful forests (HUECK 1966, KLEIN 1975, LÖTSCHERT 1981, RÖNICK 1981). Several scientists, e.g. HUECK 1952, RAMBO 1956, LINDMAN 1906, LEITE & KLEIN 1990, suppose that grasslands are relicts from a former drier climate which are joined in by *Araucaria* forest. However, a reconstruction

of the former vegetation is difficult due to serious anthropogenic influences (BEHLING 2002, BEHLING et al. 2001, HUECK 1966, OVINGTON 1983).



Fig. 19: Swamped Campo

Campo is differentiated in two expressions:

6.1 “Clean” grassland (Campo limpo, Campinas, Fig. 20)

The vegetation of the “campo limpo” is highly diverse and is characterized by hemi-cryptophytes and geophytes. Different species of Poaceae, especially the genera of *Andropogon*, *Agrostis*, *Axonopus*, *Eragrostis* and *Paspalum*, Asteraceae (*Achyrocline*, *Baccharis*, *Eupatorium*, *Senecio*), Cyperaceae (*Rhynchospora*), *Desmodium* (Leguminosae), *Relbunium* (Rubiaceae) are dominant (LEITE & KLEIN 1990, RAMBO 1956). Further species of this vegetation type are *Tibouchina gracilis* (Melastomataceae), *Eryngium spp.* (Apiaceae) and *Pteridium aquilinum* (Dennstaedtiaceae) (KLEIN 1978, RAMBO 1956, RÖNICK 1981). The occurrence of this vegetational pattern is associated with shallow soil and with cold winds during winter.



Fig. 20: Campo with *Tibouchina gracilis* (Melastomataceae, magenta blossoms)

6.2 “Dirty” grassland (Campo sujo, Capoeira, Fig. 21)

In contrast with the “campo limpo”, the “campo sujo”, in addition to grasses and sedges, contains irregular dense and high shrubs, as well as small wooden plants. It is characterised by the impressive “Pampass grass” *Cortaderia selloana* (Poaceae), *Blechnum imperiale* (Blechnaceae), *Eryngium sp.* (Apiaceae), *Baccharis uncinella* and *Baccharis trimera* (both Asteraceae). Originally, forests grew on this ground, but were completely cleared in the course of settlement. Capoeiras represent the first stage of succession to the point of the forest stage.



Fig. 21: Capoeira with *Baccharis uncinella* (Asteraceae)

Both types of grasslands include a high number of different herb species.

7. Anthropogenic affected area

Beside a classification in vegetation types, it is necessary to distinguish forest types, depending on the kind and intensity of exogenous factors. They are divided in **primary forests**, **secondary forests** and **exploited forests**. **Primary forests (Mata primária)** are primeval forests or forests subjected to a lower extent of anthropogenic influence. **Secondary forests (Mata secundária**, Fig. 22) include all stages of succession. *Dasiphylum spinescens*, *Dasiphylum tomentosum*, *Vernonia sp.* (Asteraceae) and *Mimosa spp.* (Leguminosae) are indicator plants pertaining to this forest type. Exploited forests are characterised by a selective exploitation of valuable timbers (LAMBRECHT, 1986). Generally, a vast area was transformed to cropland (**Áreas de cultivo**). Beside the cultivation of several economic plants, e.g. *Zea mays* (maize) and *Saccharum officinarum* (sugar cane), large areas of the region are covered by plantations of exotic species, such as

Pinus elliotii, *Pinus taeda* and *Eucalyptus*. However, most parts of the area are used extensively as pasture land.



Fig. 22: *Typical secondary forest on the slopes of the Serra Geral*

2.2 Characterisation of vegetation by palynological analysis

For the characterisation of vegetation, 85 localities were selected at random in the 3 sub-areas of the study region (Fig. 23). At each locality the geographical position (GPS) and the altitude (altimeter) were recorded (App. IV). The vegetation around each sample point was described and a surface-soil sample was collected for the palynological analysis.

Vegetation types	Sub-area		
	1	2	3
Semi-deciduous forest	1	1	5
Deciduous forest	0	0	2
<i>Araucaria</i> forest	3	8	12
Gallery forest	1	2	1
Capão	9	6	4
Capoeira	1	2	2
Grassland	5	9	3
Anthropogenic affected area	1	1	6

Tab. 1: Overview of sample point distribution in the different sub-areas and vegetation types

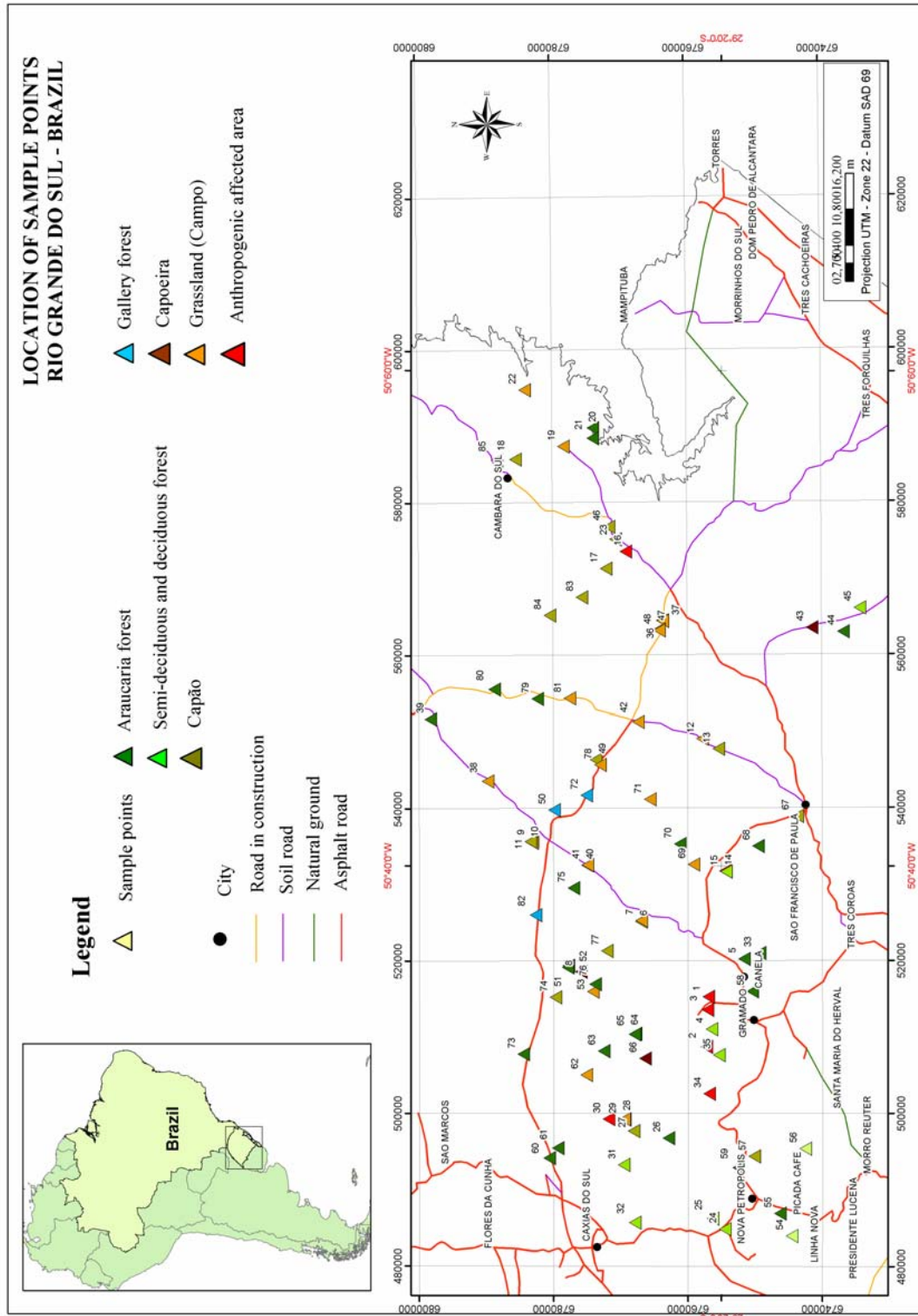


Fig. 23: Locations of sample points – Rio Grande do Sul - Brazil

2.2.1 Sampling of surface-soil samples

The surface-soil samples were collected to extract and analyse the palynomorphs from these soils. Metal samplers measuring 7/ 5.5/ 3.6 cm ($\cong 139 \text{ cm}^3$) (Fig. 24) were used for sampling the surface soil. Before the sampler was driven into the soil with a sledge, the surface of the ground was softly cleaned from leafage, grass-tufts etc. The samples were transported in tagged plastic bags, sealed and kept in a freezer until preparation.



Fig. 24: Soil sampler

2.2.2 Preparation of surface-soil samples

Before the soil samples were prepared by the modified method of KAISER & ASHRAF (1974) and EBNER (2005) they were dried at 40 – 45°C in a drying kiln overnight.

The samples underwent the following steps:

- 1) Crushing of the sample with mortar and pestle.
- 2) **10g** of the dried soil and **2 Lycopodium tablets** were mixed with **100 ml distilled water** in a glass beaker and heated at once.

- 3) **5g KOH lumps** were added to deflocculate and remove humic colloides.
- 4) The reaction was allowed to proceed under heating and stirring for 5 min.
- 5) Stopping of reaction by addition of **40 ml of 36% HCl**.
- 6) Filtration of the solution over a **125 µm sieve** (Fig. 25) with great quantities of water.
- 7) Filtration over a **6 µm filter** (Fig. 26) with great quantities of water.
- 8) Placing of the filter on paper towel for drying.
- 9) Rinsing the dried filter with saturated **ZnCl₂-solution** and transfer of the soil precipitate in a glass beaker.
- 10) The soil solution and the saturated ZnCl₂ were transferred into centrifuge tubes and allowed to spin at **3000 rpm** for **5 min** (heavy liquid separation) without using brakes as to avoid turbulences. The heavy liquid separation is based on a separation of organic material from inorganic material. The saturated solution which has a specific weight higher than that of the palynomorphs but lower than that of the inorganic material. Whereas the specific weight of the palynomorphs is between 1.4 g/cm³ (RT) and 2.1 g/cm³ in RT (depending on the density of the exine) the specific weight of most inorganic materials is over 2.64 g/cm³ in RT (cp. JUVIGNÉ 1973). Saturated ZnCl₂-solution has a specific weight ranging between 1.96 g/cm³ in RT (cp. with FUNKHOUSER & EVITT, 1959) and 2.1 g/cm³ in RT.
- 11) Decantation of the supernatant from the tube over a **6 µm filter** (Fig. 27) and washing with great quantities of water.
- 12) Transfer of the filter in **99.7% CH₃COOH**.
- 13) Transfer of the filter in **Acetolytical solution (H₂SO₄ :(CH₃CO)₂O = 1:10)** and boiling in a water bath for **20 min**.
- 14) Washing of the filter with great quantities of water.
- 15) Preparation of microscopic slides with glycerine gelatine.
- 16) Sealing the slides with nail polish the next day.



Fig. 25: soil sieve, mesh size 125 μ m



Fig. 26: Filtration over 6 μ m filter



Fig. 27: Decantation after centrifugation

Used materials and chemicals:

MATERIALS/CHEMICALS	COMPANY
<i>Lycopodium</i> spore tablets (marker)	Quaternary Sciences University Lund/ Sweden
KAISER's glycerine gelatine	MERCK
KOH (potassium hydroxide)	SYNTH, DINÂMICA
HCl (hydrochloric acid)	VETEC, NUCLEAR
ZnCl ₂ (zinc chloride)	NUCLEAR
CH ₃ COOH (acetic acid)	SYNTH
(CH ₃ CO) ₂ O (acetic anhydrite)	MERCK, NUCLEAR
H ₂ SO ₄ (sulphuric acid)	DINÂMICA
Centrifuge T23	JANETZKI

2.2.3 Palynological analysis

The palynological analysis was carried out with a ZEISS-Standard-LAB-light microscope with a magnification of 400x. As supposed by FIRBAS (1952) 200 palynomorphs were counted, including pollen, spores, fungi and *Lycopodium*-marker. Their morphological identification was based on reference atlases from the states of Amazonas, Rio de Janeiro and Rio Grande do Sul, from Argentina, Barro Colorado Islands as well as from other publications (Para. 5.2). Based on morphological similarity of pollen/ spores, the taxonomic relationship or the similarity of the habitat, 116 pollen/ spore types were distinguished and, assigned in 62 groups (App. II). This assignment facilitates a more concise treatment of the data with the following different analysis methods. The palynomorphs were catalogued by photographs (App. I) for further comparisons.

2.2.4 Data analysis

For comparative analyses of pollen and spore data and of recent vegetation and pollen and spore data, the results of the palynological analysis were evaluated. Before analysing pollen and spore data with the different methods, their composition was scrutinised and some groups were excluded. For a more concise treatment of the data it was agreed upon the following definition: A group of palynomorphs has to exhibit more than 5% of the total amount of palynomorphs per sample. Only these were taken into consideration. As a result 25 groups of palynomorphs (taxa) were considered. Indicator-palynomorphs characterising the different vegetation types were at the focus of attention.

EXCEL was used for the calculation of the percentages of counted palynomorphs for adjacent TILIA-analyses as well as for data plotting in PCA charts.

The software programs TILIA, TILIAGRAPH, TILIAVIEW were used for data plotting in pollen diagrams.

The distribution of the indicator-spores of *Cyatheaceae* and *Dicksonia sellowiana* was investigated with the *raster interpolation method* ArcGIS 8/ArcMap (1999-2002).

CANOCO (Version 4, 1988-98) and PAST (2002) were used for *multivariate statistical analysis*.

Principal components analysis (PCA) was one of the applied *multivariate statistic methods*. It is an ordination method that produces hypothetical variables (components), accounting for as much of the variation in the data as possible. These new variables (components) are linear combinations of the original variables. PCA is a descriptive method (HAMMER 2002). In this thesis the abundance of 25 taxa of pollen and spores in 81 samples was determined. PCA resulted in 25 components from these variables. The program also presents 'loadings'

of the components, illustrating to what extent each original variable contributes to the components.

Cluster analysis is the second utilised method of multivariate analysis. It aims to find groups of samples (or taxa) based on an appropriate distance measure. Hierarchical cluster analysis produces a 'dendrogram' where similar samples are grouped together. Cluster analysis of samples ('Q' mode) is used for detecting associations. Cluster analysis of taxa ('R' mode) is used for detecting of associations of taxa. Several algorithms are available for hierarchical clustering. Generally they cluster the most similar items first, and then proceed by grouping the next most similar cluster until a single is left. Cluster analysis was executed on the sample level to emphasise the results of PCA. In this study the mean linkage algorithm, also known as *Unweighted Pair-Group Moving Average (UPGMA)*, was used for hierarchical clustering. This algorithm was combined with *MORISITA's similarity index* (KREBS 1989). 80 samples of a total of 85 were clustered. In four samples the palynomorphs were under-represented and one sample (sample No. 55) yielded a misinterpretation. They were therefore not considered in the data analysis. For obtaining more detailed information, a cluster analysis was executed separately for the forest samples, the grassland samples and samples with intense interaction with grassland (samples of Capões, Capoeira, gallery forests).

2.3 Mapping of the vegetation

For comparison of the relationship between the pollen/ spore data and the vegetation, the recent vegetation around each sample point was identified and recorded. Therefore, the vegetation around the points was identified in a 10m x 10m square by a simplified BRAUN – BLANQUET-technique. The application of this method to forests in subtropical and tropical regions is intricate because the identification of high trees is difficult to ascertain.

Therefore, it is emphasised that this work does not claim completeness of the vegetation record. The mapping of vegetation was merely done to get a first impression about the local vegetation.

For the mapping of vegetation the following keys for dominance and species abundance were used:

Dominance:

scale	covering in %
5	>75 - 100
4	50 - 75
3	25 - 50
2	5 - 25
1	< 5

Species abundances:

r (raro) sporadic, mostly one specimen only
p (paululum) few specimens
a (amplius) numerous specimens
m (multum) multiple specimens

The vegetation was identified on site and specimens of unknown plants were collected for their further identification in the laboratory. The collected plant specimens were identified in the herbaria of LPB/PUCRS and UFRGS, generously supported by Professors J. A. JARENKOW (UFRGS), M. SOBRAL (UFMG) and N. I. MATZENBACHER (PUCRS). In addition, several references were used for the identification (Para. 5.3)

For concise comparison between the different vegetation types and the sample sites, the most frequent taxa were further analysed. Therefore the BRAUN-BLANQUET-abundance values were substituted by numbers as follows: $r = 1,1$; $p = 1,2$; $a = 1,3$; $m = 1,4$. For each selected taxa (Para. 3.5, Tab. 4) the mean value of dominance and abundance were selected independent from the different tree or shrub layers. Then, by multiplying of these

abundance values with the dominance values, a new factor (AD-value = abundance-dominance-value) was calculated. One example should explain the procedure: The family of Myrtaceae occurs in sample No. 5 (*Araucaria* forest) in the first tree layer with the BRAUN-BLANQUET-value of 2p in the second tree layer of 2a. For further calculations the value 2a was used and converted in 2,6. Finally, the average of the AD-values of the selected taxa (Tab. 4) for the different vegetation types in the different sub-areas was calculated.

3. Results

3.1 Pollen diagrams - distribution of pollen and spores in the different vegetation types

The pollen diagrams represent the relative percentage variation of pollen and spore types found in the according vegetation type. Each of the presented pollen/ spore type has at least relative percentages of 5% in one sample. Top down it moves from taxa frequency in sub-area 1 to those in sub-area 2 and 3.

3.1.1 Distribution of pollen and spores in semi-deciduous forests

In semi-deciduous forests (samples No. 4, 15, 24, 25, 31, 32 and 45) the phylum of pteridophytes was the most frequent group (60%, Fig. 28). Within the pteridophytes the *Blechnum* group, the *Microgramma* group, the *Asplenium* group and the *Cyatheaceae* group were predominant. It has to be emphasised that the same spores of the *Blechnum* and *Microgramma* group appeared both, in the forests on the south-eastern slopes of the Serra Geral (sample No. 45) and in the valley of the river Caí (samples No. 24, 25). The striking presence of the *Asplenium* group, mainly in the semi-deciduous forests in the valley of the river Caí (sample No. 24, about 3% and sample No. 25, about 10%) and on the Western slopes of Serra Geral (sample No. 31, more than 20%); is worth a careful observation. The *Asplenium* group was scarce in the sites on the southern (samples No. 4, 15) and south-eastern slopes of the Serra Geral (sample No. 45). In addition, sample No. 31 aroused attention by the occurrence of spores of the *Dennstaedtia/ Pteridium*-group and the *Anogramma/ Pteris*-group. The spores of *Dennstaedtia/ Pteridium*-group amount to more than 10% of all counted palynomorphs. The *Cyatheaceae* group was most abundant in the total amount of Pteridophyta spores. In sample No. 15 it reached over 40% of the total amount of pollen and spores.

The spores of *Cyatheaceae* and conditionally of *Asplenium* are characteristic marker of semi-deciduous forest. Pollen grains of gymnosperms and angiosperms were generally

under-represented in semi-deciduous forests with exception of *Ilex* pollen (more than 10%). *Ilex* pollen was found to occur solely in sample No. 15 and 45.

3.1.2 Distribution of pollen and spores in deciduous forests

The semi-deciduous forests and deciduous forests were intricate to distinguish by palynological analysis. The combined illustration (Fig. 28) ought to emphasise this complexity of separation of these two forest types.

As shown for semi-deciduous forests, the phylum of pteridophytes was the most frequent group in deciduous forests. However, fern spores did not appear in the same quantity in deciduous forests as they did in semi-deciduous forests. The spores of the members of the *Microgramma* group, and even *Microgramma* itself (Tab. 2), were rare. In sample No. 56 they even lacked. In contrast to the semi-deciduous forests, the *Asplenium* group appeared to be rare in deciduous forests in this investigation.

Pollen grains of gymnosperms and angiosperms were also under-represented in deciduous forests with exception of Euphorbiaceae pollen (Fig. 28). Pollen grains of Euphorbiaceae appeared in samples No. 54 and 56 and were more frequent in sample No. 54 with almost 7%.

Sub-area	Sample points	Blechnum group		Asplenium group			Dennstaedtia/ Pteridium group	Cyatheaaceae group				Euphorbiaceae group	
		<i>Blechnum</i>	<i>Microgramma</i>	<i>Asplenium 1</i>	<i>Asplenium 2</i>	<i>Dryopteris</i>		<i>Dennstaedtia</i>	<i>Alsophila</i>	<i>Cyathea schaschkin</i>	<i>Cyathea spp.</i>	<i>Nephelea</i>	<i>Alchornea trinlinerva</i>
1	45	0.2	1.7	1.2	0.2	0.5	0.2	0.0	0.2	0.0	0.0	0.0	0.0
2	15	8.4	9.3	0.0	0.0	0.0	0.0	15.2	12.7	2.1	10.5	0.0	0.0
3	4	1.7	0.4	0.0	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3	24	0.7	2.0	0.0	0.0	3.2	0.0	0.5	0.5	0.0	0.0	0.0	0.0
3	25	0.5	1.4	0.0	0.0	10.3	0.0	2.6	0.0	0.0	0.0	0.0	0.0
3	31	1.8	0.4	3.5	12.7	9.9	14.1	0.4	0.0	1.4	0.0	0.4	0.0
3	32	10.0	6.1	0.0	0.0	0.0	0.0	14.3	8.2	8.2	5.4	0.0	0.0
3	54	3.8	3.1	0.0	0.0	1.5	0.0	0.0	0.0	0.0	0.0	2.3	4.6
3	56	0.4	0.0	0.0	0.0	1.8	0.0	0.4	0.0	0.0	0.0	0.0	0.0

Tab. 2: Relative percentage of important pollen and spore types within selected groups in semi-deciduous and deciduous forests (conspicuous values are accented)

3.1.3 Distribution of pollen and spores in *Araucaria* forests

In *Araucaria* forests, like in semi-deciduous and deciduous forests, pteridophytes were among the most frequent group. The palynomorphs in *Araucaria* forests were characterised by the *Blechnum* (almost 30%) and the *Microgramma* group (almost 40%) (Fig. 29). Interestingly enough, it is observed that *Vittaria* occurred mainly in *Araucaria* forests situated on the slopes of Serra Geral (sample No. 44, almost 2%) or on slopes alongside rivers (samples Nos. 5, 33, maximum about 3 %) (Tab. 3). The increased amount of *Hymenophyllum* (more than 3% in sample No. 79) and *Trichomanes* spores in *Araucaria* forests was remarkable compared to the semi-deciduous and deciduous forests. The high percentage of tree fern spores is worth being paid attention to. *Dicksonia sellowiana* reached an amount of almost 44% (sample No. 44). Depending on the location, the tree ferns of Cyatheaceae unfold a peculiar significance (Para. 3.2).

Pollen grains of *Araucaria angustifolia* and *Podocarpus lambertii* occurred but rather occasionally (Fig. 29). They were clearly evident in samples located in the transition zone of *Araucaria* forest with the broad-leaved evergreen forest (about 8%) and on the Planalto (over 6% in sample Nos. 64 and 79 or rather more than 18% in sample No. 61).

Pollen of angiosperms, in particular Poaceae (almost 30% in sample Nos. 73 and 79), Asteraceae (over 10% in sample Nos. 73 and 76), Euphorbiaceae, Myrtaceae, Meliaceae and Sapindaceae, as well as the genus *Ilex* occurred more often than in semi-deciduous and deciduous forests. However, they do not exclusively represent the *Araucaria* forests which are located on the Planalto as they appeared also in *Araucaria* forests which are conspicuously influenced by subtropical elements. Sample No. 55 is eminently mentioned because in this sample Myrtaceae pollen grains were represented with almost 25%, Meliaceae pollen with about 17% and Sapindaceae pollen were detected with a percentage of about 15%. The pollen assemblages Nos. 65, 68, 70, 73, 75 and 76 generated an unusual low amount of pollen grains and were represented rather exclusively by Asteraceae, therefore they did not represent a typical pollen spectrum of *Araucaria* forest.

Sub-area	Sample points	Blechnum group					Microgramma group		Pteridophyta 2	Euphorbiaceae group		
		<i>Blechnum</i>	<i>Dicranopteris</i>	<i>Gleichenia</i>	<i>Pteridophyta 1</i>	<i>Vittaria</i>	<i>Microgramma</i>	<i>Davallia</i>		<i>Actinostemon concolor</i>	<i>Alchornea triplinervia</i>	<i>Sebastiania commersoniana</i>
1	20	1.1	0.0	0.0	3.5	0.0	0.9	0.0	0.2	0.2	0.9	0.7
1	21	9.1	3.8	3.2	1.6	0.0	8.6	3.2	0.0	0.0	0.0	1.1
1	44	5.7	2.3	3.1	10.0	1.9	7.3	3.8	0.0	0.0	0.4	1.5
2	5	2.4	0.0	0.0	13.3	2.4	0.3	0.0	0.0	0.0	0.3	1.0
2	33	8.4	1.3	0.9	0.0	2.7	24.8	4.4	0.0	0.0	0.0	1.3
2	39	4.9	4.9	7.7	8.1	0.4	10.2	2.4	0.0	0.0	0.0	1.2
2	68	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2	70	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2	75	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2	79	0.5	0.0	0.8	0.5	0.0	1.8	0.5	0.0	0.0	0.5	0.0
2	80	1.2	0.0	1.2	2.8	0.0	4.8	0.8	0.0	0.0	1.6	1.6
3	26	0.6	0.0	1.9	1.2	0.0	9.0	0.9	0.0	0.0	0.0	0.0
3	52	5.0	7.0	4.0	5.4	0.0	10.7	2.3	0.3	0.3	0.0	1.7
3	55	0.9	0.0	0.4	0.0	0.4	2.2	0.0	0.0	0.0	0.0	0.4
3	58	0.5	0.0	0.0	0.0	0.0	1.0	0.0	0.0	0.0	0.0	0.0
3	59	7.5	2.9	2.2	11.0	0.4	24.1	3.5	0.0	0.0	0.4	3.1
3	60	0.9	2.7	1.7	3.0	0.4	11.6	2.6	0.0	0.0	0.0	0.0
3	61	1.6	2.1	0.5	1.6	0.0	18.1	0.0	1.6	1.6	0.0	0.0
3	63	5.6	0.0	0.0	3.1	0.0	1.5	0.0	3.1	3.1	21.9	4.8
3	64	2.2	2.2	1.3	1.3	0.0	8.7	1.3	0.0	0.0	0.9	1.3
3	65	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3	73	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3	76	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0

Tab. 3: Relative percentage of important pollen and spore types within selected groups in *Araucaria* forests (conspicuous values are accented)

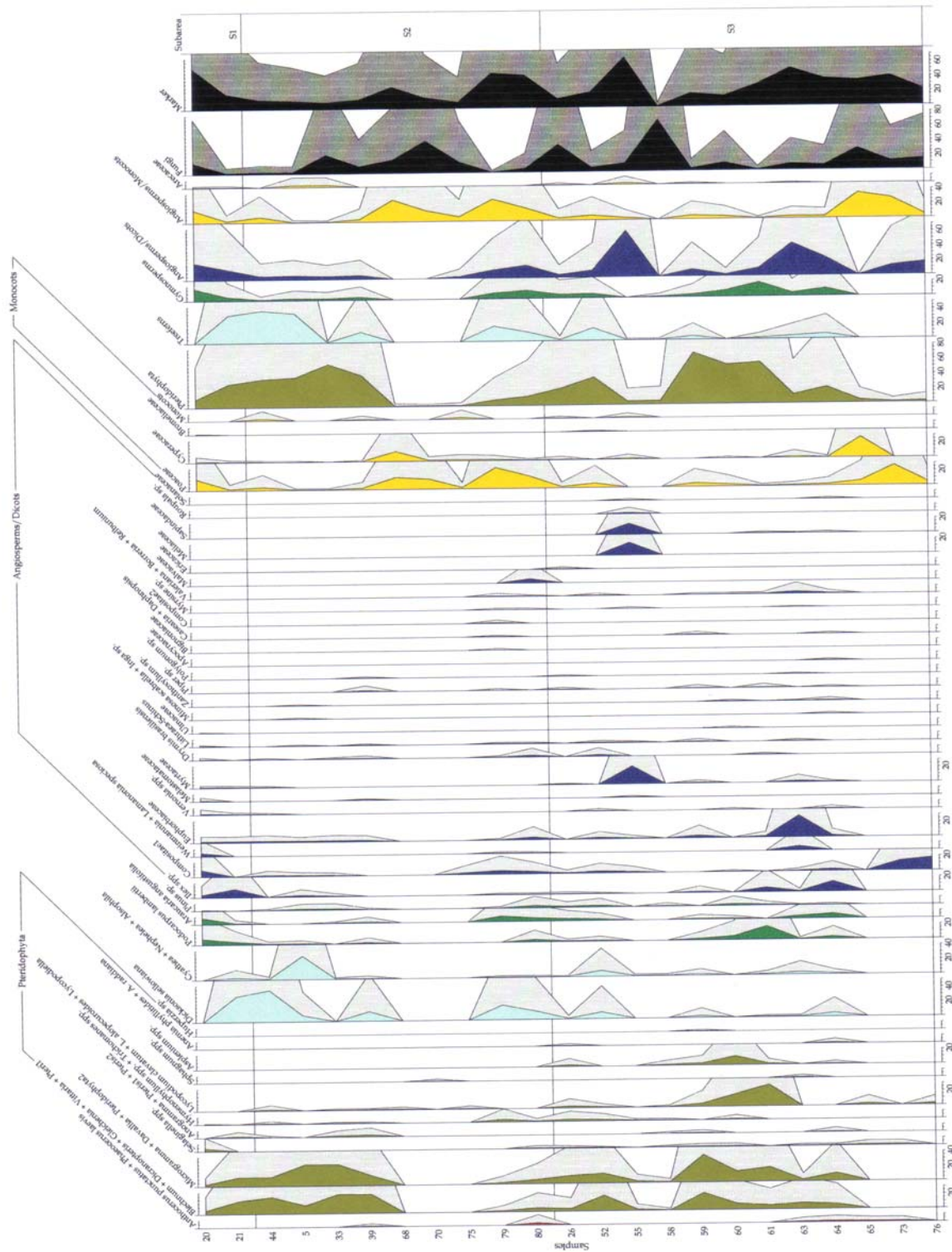


Fig. 29: Distribution of pollen and spores in Araucaria forests (%)

3.1.4 Distribution of pollen and spores in Capões

Capões are comparable to *Araucaria* forests with respect to the *Blechnum* and *Microgramma* group which occurred most frequently. Tree ferns were less abundant in Capões (about 8%) than in *Araucaria* forests (Fig. 30). The percentage of *Dicksonia sellowiana* spores was slightly higher (7.6%) than Cyatheaceae spores (6.6%).

Pollen grains of the gymnosperms *Araucaria angustifolia* and *Podocarpus lambertii* occurred more constantly in Capões than in *Araucaria* forest, with *Podocarpus lambertii* reaching a high pollen prevalence of almost 75% (sample No. 13, Fig. 30) or almost 30% (sample No. 36). However, *Podocarpus lambertii* was almost inexistent in Capões situated on flat declivities of the Western Serra Geral (samples No. 27, 28 and 57).

In Capões, dicotyledons were found to be less abundant than in *Araucaria* forests but the monocotyledons, particularly Poaceae (over 40%) and Cyperaceae (almost 15%), were more abundant in Capões compared to *Araucaria* forests. *Ilex* pollen (11%), pollen of Asteraceae (about 8%) and Euphorbiaceae (up to 8%), and *Drymis brasiliensis* pollen (nearly 5%) featured a distinct abundance (Fig. 30). While *Drymis brasiliensis* pollen grains appeared sporadically in *Araucaria* forests, they appeared more frequently in Capões. Their rare appearance in Capões situated on flat declivities of the Western Serra Geral (samples No. 27, 28, 57) is worth being mentioned.

In general terms, Capões are characterised by pollen of *Podocarpus lambertii*, Poaceae, *Ilex* sp., Euphorbiaceae and *Drymis brasiliensis*.

3.1.5 Distribution of pollen and spores in gallery forests

With regard to the composition of pollen and spores, gallery forests did not provide the clear pattern of an *Araucaria* forest or Capão (Fig. 31). The *Microgramma* group was specifying the pollen spectrum with a total amount of almost 21% (sample No. 82). Other spores were rarely present.

Pollen of gymnosperms and angiosperms were also rare. Only *Podocarpus lambertii*, *Ilex* and Asteraceae, Euphorbiaceae and Poaceae occasionally presented notable pollen abundance (3-8%).

With regard to gallery forests, bryophytes obtained a higher quantity (about 5%) than in the semi-deciduous and deciduous forests and *Araucaria* forests (Fig. 31).

The characterisation of gallery forests by palynological spectra is less distinct. They are also characterised by *Podocarpus sp.*, Poaceae, Euphorbiaceae and *Ilex sp.* but in lower amounts than Capões.

3.1.6 Distribution of pollen and spores in Capoeira vegetation

Like in forests, pteridophytes were also abundant in Capoeiras, but the importance is no longer related to the *Microgramma* and *Blechnum* groups (Fig. 32). Spores of *Hymenophyllum*, *Trichomanes*, *Lycopodium* and *Lycopodiella* become more important. The tree ferns (mainly Cyatheaceae) are notably present. In sample No. 43 spores of Cyatheaceae are notable present (37%).

Records of Capoeiras mainly contained pollen of Poaceae (more than 18% in sample No.51) and of the *Compositae* group (over 21% in sample No. 8) including pollen of Asteraceae such as *Achyrocline*, *Artemisia*, *Baccharis*, *Calea*, *Eupatorium*, *Senecio* and *Trixis* which mainly grow on grassland.

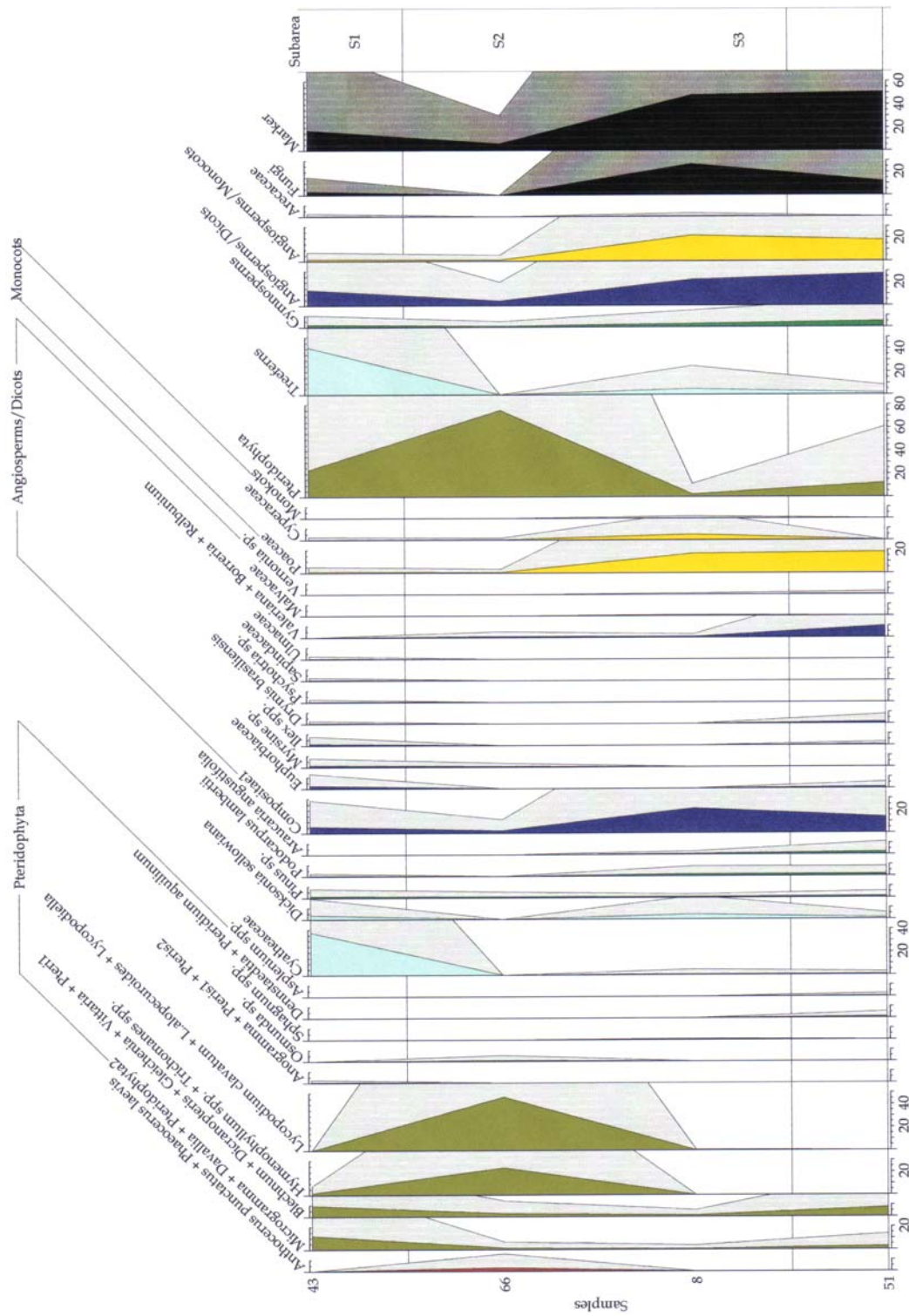


Fig. 32: Distribution of pollen and spores in Capoeira vegetation (%)

3.1.7 Distribution of pollen and spores in Campo vegetation

The pollen of Poaceae and Cyperaceae dominated in grassland samples, with about 50% and 30% respectively, of the total sum of palynomorphs (Fig. 33).

The bryophytes *Anthoceros punctatus* and *Phaeoceros laevis* (about 50% in sample No. 71 and over 25% in sample No. 42) and the peat moss *Sphagnum* (over 2%) reached a conspicuous relevance.

Pollen grains of angiosperms were rare. The group of *Compositae I* amounted to more than 9% (sample No. 22). However, this was lower than in Capoeira vegetation. Pollen and spores from the surrounding vegetation, such as *Dicksonia sellowiana*, *Araucaria angustifolia* or *Podocarpus lambertii* occurred scarcely (about 1-4%).

3.1.8 Distribution of pollen and spores in anthropogenic affected area

There were noticeable amounts of pollen of *Araucaria angustifolia* (maximum 6.5% in sample No. 14, Fig. 34), *Podocarpus lambertii* (maximum 16% in sample No. 1) and of Myrtaceae (almost 9% in sample No. 1) and spores of *Dicksonia sellowiana* (about 7% in sample No. 14). The pollen diagram illustrates palynomorphs, e.g. spores of the *Microgramma* (about 13% in sample No. 30) and *Blechnum* group (about 14% in sample No. 30) as well as *Vernonia* (about 2% in sample No. 14) and *Ilex* pollen (about 1% in sample No. 14), which are representative for forest vegetation (Fig. 34). In addition, typical grassland indicator pollen and spores also occurred. They were represented by pollen of the Compositae1 group, spores of *Anthoceros punctatus* and *Phaeoceros laevis* (1 – 4%).

3.2 Distribution of spores of *Cyatheaceae* and *Dicksonia sellowiana* in the study area (ArcGIS analysis)

For the distribution of spores of *Cyatheaceae* and *Dicksonia sellowiana*, the following tendency was observed: spores of *Dicksonia sellowiana* mainly occurred on the Planalto (Fig. 35). As represented in samples No. 5, 20, 21, 39, 49 and 79, the spores occurred exclusively in *Araucaria* forests, Capões and in grasslands located close to *Araucaria* forests or Capões (samples No. 19, 22 and 78). These spores occurred even in secondary forest (sample No. 14). Occasionally spores of *Dicksonia sellowiana* appeared also in semi-deciduous forests (samples Nos. 15, 33). The amount of spores of *Dicksonia sellowiana* decreased from the east westwards concerning the *Araucaria* forest, Capões and in grassland (Fig. 35). Whereas in the eastern part of the study region the amount of spores of *Dicksonia sellowiana* reached amounts of 34.9% (sample No.21) and 43.3% (sample No.44), they lacked in the western part of the study region.

In contrast to *Dicksonia sellowiana*, spores of *Cyatheaceae* occurred rarely on the Planalto. They were primarily found in forests on the Western and Southern slopes of the Serra Geral (samples No. 15, 32 and 43) and in river valleys (sample No. 5) (Fig. 36). They prevail in high percentages in these forests (40.5% in sample No.15, 36.2% in sample No.32 and 32% in sample No.5). In addition, these spores also occurred on the Eastern margin of the Planalto (sample No.19, 7.6%). They appeared most remarkably in soils of semi-deciduous and deciduous forests and of *Araucaria* forests situated on rivers.

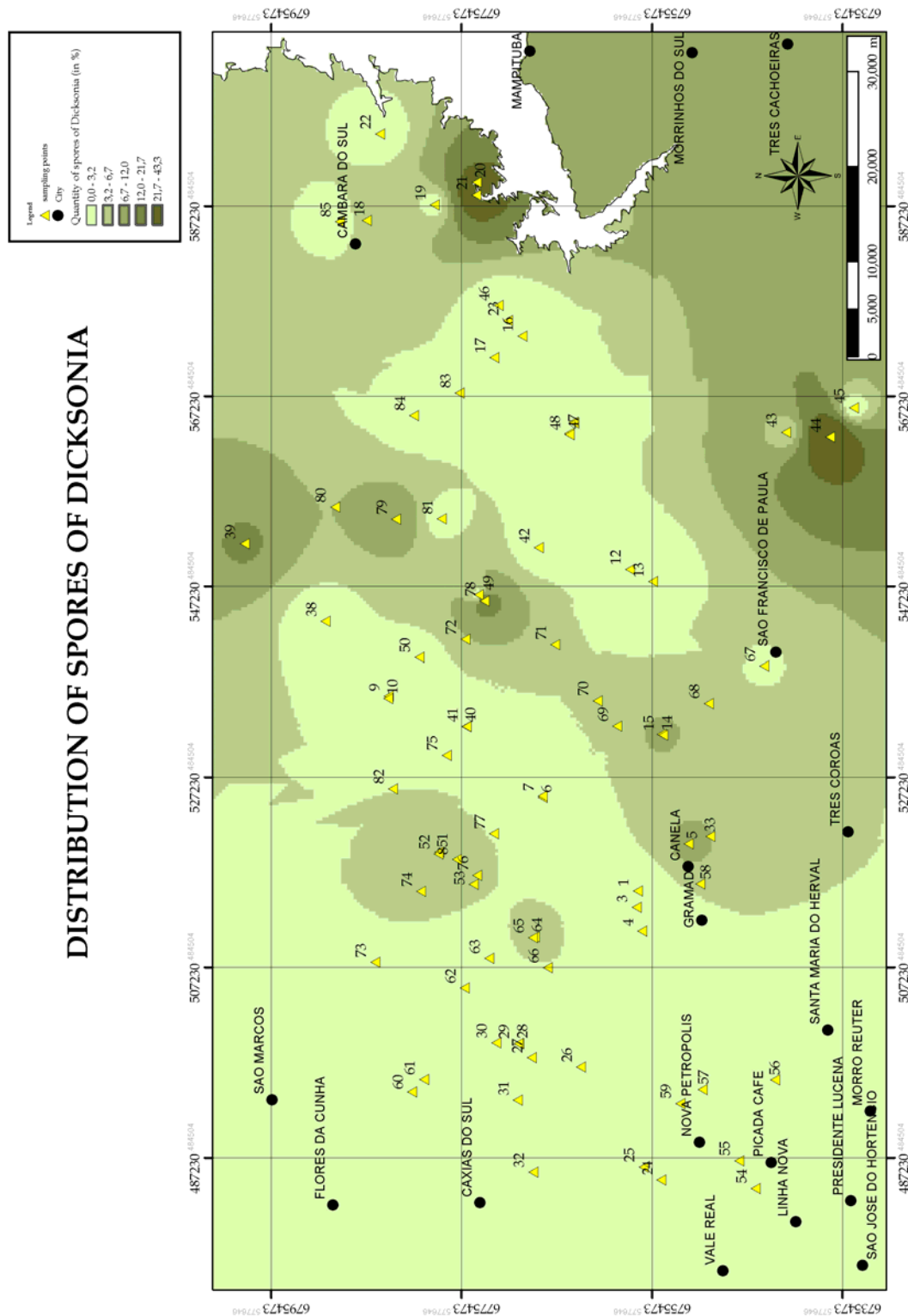


Fig. 35: *Distribution of spores of Dicksonia sellowiana*

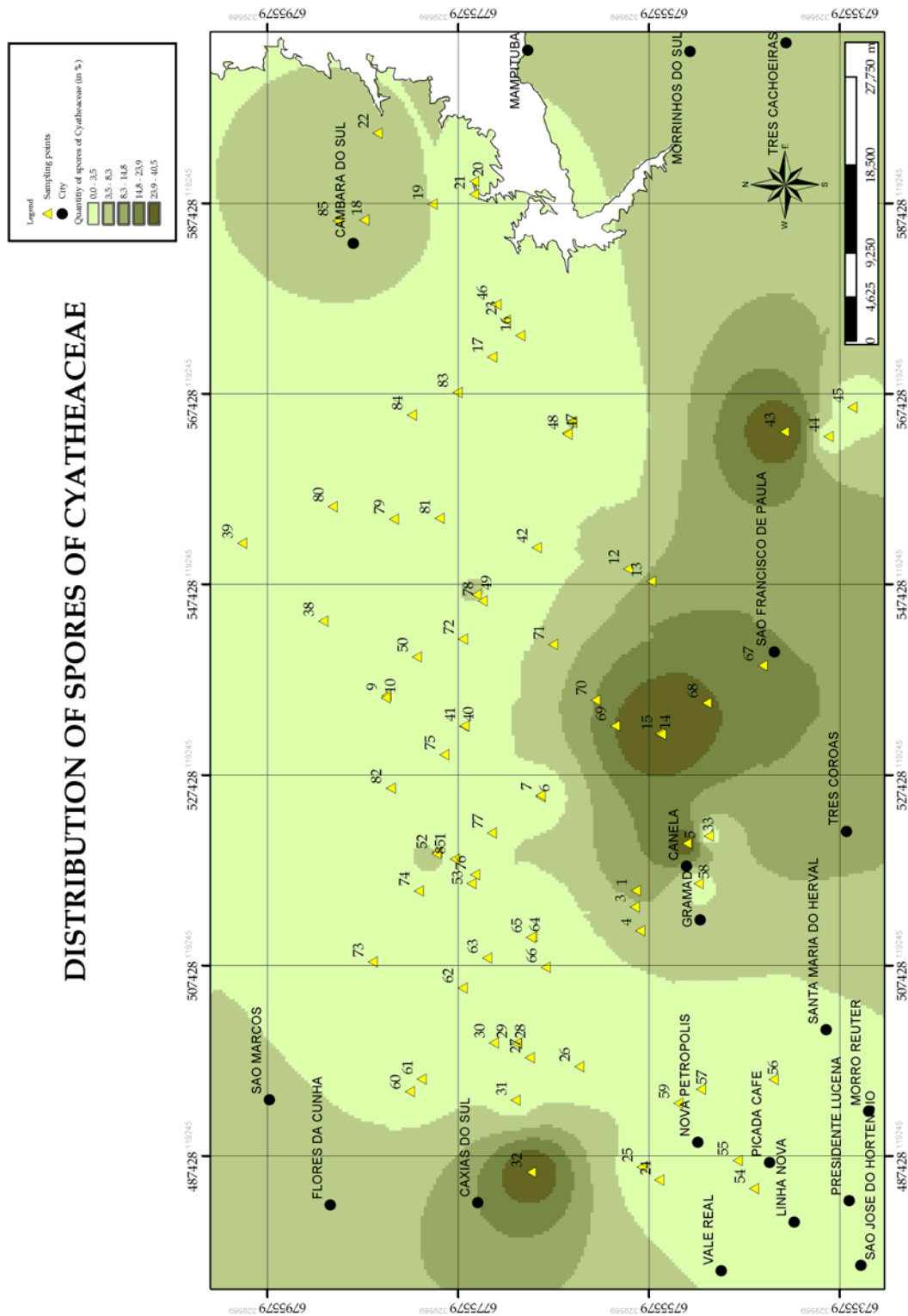
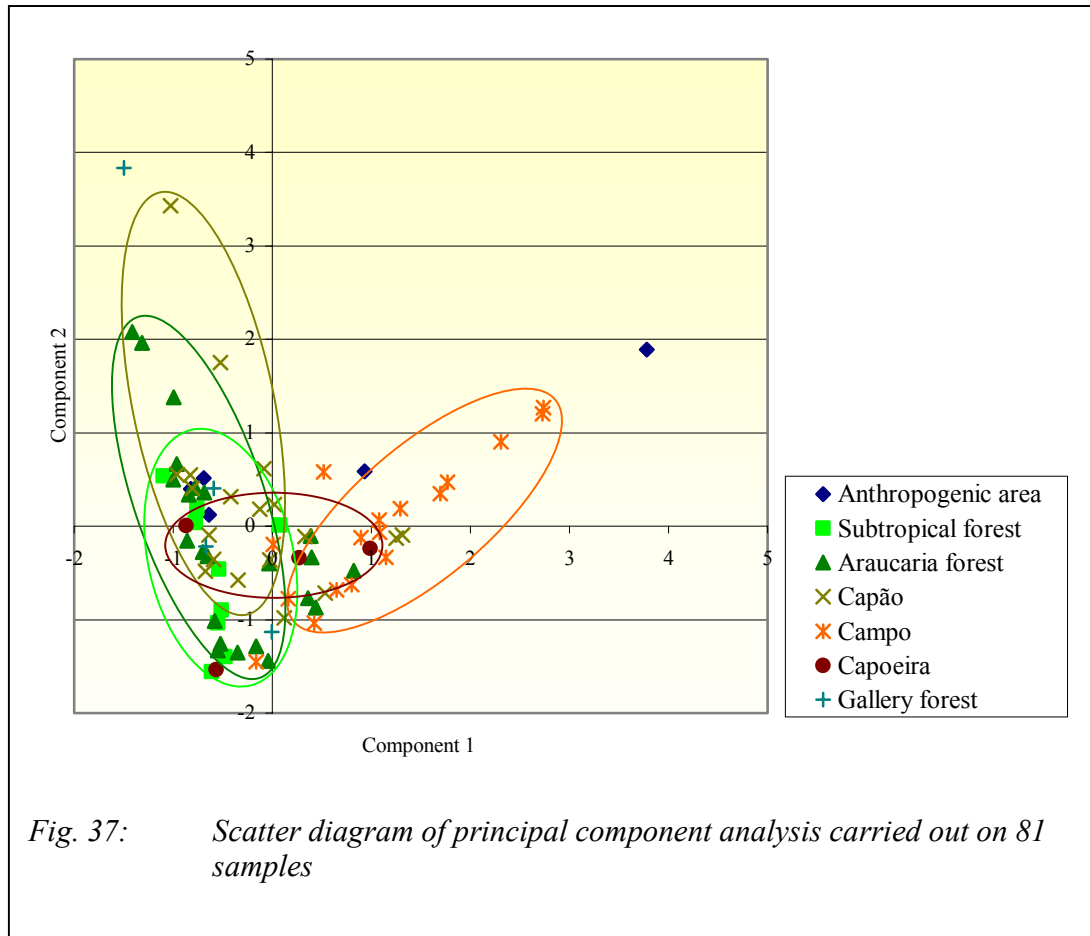


Fig. 36: *Distribution of spores of Cyatheaaceae*

3.3 Principal components analysis (PCA)

3.3.1 Dispersion of samples

PCA demonstrates that a separation of different vegetation types by dint of the palynological analysis is feasible. It results in a distinct separation of grassland vegetation and forest vegetation. Based on 25 variables (pollen and spore types), the PCA analysis produced 25 components (axes). Four of these components were responsible for the variation in the samples of the different vegetation types. The results of PCA the first component explains 40.2% of the data variation, the second explains 12.9 %, the third 9.7%, while the fourth explains 7.7%. Whereas the positive end of the first axis groups primarily grassland vegetation, the negative end of the first axis represents primarily forest vegetation. The results of the PCA can be interpreted in terms of environmental variables. The first axis represents the humidity gradient. Humidity increases from the positive to the negative end of the first axis, and separates clearly the grassland from the forests. The samples corresponding to transitional vegetation (Capoeira, Capões and gallery forests) are in the middle of the scatter indicating that they cannot be distinguished distinctly as separate groups. The second axis is interpreted possibly as the elevation gradient corresponding to the different forest types and Capoeira vegetation. From bottom to top, the second axis represents vegetation types from the slopes of the Serra Geral to the Planalto. Capões represent a binding tie between grassland vegetation and forest vegetation with higher tendency to forest vegetation. The assemblages of Capoeira vegetation displayed a diffuse distribution pattern. They were located along the axis of grassland vegetation as well as at the axis of forest vegetation. Assemblages of the anthropogenic influenced area showed the same distribution pattern.



3.3.2 Dispersion of taxa

The arrangement of components on the level of pollen and spores types demonstrates principally the same pattern as shown above on the sample level. Taxa, such as Poaceae, Cyperaceae, herbaceous members of Asteraceae and Rubiaceae, which are characteristic for grassland vegetation, are arranged along the first axis (Fig. 38, brown ellipse). Taxa which belong to the forest vegetation, e.g. *Araucaria*, *Podocarpus*, *Ilex*, *Dicksonia*, *Microgramma* and *Blechnum*, were mainly clustered on the second axis (Fig. 38, dark green ellipse).

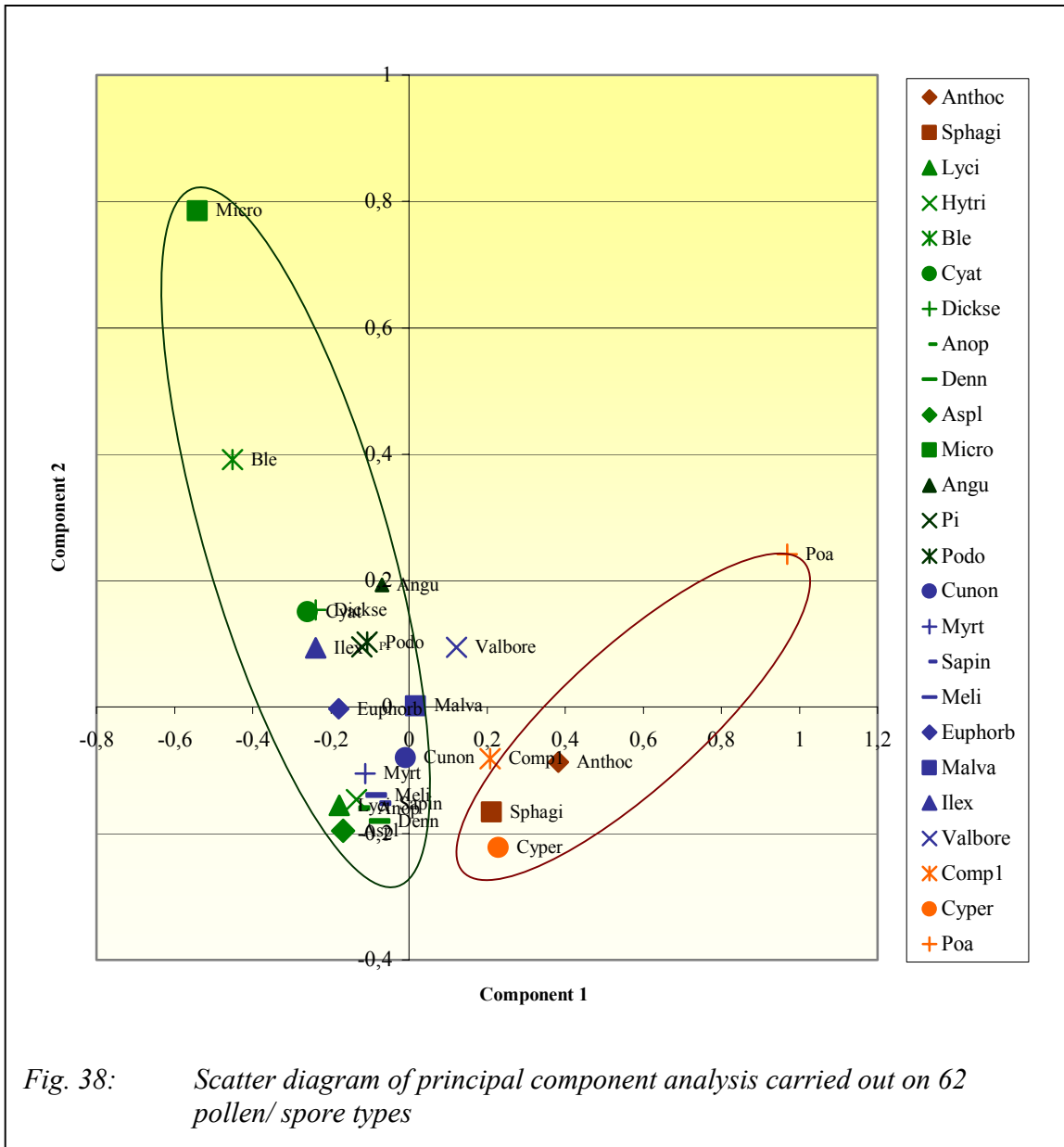


Fig. 38: Scatter diagram of principal component analysis carried out on 62 pollen/ spore types

3.3.3 Loadings of the components

The first component (axis) is defined by taxa of grassland vegetation because they show a positive value (Fig. 39). Poaceae, which contribute with 97%, is the determining group whereas other taxa contribute 38% (e.g. Anthocerataceae) to the first component. Further taxa contributing grassland vegetation are Cyperaceae, Asteraceae (Comp1) and the Sphagnum group. Taxa of the pteridophytes, such as *Microgramma* (76%) and *Blechnum* (39%) define the second component (Fig. 40). The measured contribution of Poaceae (24%) is not to be scoffed.

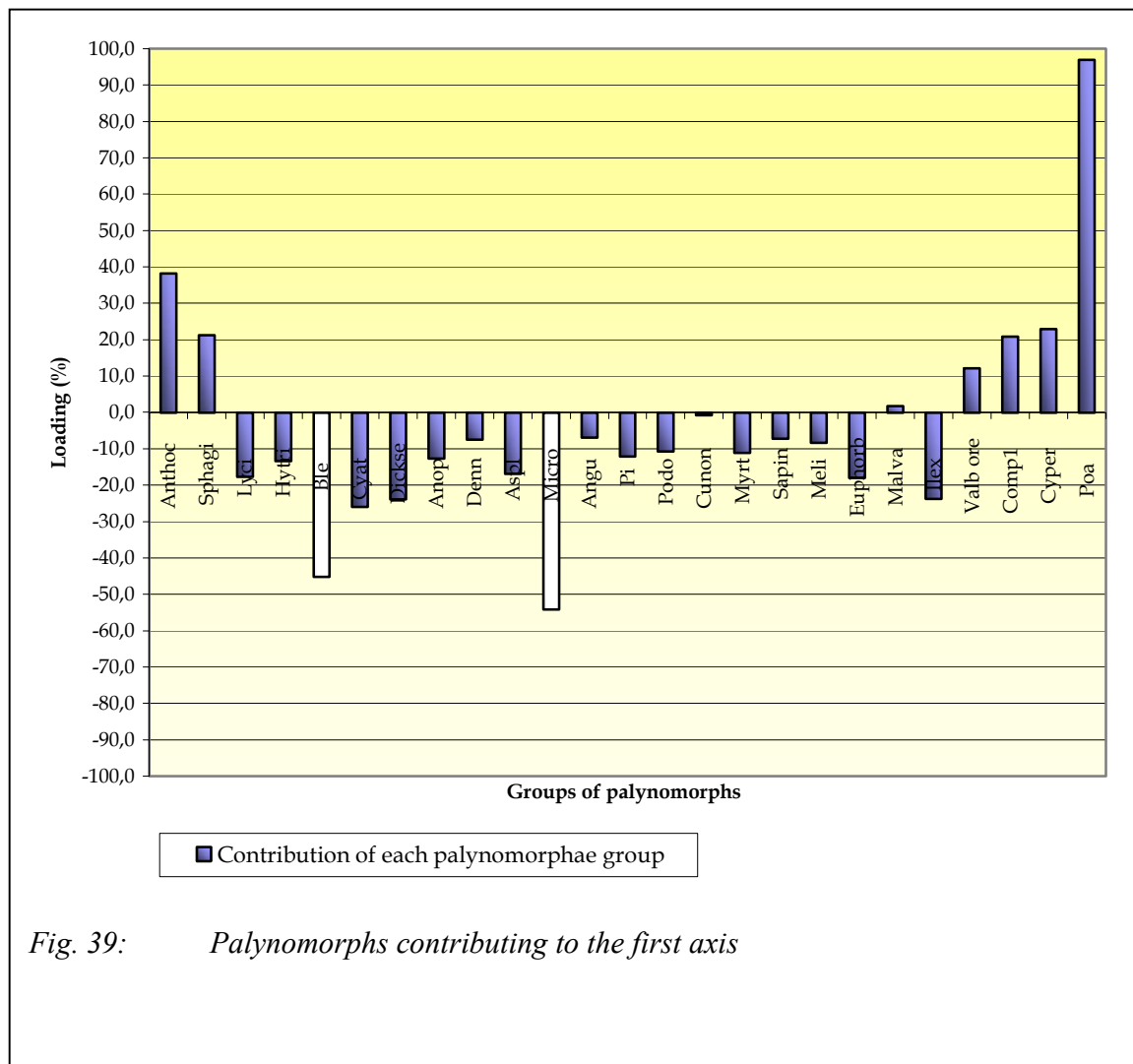


Fig. 39: Palynomorphs contributing to the first axis

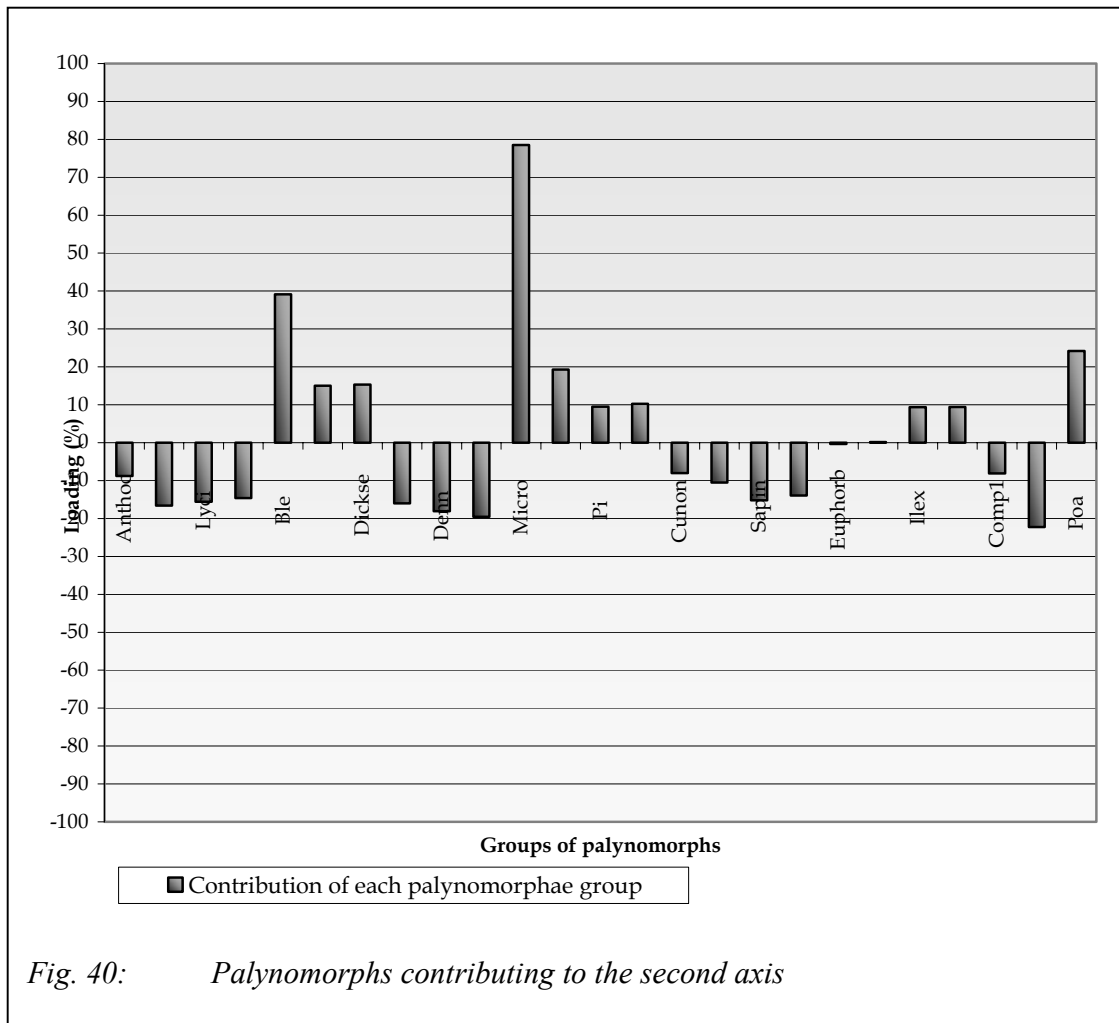


Fig. 40: Palynomorphs contributing to the second axis

3.4 Cluster analysis

3.4.1 Cluster analysis for all samples

Two main clusters are distinguished: The separation is located between sample No. 21 and 31 (Fig. 41, red arrow). The first cluster is characterised by the different forest types (*Araucaria* forest (dark green numbers), semi-deciduous and deciduous forest (magenta numbers), gallery forest (blue numbers) and Capão (dark yellow numbers) as well as by some assemblages from Capoeira vegetation (brown numbers) and assemblages from anthropogenic affected areas (red numbers). The second cluster implies all grassland samples (light blue numbers) and isolated samples of the different forest types, Capoeira and anthropogenic affected areas. In the first cluster the different forest types intermix. A clear separation between the different forest types is not possible. The second cluster shows, in matters of separation, a similar pattern. It presents the grassland cluster spotted by forest vegetation. While the second cluster ('grassland cluster') includes also forest samples, the first cluster ('forest cluster') includes exclusively samples of forest vegetation and transitional vegetation.

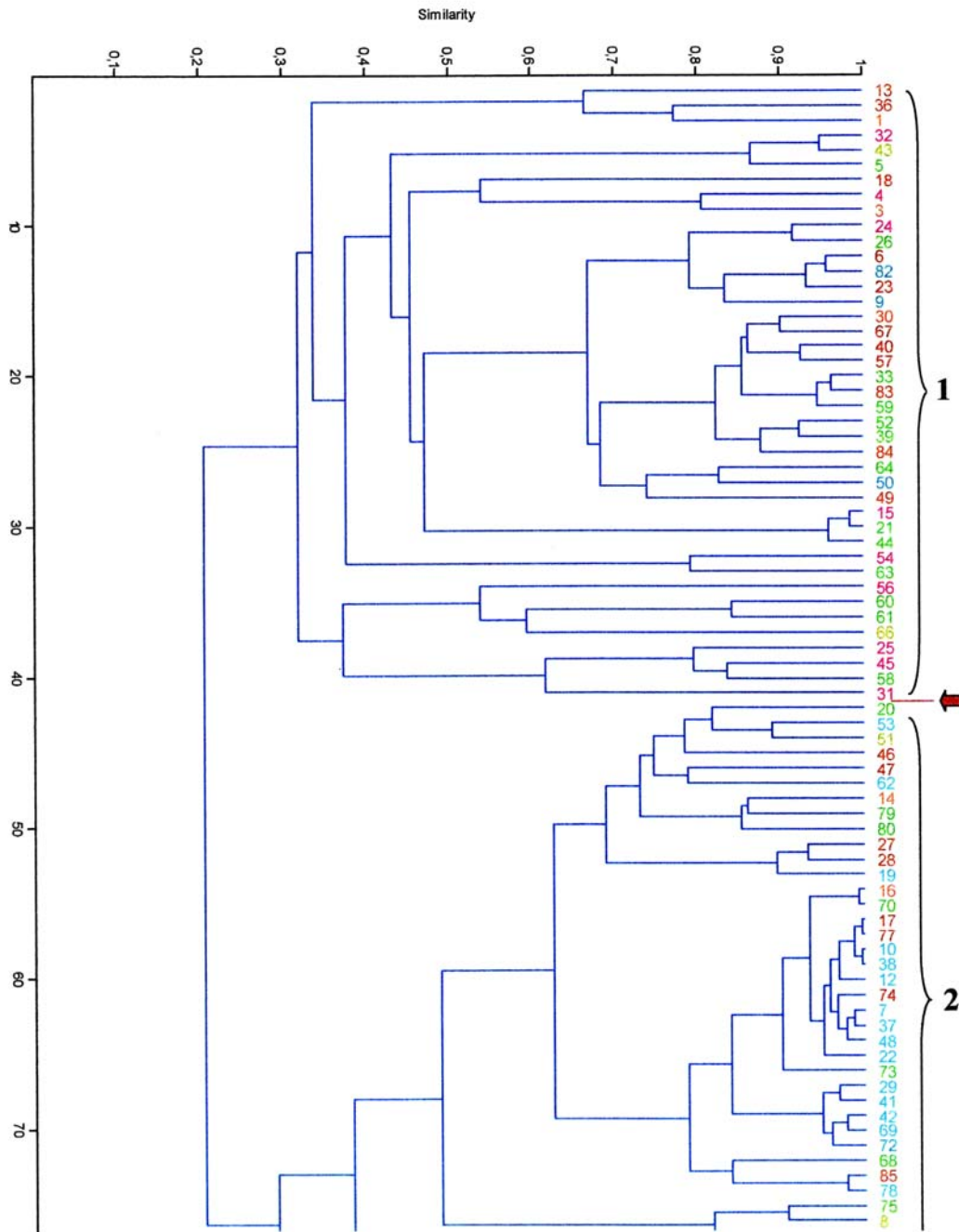


Fig. 41: Cluster analysis – clustering of all samples, using the algorithm of UPGMA and MORISITA's index

3.4.2 Cluster analysis for samples of forest vegetation

The dendrogram illustrates a splitting up of samples in two main branches (branch 1 and branch 2) but within these branches there is no order identifiable (Fig. 42). The branch 2 aggregates mainly samples of Capões collected in sub-area 1, whereas the first branch aggregates samples of Capões of all sub-areas. The other forest types point out similar results: Samples of semi-deciduous and deciduous forest sites showed a diffuse pattern.

3.4.3 Cluster analysis for samples of Campo vegetation and samples with intense interaction with Campo vegetation

The dendrogram of this cluster analysis presented a splitting up of the samples in two main branches (Fig. 43). In contrast to the previous dendrogram, the first main branch clusters all samples of Campos. Samples of Capão-, Capoeira and gallery forest vegetation appear in branch 1 and branch 2 independent of the origin of the sample. A clear order is not identifiable.

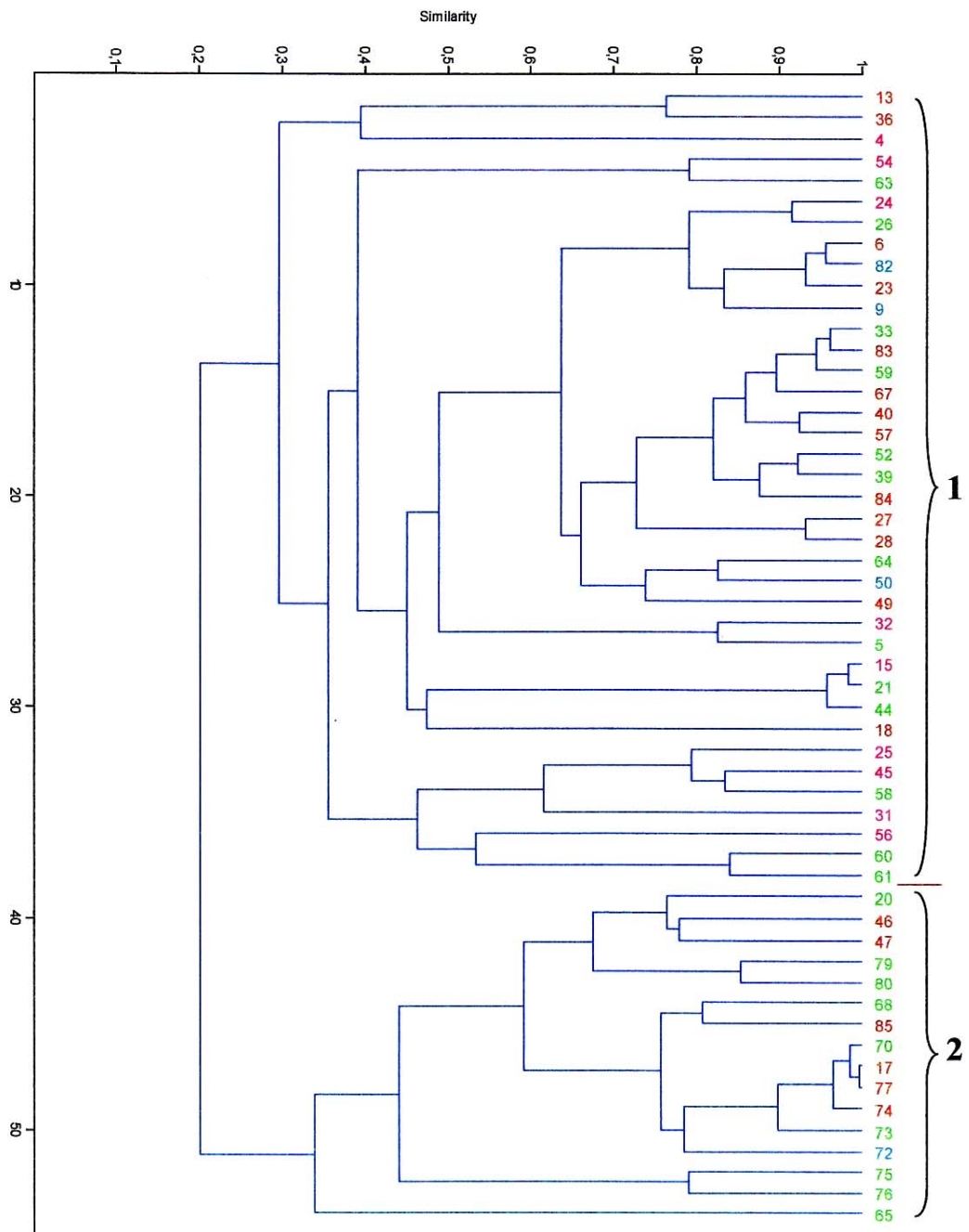


Fig. 42: Cluster analysis – clustering of samples of forest vegetation, using the algorithm of UPGMA and MORISITA's index

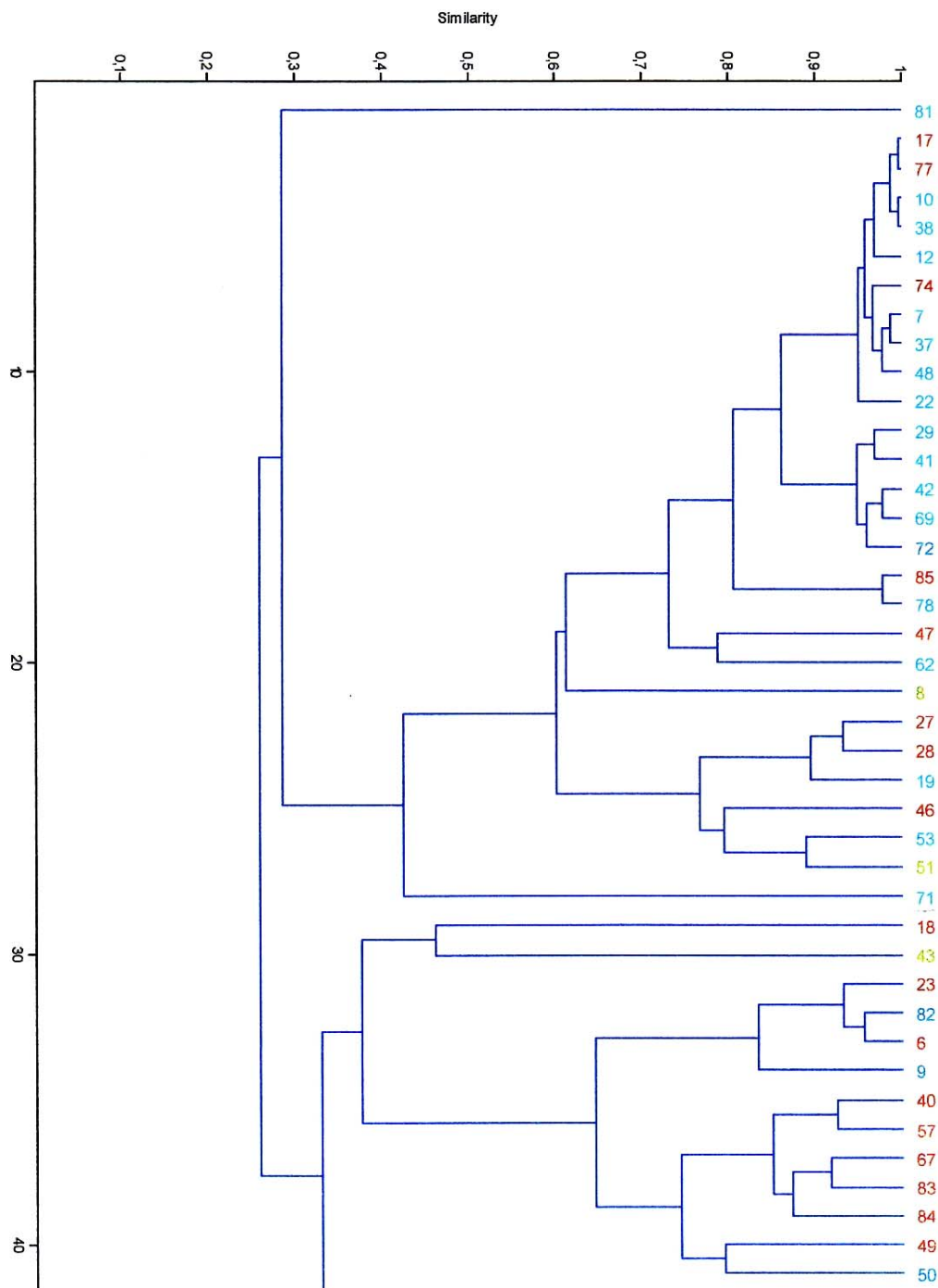


Fig. 43: Cluster analysis – clustering of samples of grassland, Capões, Capoeira and gallery forest vegetation, using the algorithm of UPGMA and MORISITA's index

3.5. Comparison of vegetation mapping with the palynological analysis

Five categories of relationships between plants and the appearance of their pollen grains or spores in the soil were detected (Tab. 4):

The first category (Tab. 4, yellow accented taxa) includes plants which occur frequently with a high dominance and abundance (high AD-value). Asteraceae (mainly herbaceous forms, but also shrubs) and Poaceae are examples for this category. Poaceae occur not only in grassland and Capoeira vegetation but also in forest vegetation where it even exhibits a high dominance and abundance. The palynological analysis presents a similar pattern. Pollen grains of Poaceae occur of course in Campo and Capoeira vegetation (there with a conspicuously higher amount) as well as in forest vegetation (Figs. 32 and 33). While herbaceous Asteraceae grow mainly in grassland and Capoeira vegetation, shrubby Asteraceae occur mainly in the skirts of forests. The palynological spectra of the respective vegetation types reflect this aspect (Figs. 28 to 34).

The second category (Tab. 4, red accented taxa) encloses taxa which are characteristic for a certain vegetation type. *Araucaria angustifolia* – the eponymous element of *Araucaria* forest - *Dicksonia sellowiana*, *Drymis brasiliensis* and *Vernonia spp.* (mainly *Vernonia discolor*) are mentioned exemplarily for taxa belonging to this group (Tab. 4, red accented taxa). Whereas *Araucaria angustifolia* is a typical element of *Araucaria* forests, Capões and gallery forests, *Dicksonia sellowiana* is almost exclusively related to *Araucaria* forests. *Drymis brasiliensis* is prevalent in Capões and partially in gallery forests. It is an element of the skirts of Capões. *Vernonia* also prefers the skirts of the wood but is less frequent than *Drymis brasiliensis*. Respectively, the palynomorphs of these taxa determine the palynological spectra of the accordant forest type.

The third category (Tab. 4, green accented taxa) encloses taxa which are characteristic for a certain vegetation type but their palynomorphs did not occur frequently in the palynological

spectra of the accordant vegetation type. Leguminosae (with dendriform habit) and Piperaceae should be mentioned exemplarily for typical taxa of broad-leaved evergreen forests, semideciduous and deciduous forests. They are characteristic taxa of these forest types. They even distinguish the character of these forest types. However, in the palynological spectra of the correspondent forest type they appear sporadically and they were not considered in further analyses. In the cases of *Casearia* spp. (Flacourtiaceae), Melastomataceae, Myrtaceae and *Daphnopsis* spp. (Thymeleaceae), it is similar (Tab. 4). The Melastomataceae-species *Miconia hiemalis* and *Miconia cinerascens* as well as the Flacourtiaceae-species *Casearia decandra* and *Casearia sylvestris* and the Thymeleaceae-species *Daphnopsis fasciculate* and *Daphnopsis racemosa* are typical elements of *Araucaria* forests, Capões and gallery forests but their pollen are rare in the palynological spectra of these forest types. Myrtaceae, which occur in an extremely high diversity of species in all forest types, appear rare in the palynological spectra.

The fourth category (Tab. 4, blue accented taxa) includes taxa which emerge in forests but with a high variability of abundance and dominance. The palynological spectra of these forests demonstrate this variability related to the palynomorphs. *Podocarpus lambertii* represents one example of this relationship between vegetation and pollen presence. *Podocarpus lambertii* occurs in forests but with a different abundance and dominance of specimens. *Podocarpus lambertii*-pollen grains showed the same variability.

Finally, the fifth category (Tab. 4, brown accented taxa) includes epiphytes like Bromeliaceae and Orchidaceae. They mainly emerged in *Araucaria* forests, Capões and gallery forests but in the palynological spectra they rarely appeared (Bromeliaceae) or are absent (Orchidaceae).

The other taxa (non-accented taxa) do not show a clear relationship between the taxon and a certain vegetation type. They cannot be used as indicator-taxa.

	semi-deciduous and deciduous forest			Campo			Capoeira		
	1	2	3	1	2	3	1	2	3
subareas									
Lithraea+Schinus (Anacardiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	2,2
Ilex spp.(Aquifoliaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Vernonia spp. (Asteraceae)	0,0	0,0	0,0	0,0	0,1	0,8	0,0	0,0	0,0
Asteraceae (general)	0,0	0,0	0,0	0,5	2,1	2,5	5,2	4,7	3,9
Sloanea + Weinmannia	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Sebastiania commersoniana (Euphorbiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Actinostemon concolor (Euphorbiaceae)	0,0	0,0	0,7	0,0	0,0	0,0	0,0	0,0	0,0
Alchornea triplinervia (Euphorbiaceae)	0,0	0,0	0,5	0,0	0,0	0,0	0,0	0,0	0,0
Euphorbiaceae (general)	0,0	0,0	0,3	0,0	0,0	0,0	0,0	0,0	0,0
Casearia spp.(Flacourtiaceae)	0,0	0,0	0,0	0,0	0,0	0,8	0,0	0,0	0,0
Lauraceae	0,0	0,0	0,7	0,0	0,0	0,0	0,0	0,0	0,0
Leguminosae	2,4	1,2	1,4	0,2	0,3	0,0	0,0	0,0	0,0
Melastomataceae	0,0	0,6	0,0	0,2	0,5	0,4	0,0	0,6	0,0
Cabralea canjerana (Meliaceae)	0,0	0,0	0,3	0,0	0,0	0,0	0,0	0,0	0,0
Trichilia spp.(Meliaceae)	0,0	0,0	0,7	0,0	0,0	0,0	0,0	0,0	0,0
Meliaceae (general)	0,0	0,0	0,7	0,0	0,0	0,0	0,0	0,0	0,0
Myrsine spp.(Myrsinaceae)	0,0	0,0	0,9	0,0	0,0	0,0	0,0	0,0	0,0
Myrtaceae (general)	0,0	2,0	1,7	0,0	0,0	0,4	2,4	0,0	0,0
Piperaceae	2,4	0,0	2,1	0,0	0,0	0,0	0,0	0,0	0,0
Matayba elaeagnoides (Sapindaceae)	0,0	0,0	0,4	0,0	0,0	0,0	0,0	0,0	0,0
Cupania spp. (Sapindaceae)	0,0	0,0	1,6	0,0	0,0	0,0	0,0	0,0	0,0
Allophylus spp. (Sapindaceae)	0,0	0,0	0,3	0,0	0,0	0,0	0,0	0,0	0,0

Tab. 4: Average dominance and abundance (AD-values) of selected plant taxa in the different vegetation types and the different sub-areas

	semi-deciduous and deciduous forest			Campo			Capoeira		
	1	2	3	1	2	3	1	2	3
subareas									
Solanaceae (general)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Solanum spp. (Solanaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Daphnopsis spp. (Thymeleaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	2,4
Drymis brasiliensis (Winteraceae)	0,0	0,0	0,0	0,0	0,0	0,8	0,0	0,0	0,0
Bromeliaceae	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Cyperaceae	0,0	0,0	0,0	0,0	0,7	0,0	0,0	1,2	0,0
Orchidiaceae	0,0	0,6	0,5	0,0	0,0	0,0	0,0	0,0	0,0
Poaceae	2,4	0,0	0,6	6,4	6,4	2,3	7,0	6,3	7,0
Araucaria angustifolia (Araucariaceae)	0,0	2,3	0,0	0,0	0,0	0,0	0,0	1,1	0,0
Podocarpus lambertii (Podocarpaceae).	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Lycopodium sp. (Lycopodiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,6	0,0
Sphagnum sp. (Sphagnaceae)	0,0	0,0	0,0	0,5	0,4	0,0	0,0	0,0	0,0
Asplenium spp. (Aspleniaceae)	1,1	0,6	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Blechnum imperiale (Blechnaceae)	0,0	0,0	0,6	0,0	0,3	0,0	0,0	0,0	0,0
Alsophila sp. (Cyatheaceae)	0,0	0,0	0,3	0,0	0,0	0,0	0,0	0,0	0,0
Dennstaedtiaceae	0,0	3,5	1,6	0,0	0,7	1,6	0,0	0,0	1,2
Dicksonia sellowiana (Dicksoniaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Gleichenia sp. (Gleicheniaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Pteridophyta (general)	0,0	0,6	0,0	0,0	0,0	0,0	0,0	1,3	0,0

Tab. 4: Average dominance and abundance (AD-values) of selected plant taxa in the different vegetation types and the different sub-areas

subareas	gallery forest			Capão			Araucaria forest		
	1	2	3	1	2	3	1	2	3
Lithraea+Schinus (Anacardiaceae)	0,0	0,0	0,0	0,0	1,0	0,0	0,0	0,3	0,2
Ilex spp.(Aquifoliaceae)	0,0	0,0	2,4	0,2	0,0	1,0	0,0	0,3	1,6
Vernonia spp. (Asteraceae)	0,0	0,0	0,0	0,3	0,4	0,0	0,0	0,0	0,2
Asteraceae (general)	0,0	2,8	0,0	0,3	0,0	0,6	0,0	0,6	0,0
Sloanea + Weinmannia	0,0	0,0	0,0	0,0	0,0	0,0	0,6	0,3	0,2
Sebastiania commersoniana (Euphorbiaceae)	0,0	0,0	2,4	0,0	0,0	0,0	0,0	0,0	0,2
Actinostemon concolor (Euphorbiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,2
Alchornea triplinervia (Euphorbiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Euphorbiaceae (general)	0,0	0,0	0,0	0,0	0,0	0,6	0,0	0,5	0,3
Casearia spp.(Flacourtiaceae)	0,0	0,0	0,0	0,0	0,8	0,6	0,0	0,7	1,1
Lauraceae	0,0	1,8	0,0	0,4	0,0	0,0	0,6	0,8	1,1
Leguminosae	0,0	0,0	0,0	0,1	0,0	0,0	0,0	0,0	0,4
Melastomataceae	3,9	0,6	2,4	1,5	2,5	0,7	1,2	1,4	1,9
Cabralea canjerana (Meliaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,4
Trichilia spp.(Meliaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,2
Meliaceae (general)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Myrsine spp.(Myrsinaceae)	0,0	0,0	0,0	0,5	0,2	0,0	0,6	0,3	1,0
Myrtaceae (general)	3,6	2,0	3,6	3,4	3,0	3,2	3,2	2,0	2,4
Piperaceae	0,0	0,0	0,0	0,0	0,0	0,7	0,0	0,0	0,5
Matayba elaeagnoides (Sapindaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,6
Cupania spp. (Sapindaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,9
Allophylus spp. (Sapindaceae)	0,0	0,6	0,0	0,3	0,0	0,0	0,0	0,0	0,1

Tab. 4: Average dominance and abundance (AD-values) of selected plant taxa in the different vegetation types and the different sub-areas

	gallery forest			Capão			Araucaria forest		
	1	2	3	1	2	3	1	2	3
subareas									
Solanaceae (general)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,3	0,2
Solanum spp. (Solanaceae)	0,0	0,0	2,4	0,0	0,2	0,6	0,0	0,0	0,1
Daphnopsis spp. (Thymeleaceae)	2,4	0,0	0,0	1,4	2,4	1,3	0,0	1,0	0,8
Drymis brasiliensis (Winteraceae)	0,0	0,0	2,4	1,2	0,8	0,0	0,0	0,4	0,3
Bromeliaceae	1,4	0,7	1,3	0,9	1,3	0,3	1,3	0,8	0,4
Cyperaceae	0,0	0,0	1,1	0,0	0,0	0,0	2,6	0,0	0,6
Orchidiaceae	0,0	0,0	0,0	0,3	0,2	0,3	0,0	0,4	0,2
Poaceae	2,4	0,0	2,6	2,2	2,1	4,4	0,0	2,4	1,4
Araucaria angustifolia (Araucariaceae)	2,4	2,6	0,0	2,7	3,3	3,4	3,0	3,0	4,0
Podocarpus lambertii (Podocarpaceae)	2,4	0,0	0,0	0,7	0,4	0,0	0,0	0,5	0,7
Lycopodium sp. (Lycopodiaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Sphagnum sp. (Sphagnaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Asplenium spp. (Aspleniaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Blechnum (Blechnaceae)	0,0	0,0	0,0	0,3	0,4	0,6	0,0	0,1	0,0
Alsophila sp. (Cyatheaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Dennstaedtiaceae	0,0	0,6	0,0	0,0	0,0	0,0	0,0	0,0	0,0
Dicksonia sellowiana (Dicksoniaceae)	0,0	0,0	0,0	0,0	0,0	0,9	1,2	1,1	0,4
Gleichenia sp. (Gleicheniaceae)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,1
Pteridophyta (general)	0,0	0,0	0,0	0,0	0,0	0,0	0,0	0,3	0,0

Tab. 4: Average dominance and abundance (AD-values) of selected plant taxa in the different vegetation types and the different sub-areas

4. Discussion and conclusion

4.1 Discussion

The intense correlation between vegetation and palynological spectra had already been demonstrated by different authors. While e.g. BIRKS (1981), FIRBAS (1949), SALGADO-LABOURIAU (1997) and STRAKA (1957) attempted to reconstruct the palaeovegetation based on palynological analysis, other authors accomplished a direct comparison of palynological spectra and the contemporary vegetation. DAVIES & FALL (2001) studied the modern pollen precipitation from an elevational transect in central Jordan and layed open its relationship to vegetation. CONNOR et al. (2004) conducted a similar study in southern Georgia, Caucasus region. ELENGA et al. 2000, EL GHAZALI & MOORE 1998, GAJEWSKI 2002 executed similar studies in Africa.

This work has shown that recent pollen data from the Planalto region of Rio Grande do Sul show differences between various vegetation types. The different vegetation types can be separated by the relative proportion of different taxa as shown in the palynological spectra, by the occurrences of pollen/ spore markers sorted by interpolated raster data, by principal component analysis and cluster analysis.

4.1.1 Interpretation of palynological spectra from TILIA-analyses and interpolated raster data (ArcGIS)

Palynological analyses of the present study, performed with TILIA and also interpolated raster data from ArcGIS, demonstrate that a separation between the different vegetation types is feasible. Like MARCHANT et al. (2001) who reconstructed different biomes derived from modern pollen data along an altitudinal gradient in Colombia, this study aimed to reconstruct different vegetation types along a regional gradient in the Planalto region of Rio Grande do Sul. The presence and abundance of pollen and spores are controlled by the

chemism of the pollen-exine and the sporoderm, by the dispersion mechanism of pollen and spores. DENG et al. (2004) refer emphatically that the quantitative relationship between source taxon vegetation frequency and its pollen presentation varies between species due to differential pollen production and dispersal. VINCENS et al. (2000) disclose this as a general feature in many tropical regions. Processes of degradation, which are controlled by climatic factors, also affect on the presence and abundance of the palynomorphs. The climatic factors include intense rain, subtropical temperatures, etc. interacting with the soil texture and stratigraphic conditions (EBNER 2005). This point of view has always to be taken into consideration. For example, soils situated on slopes, subjected to erosion and rearrangement including their palynomorphs which, themselves, modify the palynological spectrum. Taking into account all these factors, local vegetation influences the palynological spectra and consequently, the different vegetation types will still be distinguished on the basis of palynological spectra.

The fundamental question which was posed was confirmed in this work with regard to:

1. grassland/forest-level
2. different grassland types and
3. different forest types

1. *grassland/forest-level*

The present study confirms that grassland is determined primarily by pollen grains of Poaceae (almost 50%) while the soil samples of forests contained pollen grains of Poaceae from 20 to 30%. The conspicuous amount of Cyperaceae pollen is worth being mentioned in soil samples of grassland. Accessory pollen of *Araucaria angustifolia*, *Podocarpus lambertii* and taxa of Asteraceae are worth being listed as well. REESE & LIU (2005) described the dominance of grass (Poaceae) in combination with moderate frequencies of Asteraceae pollen, which tend to increase slightly towards the southern (drier) locations as a characteristic of the Altiplano of the central Andes region of South America.

The pteridophytes group is the predominant one in soil samples of forests, where up to almost 60% of the total amount of palynomorphs were found in *Araucaria* forests, up to almost 50% in semi-deciduous and deciduous forests and of 40-50% in Capões. Furthermore, pollen of shrub and tree species, e.g. *Araucaria angustifolia*, *Podocarpus lambertii*, *Ilex sp.*, Euphorbiaceae and Myrtaceae, emerge stronger in soil samples of forest vegetation than in soil samples of grassland vegetation. In contrast to forests, the amount of pteridophytes reaches a maximum of 20% in soil samples of grassland. In general, the proportion of fern spores, arboreal pollen (e.g. of *Araucaria angustifolia*, *Podocarpus lambertii*, *Ilex sp.*, Euphorbiaceae and Myrtaceae) and non-arboreal pollen, especially Poaceae pollen, reflect the physiognomy of the forests and grasslands. EBNER (2005) described similar results for the Pró-Mata area. Gallery forests reveal an intermediate situation with maximum 25% spores of pteridophytes and a scarce pollen amount of tree species. This reflects the situation of the recent vegetation where gallery forests represent open vegetation alongside rivers interacting intensely with grasslands. BROWN (1987) constituted gallery forests as paths for exchange of floristic elements of different forest biomes like Mata Atlântica and Amazonia. Gallery forests present a transitional vegetation which is not clearly separable from other forest types.

2. *different grassland types*

On the basis of palynological analysis two types of grassland are distinguished. While spores of the bryophytes *Anthoceros punctatus* and *Phaeoceros laevis* were found in soil samples of dry grassland, spores of *Sphagnum* occurred exclusively in wet grassland. Consequently, *Sphagnum* represents an indicator for wet subsurface and swamp. Swamped grassland interferes frequently in recent vegetation with dry grassland, especially with neighbouring Capões. PAEZ et al. (1998) separated three different steppe types by multivariate analysis of pollen samples.

3. *different forest types*

A determination of the three forest types – *Araucaria* forest, semi-deciduous and deciduous forest – is feasible by means of availability and frequency of tree fern spores. Investigations of the vegetation carried out by KLEIN (1984) confirmed that genera of Cyatheaceae occur on slopes of the Serra Geral especially in subtropical broad-leaved evergreen forests while species of *Dicksonia* prefer the Planalto. In this investigation, spores of Cyatheaceae appear mainly on slopes of the Serra Geral, the thick-walled spores of *Dicksonia sellowiana* emerge exclusively on the Planalto. This aspect correlates with the forest types because the eastern and south-eastern slopes of Serra Geral are covered with semi-deciduous forests influenced by coastal, subtropical broad-leaved evergreen forests while the Planalto is exclusively covered by *Araucaria* forests and Capões. This aspect corroborates the hypothesis that it is feasible to determine the two types of forests - *Araucaria* forest and semi-deciduous forest. In relation to deciduous forests, this hypothesis is invalid. In fact, deciduous forests are located on the slopes of the Serra Geral but tree fern spores are less relevant. The drier climate upcountry (SCHULTZE 2000) that requires an equivalent forest – a deciduous forest – is probably the reason why the amount of tree ferns and consequently tree fern spores decline. Furthermore, the influence of elements of the broad-leaved evergreen forest of the coastal slopes of the Serra Geral, which passed the “Gate of Torres” (RAMBO 1951), diminishes from the East westward. That means that a determination of the three mentioned forest types is practicable by tree fern spores of palynological spectra. This result corresponds with EBNER’s result (2005) who detected the frequency of the two spore types of tree ferns depending on the location/ forest type within the Pró-Mata area. Apart from the above mentioned, spores of *Asplenium sp.*, pollen of *Araucaria angustifolia*, *Podocarpus lambertii* and accessory palynomorphs are relevant for a determination of the forest types. With exception of two soil samples (samples No.15 and 32), *Asplenium* spores were found regularly in semi-deciduous and deciduous forests, but in a low amount. In contrast, *Asplenium sp.* is irrelevant in *Araucaria* forests. *Asplenium sp.* could represent a discriminatory species which is decisive for a vegetation type (DIERSCHKE 1994) – in this

case for semi-deciduous and deciduous forests. All these facts prove the *Araucaria* forests can be distinguished from the deciduous forests. But in contrast to the proposition of BONNEFILLE et al. (1999), detecting deciduous forests are characterised by abundant Melastomataceae pollen, this study does not confirm this statement. In the present study Melastomataceae pollen are rare or absent in *Araucaria* forests, Capões, gallery forests as well as in deciduous forests, although the source plants are well-presented in the contemporary vegetation. Pollen of *Araucaria angustifolia* and *Podocarpus lambertii* indicate *Araucaria* forests and Capões, while they fail to appear in semi-deciduous and deciduous woods. DENG et al. (2004) described similar effects for modern pollen-vegetation relationships along transects on the Whangapoua Estuary. They emphasise the presence of discriminatory species, even in small pollen amounts, will allow correct identification of the local vegetation represented in sedimentary palynological sequences. In the course of the present work it was also ascertained that the number of pollen/ spore taxa identified in each sample of the semi-deciduous and deciduous forests is notably less than that obtained in each sample of the other vegetation types. BONNEFILLE et al. (1999) also mentioned this observation in the deciduous forests of South India. This result would agree with a lower plant diversity estimated for deciduous forests (GHATE et al. 1998)

Similar to the intermediate position of gallery forest implied by pteridophytes spores, gallery forests are exceptions in relation to tree fern spores and pollen of *Araucaria angustifolia* and *Podocarpus lambertii* because they were rare.

The position of Capões is in so far interesting as the latter are less characterised by pollen of *Araucaria angustifolia* rather than *Podocarpus lambertii*, *Ilex sp.* and *Drymis brasiliensis* which are well-represented. *Podocarpus lambertii*, *Ilex sp.* and *Drymis brasiliensis* are classified as settlement pioneers of vegetation and prefer rather open vegetation (EBNER 2005). These conditions match those of Capões which have an open mantle of vegetation interacting with surrounding grassland vegetation. Their margin is favoured by species of *Podocarpus lambertii*, *Ilex sp.* and *Drymis brasiliensis*. The increased value of Poaceae pollen (up to 40%) in Capões can be attributed to aeolian input

from nearby grasslands. This reflects the mosaic pattern of Capões and grasslands. BONNEFILLE et al. (1999) ascertained similar effects for the mosaic pattern of shoals and grasslands in South India.

4.1.2 Interpretation of the PCA results

Results deriving from two different methodologies were compared in juxta position: Palynomorphs, extracted from soil and counted thereafter, were analysed with multivariate statistical methods to verify values resulting from assessing palynological spectra. PCA was one of the applied multivariate statistical methods. The palynological spectra arrange in distinct cluster groups subjected to the main vegetation types. These results correspond to EBNER (2005) who introduced the term 'fingerprint' for palynological spectra of upper floor samples for identification of ecological units. ANUPAMA et al. (2000) distinguished between deciduous and evergreen forest of the southern Eastern Ghats of India. But in contrast to his work, in the present study the separation of semi-deciduous and deciduous forests is less distinct. This reflects the secret transition from semideciduous to deciduous forest. PCA resulted in a separation of palynological spectra of forest vegetation and grassland vegetation along the first axis (component). The separation of *Araucaria* forest and semi-deciduous/ deciduous forest takes place along the negative end of the first axis (component). The smooth transition, from semi-deciduous/ deciduous forest on the slopes of the Serra Geral to *Araucaria* forest on the border of the Planalto, is reflected in the PCA-charts.

Capões represent ties between forests on the one hand and grassland on the other hand accordingly they arrange in the PCA between the forest clusters and the grassland cluster.

Pollen assemblages of Capoeira vegetation cluster in a large diffuse cloud. Their palynological spectra seem to represent pollen and spores of the previous vegetation (vegetation before deforestation) because the palynological spectra vary intensively from the source vegetation. If semi-deciduous forest presented the previous vegetation, the

palynological spectra of Capoeira is analogue to the palynological spectra of semi-deciduous forest.

Capões, gallery forests and Capoeira are transitional vegetation types and their palynological spectra are not clearly separable from those of the other vegetation types. BONNEFILLE et al. (1999) described a similar behaviour of transitional forests in South India.

The loadings of taxa corroborate the mentioned characteristics. This signifies that the vegetation types are characterised by a special composition of palynomorphs. Poaceae, Cyperaceae and Anthocerotaceae cluster on the grassland side while typical forest taxa like spores of *Blechnum*, *Microgramma*, *Dicksonia sellowiana* and Cyatheaceae cluster on the forest side. LIU et al. (1999) noted characteristic combinations of pollen types for each analysed vegetation zone in south-eastern Inner Mongolia. The quintessence of EBNER's thesis (2005) expresses the same idea. However, the separation of the different types of vegetations is less distinct than shown in EBNER (2005). He described the separation of the vegetation types as a result evoked by a moisture gradient. This aspect can also be seen in the study area because the mean annual rainfall, which presents an east-to west gradient caused by orography, induces the release of rains on the eastern slopes of the Serra Geral and the eastern edge of the Planalto and blocks the inland progression (MORENO 1961). The decreasing east to west rainfall gradient limits both the expansion of subtropical broad-leaved evergreen forest on the slopes of the Serra Geral and of widespread *Araucaria* forest on the Planalto. SCHNECK (2005) described PCA as an excellent method for separation of vegetation types. This study is in accordance with his results and results of other groups: VINCENS et al. (2000) confirm that pollen data from soil samples reflect well the recent transgression of forest versus savanna vegetation in Cameroon. STUTZ et al. (2003) adduce the relationship of modern pollen and vegetation on the basis of a study along a coast to inland gradient (coastal lagoon area and Pampa grassland) in Mar Chiquita (Argentina).

4.1.3 Interpretation of the cluster analysis results

The second applied multivariate statistic method – the cluster analysis – confirms the results of the previous methods shown above. By applying cluster analysis, the soil samples cluster on the basis of palynological spectra in two cardinal groups – the ‘grassland group’ and the ‘forest group’. Further divisions within these cardinal groups are not feasible. This is also reflected by two distinct paths in the recent vegetation. One path describes semi-deciduous and deciduous forests on the slopes of Serra Geral which pass into *Araucaria* forest on the border of the Planalto. The second path describes *Araucaria* forest, Capões and gallery forest interacting intensively with grasslands (Campos). The palynological spectra of soil samples of Capoeira and anthropogenic areas represent, beside the pollen and spores of the recent vegetation, also the palynomorphs of the previous vegetation. Therefore, they scatter within the cardinal clusters. The cluster analysis of EBNER (2005) demonstrates a similar disperse distribution of the vegetation types. The present study confirms the proposition of SCHNECK (2005) referring to an indistinct separation of different vegetation types. In contrast to the present work, COURT-PICON et al. (2005) observed a good agreement between classifications and ordinations of the data-sets and the palynological separation of different vegetation types.

4.1.4 Interpretation of the palynological analysis and the mapping of vegetation

The palynological spectrum of soil depends not only on the abundance of the taxa or their pollen production but also depends on:

1. the dispersion mechanism of plants
2. the texture of pollen/ spores and
3. soil processes

1. Dispersion mechanism of plants

Some plants, e.g. ferns, gymnosperms (*e.g. Ilex, Pinus*) and Poaceae do not dispose on a special dispersion mechanism or are anemophilous plants, respectively. Considering this aspect, these plants produce numerous pollen/ spores which are typically well-represented in the palynological spectra of soil. Their presence ought to be interpreted carefully. In contrast, pollen of zoophilous plants, as Bignoniaceae, Malpighiaceae, Orchidaceae and Piperaceae, are rare or lack in the palynological spectra. Orchidaceae belong to zoophilous plants and the pollen grains are clustered to a polinarium. Orchids are well-represented in forests but their pollen lack in palynological spectra.

2. Texture of pollen/ spores

The rare presence or absence of *Araucaria angustifolia* pollen and Lauraceae pollen, respectively, are the consequence of their delicate pollen structure (ROJ & VAN DER WERFF 1988). Therefore, they are expeditiously damageable. The spores of *Dicksonia* are thick-walled and therefore they are well-presented in palynological spectra.

3. *Soil processes*

The palynomorphs in the soil are exposed to processes of pedogenesis and degradation. As a result, delicate pollen/ spores are corroded strongly and lack in palynological spectra. The adjacent soil preparation for extraction of the palynomorphs enforces this effect.

4.2 Conclusion

Although pollen/ spore diversity does not exactly reproduce the plant diversity, this study concludes that the main features of the vegetation composition are preserved in the pollen assemblages. The separation of different vegetation types has been proved by the relative proportion of different taxa as shown in the palynological spectra (TILIA analyses), by the occurrences of pollen/ spore markers sorted by interpolated raster data (ArcGIS), by principal component analysis and cluster analysis. Therefore, this study leads to the conclusion that the biomization procedure might be applicable to this region and that propositions, which are valid in a defined area, have validity in a widespread area of the Planalto and the Serra Geral as well.

Such concepts are advantageous for reconstructions of biomes from pollen/ spore data and modelling past vegetation in Rio Grande do Sul and entire Brazil. BEHLING et al. (2004, 2005) have already a share in this field of study. STUTZ & PRIETO (2003) revealed by detrended corresponded analysis that fossil pollen assemblages have modern counterparts, permitting to interpret the vegetation history of an area in greater detail than previously possible. The method of BARTH & PINTO DA LUZ (2003), executed a palynological analysis of geopropolis sediments, would be an auxiliary method added to the methodology of the present study. They demonstrated that pollen spectra of geopropolis sediments reflected a vegetation characteristic of the southeast region of Brazil.

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Appendix I

Plates with figures of an extract of identified palynomorphs

The palynomorphs were observed under 400-times magnified. The LM-micrographs were also taken with the same magnification.

Plate 1, p.III:

Gymnosperms

Fig.1: *Araucaria angustifolia* (Araucariaceae): inaperturate, scbrate, spheroidal,
diameter of 65-80 μm

Fig.2: *Pinus sp.* (Pinaceae): vesiculate, bisaccate, diameter 80 -110 μm

Fig.3: *Podocarpus sp.*(Podocarpaceae): vesiculate, bisaccate,
diameter 49 -80 μm

Gymnosperms

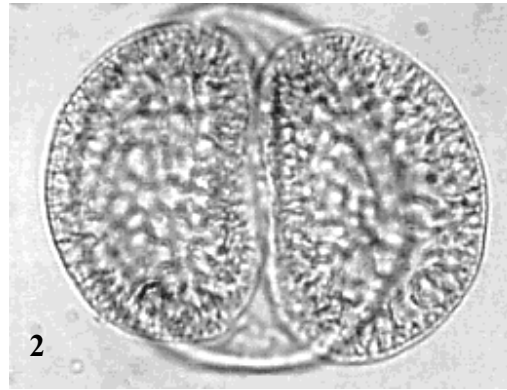
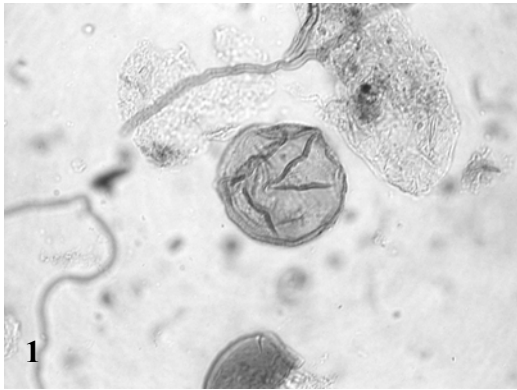


Plate 2, p.V

Angiosperms – Dicots

- Fig.4:** *Sebastiania cf. commersoniana* (Euphorbiaceae): tricolporate, microreticulate, spheroidal to prolate, colpi with smooth margin, pori conspicuously to observe
- Fig.5:** *Actinostemon concolor* (Euphorbiaceae): tricolporate, scabrate, prolate to subprolate, colpi with smooth margin, pori sometimes inconspicuous
- Fig.6:** *Alchornea triplinerva* (Euphorbiaceae): tricolporate, psilate to scabrate, suboblate
- Fig.7:** Melastomataceae: heterocolporate (tricolporate with 3 pseudocolpi), psilate, prolate
- Fig.8:** *Ilex* (Aquifoliaceae): tricolporoidate, clavate, subprolate to prolate, colpi and margin inconspicuous
- Fig.9:** *Cabrlea canjerana* (Meliaceae): tricolporate, psilate to slightly scabrate, spheroidal

Angiosperms - dicots

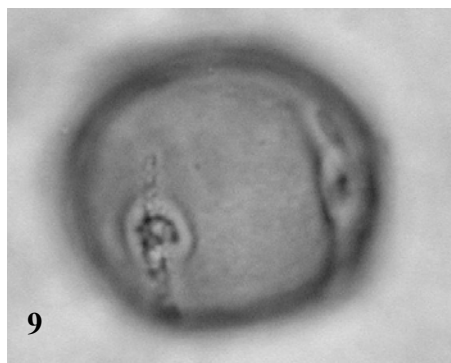
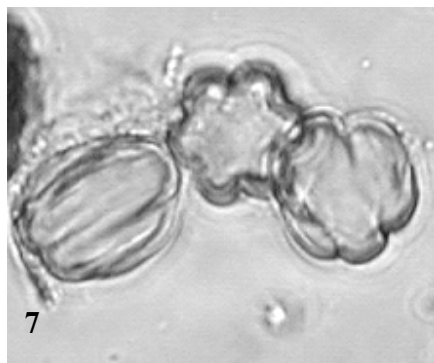


Plate 3, p. VII

Angiosperms

- Fig.10:** *Daphnopsis* (Thymeleaceae): periporate, baculate, spheroidal
- Fig.11:** *Lithraea-Schinus*-type (Anacardiaceae): tricolpate, striate, subprolate
- Fig.12:** *Celtis* (Ulmaceae): triporate, psilate to scabrate, spheroidal
- Fig.13:** *Weinmannia*-type (Cunoniaceae; including *Sloanea*/Elaeocarpaceae): tricolporoidate, microreticulate to psilate, prolate; fossil *Weinmannia* pollen are much alike *Sloanea* pollen therefore these two types were integrated in one type to avoid misidentifications (personal information of BEHLING 2003)
- Fig.14:** *Drymis brasiliensis* (Winteraceae): tetrads, single pollen grain inconspicuous zonosulcate, irregularly reticulate
- Fig.15:** *Myrsine* (Myrsinaceae): stephanocolpate (4 colpi), psilate to scabrate, spheroidal to subprolate

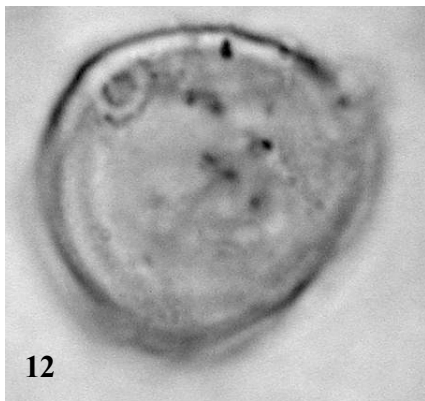
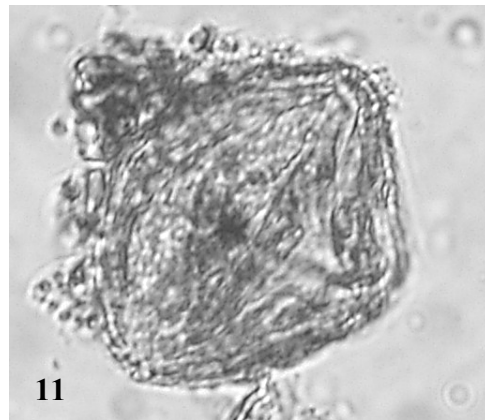
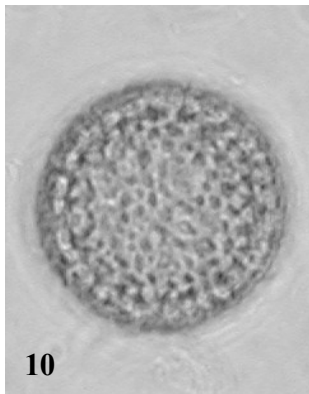


Plate 4, p. IX

Angiosperms - dicots

Fig.16: *Zanthoxylum* (Rutaceae): tricolporate, striate, subprolate to prolate

Fig.17: Myrtaceae: tricolporate, psilate to scabrate, peroblate

Fig.18: Solanaceae: tricolporate, psilate, subprolate to prolate

Fig.19: Loranthaceae: syncolpate, psilate to scabrate, peroblate, contour in polar view triangular with intense concave sides

Fig.20: *Cupania* (Sapindaceae): tricolporate, microreticulate, oblate, contour in polar view triangular with weakly sides

Fig.21: *Inga*-type (Mimosaceae): polyades, single pollen grains periporate, verrucate

Angiosperms - dicots

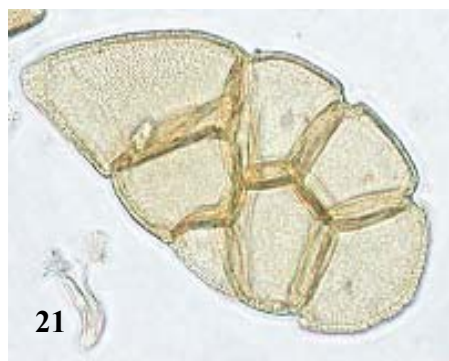
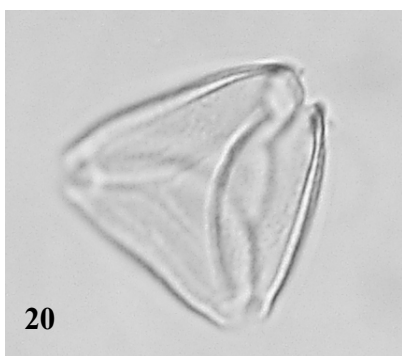
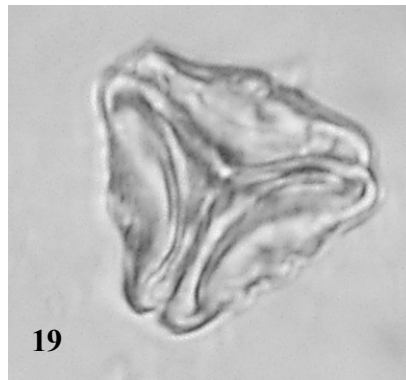
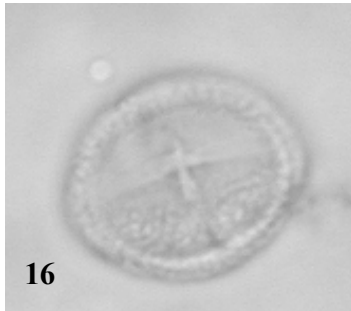


plate 5, p. XI

Angiosperms - dicots

- Fig.22:** *Vernonia*-type (Asteraceae): triporate, echinate, reticulate (fenestrate), spheroidal
- Fig.23:** Malvaceae: pantoporate, echinate, spheroidal, tectate
- Fig.24:** *Trixis* (Asteraceae): tricolporate, psilate to scabrate, prolate, exine thicker on the poles
- Fig.25:** *Polygonum* (Polygonaceae): pantoporate, reticulate, spheroidal
- Fig.26:** *Eryngium* (Apiaceae): tricolporate, psilate to scabrate, perprolate
- Fig.27:** *Borreria* (Rubiaceae): stephanocolporate, reticulate, suboblate to spheroidal
- Fig.28:** Amaranthaceae-Chenopodiaceae-type: periporate, reticulate, spheroidal

Angiosperms – dicots

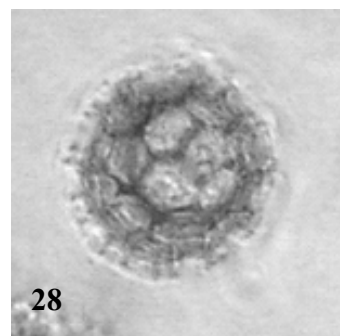
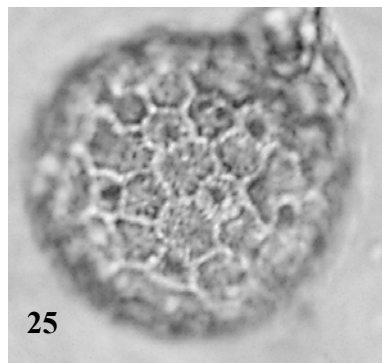
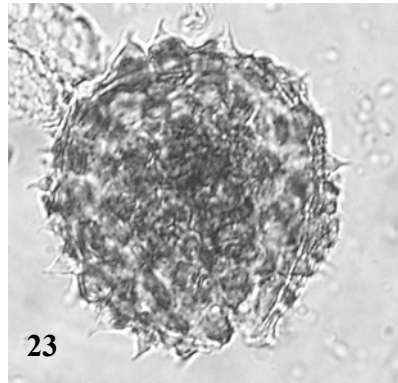
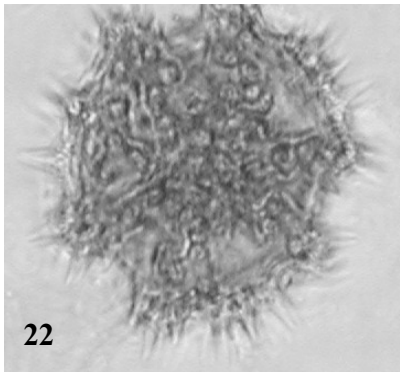


Plate 6, p. XIII

Angiosperms – dicots

Fig.29: *Baccharis*-type (Asteraceae): tricolporate, echinate, prolate spheroidal

Fig.30: *Senecio*-type (Asteraceae): tricolporate, echinate, spheroidal to subprolate

Angiosperms – monocots

Fig.31: Poaceae: monoporate, psilate to scabrate, spheroidal

Fig.32: cf. *Xyris* (Xyridaceae): dicolpate, reticulate, contour in equatorial view
biconvex

Angiosperms - dicots



Angiosperms – monocots



Plate 7, p. XV

Bryophytes

Fig.33: *Anthoceros punctatus* (Anthocerotaceae): trilete, echinate, trisymmetric

Fig.34: *Sphagnum* (Sphagnaceae): trilete, psilate to verrucate, trisymmetric

Pteridophytes – trilete spores

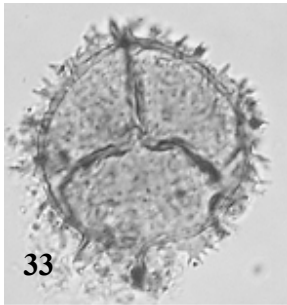
Fig.35: *Lycopodium* (Lycopodiaceae): trilete, reticulate, subtriangular-convex

Fig.36: *Selaginella* (Selaginellaceae): trilete, baculate to clavate, subtriangular-convex

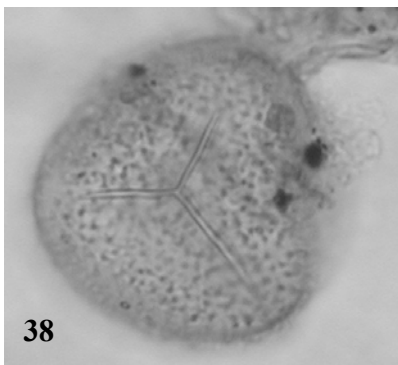
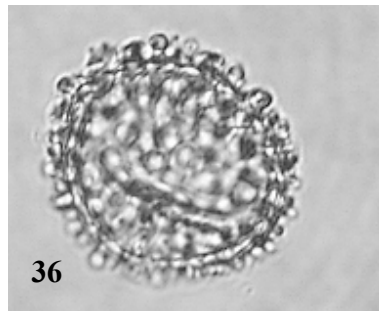
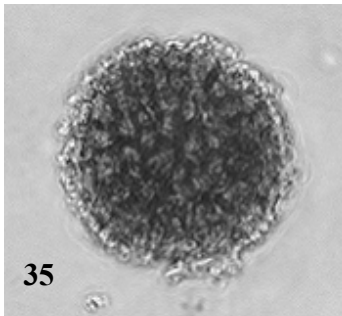
Fig.37: *Huperzia* (Lycopodiaceae): trilete, foveolate, subtriangular-convex

Fig.38: *Trichomanes* (Hymenophyllaceae): trilete, echinate-papillate, subtriangular-convex

Bryophytes



Pteridophytes – trilete spores



Plates 8, p. XVI

Pteridophytes – trilete spores

Fig.39: *Anemia raddiana* (Schizaceae): trilete, cicatricose (coarse, compact and parallel exospore ridges), triangulares

Fig.40: *Anemia phyllitides* (Schizaceae): trilete, cicatricose-baculate, triangulares

Fig.41: *Alsophila* (Cyatheaceae): trilete, laevigate, straight sides, rounded angles, triangulares

Fig.42: *Lophosoria quadripinnata* (Lophosoriaceae): trilete, tuberculate to perforate, rounded angles, triangulares

Fig.43: *Dicksonia sellowiana* (Dicksoniaceae): trilete, laevigate, straight to lightly convex sides, rounded angles, sporoderm thicker at the angles, triangulares

Fig.44: *Dennstaedtia* (Dennstaedtiaceae): trilete, eroded perispore, laevigate exospore, triangulares

Fig.45: *Dennstaedtia* (Dennstaedtiaceae): trilete, preserved perispore, verrucate, triangulares

Pteridophytes – trilete spores

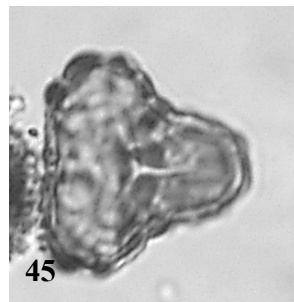
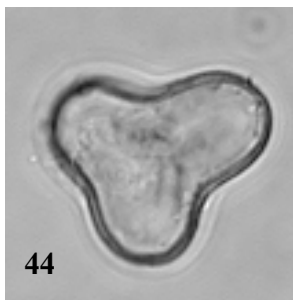
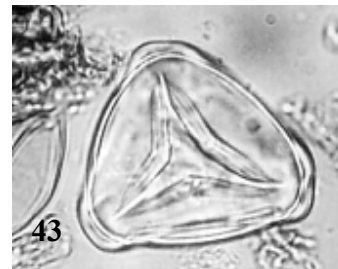
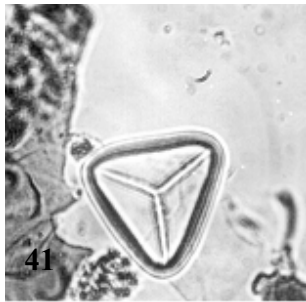
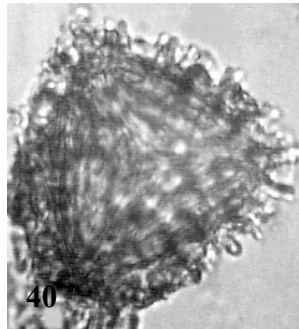


Plate 9, p. XIV

Pteridophytes – monolete spores

Fig.46: *Asplenium* (Aspleniaceae; including *Dryopteris*/ Dryopteridaceae): monolete, irregularly undulated, bilateral; fossil *Asplenium* spores are much alike *Dryopteris* spores therefore these two types were integrated in one type to avoid misidentifications

Fig.47: *Blechnum*-type (Blechnaceae): monolete, psilate, bilateral

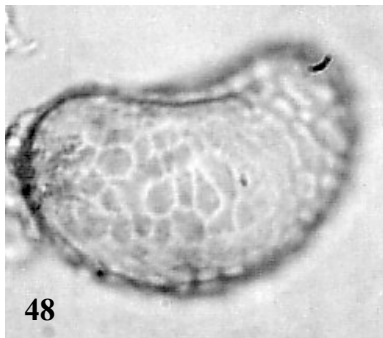
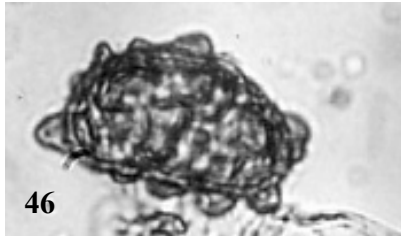
Fig.48: *Microgramma*-type (Polypodiaceae): monolete, verrucate, bilateral

Fungi

Fig.49: cf. *Helmintosporium*

Fig.50: Fungi

Pteridophytes – monolete spores



Fungi



Appendix II

1. List of identified palynomorphs

The undermentioned list numerates the indicated pollen and spores. The order follows the hierarchical system of the plant systematic (FROHNE & JENSEN, 1998). On a detailed description of pollen and spore taxa were renounced. Meticulous description about pteridophyte spores, appropriate plant, habitat and geographical distribution in Brazil are prevailed in LORSCHAITTER (1988, 1989) and in LORSCHAITTER et al. (1998, 1999, 2001 and 2002). Pollen and also spores are detailed described amongst others in BEHLING (1993), ROUBIK et al. (1991) and COLVINVAUX et al. (1999). The aforementioned references were very beneficial for pollen and spore identification on the genus level and the family level and sometimes also on the specie level.

1. *Anthoceros punctatus* (Anthocerotaceae)
2. *Phaeoceros laevis* (Anthocerotaceae)
3. *Sphagnum spp.* (Sphagnaceae)
4. *Lycopodiella* (Lycopodiaceae)
5. *Lycopodium alopecuroides* (Lycopodiaceae)
6. *Lycopodium clavatum* (Lycopodiaceae)
7. *Huperzia* (Lycopodiaceae)
8. *Selaginella* (Selaginellaceae)
9. *Hymenophyllum* (Hymenophyllaceae)
10. *Trichomanes* (Hymenophyllaceae)
11. *Osmunda* (Osmundaceae)
12. *Dicranopteris* (Gleicheniaceae)
13. *Gleichenia* (Gleicheniaceae)
14. *Anemia phyllitides* (Schizaceae)
15. *Anemia raddiana* (Schizaceae)
16. *Alsophila* (Cyatheaceae)

17. *Cyathea schaschin* (Cyatheaceae)
18. *Cyathea* (Cyatheaceae)
19. *Nephelea* (Cyatheaceae)
20. *Lophosoria quadripinnata* (Lophosoriaceae)
21. *Dicksonia sellowiana* (Dicksoniaceae)
22. *Doryopteris* (Hemionitidiaceae)
23. *Anogramma* (Gymnogrammaceae)
24. *Pteris sp.1* (Pteridaceae)
25. *Pteris sp.2* (Pteridaceae)
26. *Vittaria elongata* (Vittariaceae)
27. *Dennstaedtia* (Dennstaedtiaceae)
28. *Pteridium aquilinum* (Dennstaedtiaceae)
29. *Asplenium sp.1* (Aspleniaceae)
30. *Asplenium sp.2* (Aspleniaceae)
31. *Blechnum imperiale* (Blechnaceae)
32. *Dryopteris* (Dryopteridaceae)
33. *Davallia* (Davalliaceae)
34. *Microgramma* (Polypodiaceae)
35. *Pteridophyta 1*
36. *Pteridophyta 2*
36. *Pteridophyta 3*
38. *Araucaria angustifolia* (Araucariaceae)
39. *Pinus* (Pinaceae)
40. *Podocarpus* (Podocarpaceae)
41. *Drymis brasiliensis* (Winteraceae)
42. Annonaceae
43. Lauraceae
44. *Piper* (Piperaceae)
45. Amaranthaceae-Chenopodiaceae-type
46. *Polygonum* (Polygonaceae)

47. Ulmaceae
48. *Lamanonia speciosa* (Cunoniaceae)
49. *Weinmannia* (Cunoniaceae)
50. *Mimosa scabrella* (Leguminosae-Mimosoideae)
51. *Inga* (Leguminosae-Mimosoideae)
52. *Roupala* (Proteaceae)
53. Myrtaceae
54. Melastomataceae
55. *Fuchsia* (Onagraceae)
56. *Zanthoxylum* (Rutaceae)
57. *Lithraea-Schinus*-type
58. *Cupania* (Sapindaceae)
59. *Matayba* (Sapindaceae)
60. Erythroxyllaceae
61. *Cabrlea canjerana* (Meliaceae)
62. *Trichilia* (Meliaceae)
63. Meliaceae
64. *Polygala* (Polygalaceae)
65. *Actinostemon concolor* (Euphorbiaceae)
66. *Alchornea triplinerva* (Euphorbiaceae)
67. *Bernhardia pulchella* (Euphorbiaceae)
68. *Sebastiania commersoniana* (Euphorbiaceae)
69. Euphorbiaceae
70. *Casearia* (Flacourtiaceae)
71. *Daphnopsis* (Thymeleaceae)
72. Loranthaceae
73. *Eryngium* (Apiaceae)
74. Apiaceae
75. Malvaceae
76. Sterculiaceae

77. *Myrsine* (Myrsinaceae)
78. *Ilex divaricata* (Aquifoliaceae)
79. *Ilex 2* (Aquifoliaceae)
80. Ericaceae
81. *Clethra* (Clethraceae)
82. *Valeriana* (Valerianaceae)
83. *Borreria* (Rubiaceae)
84. *Psychotria* (Rubiaceae)
85. *Relbunium* (Rubiaceae)
86. Apocynaceae
87. Bignoniaceae
88. Acanthaceae
89. *Brunfelsia* (Solanaceae)
90. *Petunia* (Solanaceae)
91. *Solanum* (Solanaceae)
92. Solanaceae
93. Boraginaceae
94. *Achyrocline*-type (Asteraceae)
95. *Artemisia*-type (Asteraceae)
96. *Baccharis*-type (Asteraceae)
97. *Calea*-type (Asteraceae)
98. *Eupatorium* (Asteraceae)
99. *Mikania* (Asteraceae)
100. *Senecio*-type (Asteraceae)
101. *Trixis* (Asteraceae)
102. *Vernonia 1* (Asteraceae)
103. *Vernonia 2* (Asteraceae)
104. Asteraceae
105. Bromeliaceae
106. Cyperaceae

- 107. Poaceae
- 108. *Desmoncus*-type (Arecaceae)
- 109. *Palmae 1* (Arecaceae)
- 110. *Palmae 2* (Arecaceae)
- 111. *Palmae 3* (Arecaceae)
- 112. Iridaceae
- 113. Liliaceae
- 114. Xyridaceae
- 115. Amaryllidaceae
- 116. Monokotyle

2. List of pollen and spore groups

The pollen and spores were arranged in groups for the further work with different analysis methods (TILIA, CANOCO etc.). This arrangement facilitates a more concisely treatment of the data. The arrangement results from considering of morphological similarity of pollen or spores, taxonomic relationship or the similarity of the habitat respectively. The first column illustrates the groups (abbreviations were used) and the second column the taxa (reference number considering of the list above) which were incorporated in the appropriate group.

<i>Pollen or spore group</i>	<i>Incorporated taxa</i>
Anthoc	1, 2
Sphagi	3
Lyci	4, 5, 6
Hupe	7
Sela	8
Hytr	9, 10
Osmu	11
Ble	12, 13, 26, 31, 35
Anem	14, 15
Cyat	16, 17, 18, 19
Dickse	20, 21, 22
Anop	23, 24, 25
Micro	33, 34, 36
Denn	27, 28
Aspl	29, 30, 32
Pteri3	36
Angu	38

Pi	39
Podo	40
Drybra	41
Anno	42
Laura	43
Piper	44
AmaCh	45
Polygo	46
Ulma	47
Cunon	48, 49
Mimo	50, 51
Roup	52
Myrt	53
Melast	54
Fuchs	55
Zantho	56
Lithra	57
Sapin	58, 59
Ery	60
Meli	61, 62, 63
Polyga	64
Euphorb	65, 66, 67, 68, 69
Cada	70, 71
Lora	72
Apia	73, 74
Malva	75
Stercu	76
Myrs	77
Ilex	78, 79

Eric	80
Clethra	81
Valbore	82, 83, 85
Psycho	84
Apocy	86
Bigno	87
Acan	88
Solan	89, 90, 91, 92
Borangi	93
Comp 1	94, 95, 96, 97, 98, 100, 101, 104
Comp 2	99
Ver	102, 103
Brome	105
Cyper	106
Poa	107
Palm	108, 109, 110, 111
Mono	112, 113, 114, 115, 116

Appendix III

Tables of mapping of vegetation according to BRAUN-BLANQUET

- Table III.1 Mapping of vegetation of assemblages of semi-deciduous and deciduous forests
- Table III.2 Mapping of vegetation of assemblages of *Araucaria* forests
- Table III.3 Mapping of vegetation of assemblages of Capões
- Table III.4 Mapping of vegetation of assemblages of gallery forests
- Table III.5 Mapping of vegetation of assemblages of “clean” grassland
- Table III.6 Mapping of vegetation of assemblages of Capoeiras
- Table III.7 Mapping of vegetation of assemblages of anthropogenic affected areas

Zone 3

Specimen number	4	24	25	31	32	54	56
Date of record	5.5.02	18.7.02	18.7.02	21.7.02	21.7.02	23.1.04	23.1.04
1. Tree section							
Altitude (in m)	10	10	9	11	12	12	11
Angiosperms - Dicotyledons							
Bignoniaceae							
<i>Jacaranda cf. micrantha</i> Cham.							3p
Euphorbiaceae							
<i>Alchornea triplinervia</i> (Spreng.) Muell. Arg.						3p	
Myrtaceae							
<i>Eugenia neomyrtifolia</i> Sobral		2p					
Myrtaceae							
Meliaceae							
<i>Cedrela fissilis</i> Vell.				2p			
Mimosaceae	1p						
Moraceae							
<i>Ficus</i> sp.		2r					
Sapindaceae							
<i>Matayba elaeagnoides</i> Radlk.						2a	
Gymnosperms							
Araucariaceae							
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2r						
2. Tree section							
Altitude (in m)	7	7	6	9	8	10	9
Angiosperms - Dicotyledons							
Araliaceae						1p	
Bombacaceae					2r		

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Tiliaceae									
<i>Luehea divaricata</i> Mart.		2p							
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	1r								
Lianes									
<i>Arctolochia</i> sp.									r
<i>Smilax</i> sp.		p							
<i>Dioscorea multiflora</i> Mart.	p								
Shrub section									
Altitude (in m)									
Acanthaceae	2	3	2	3		2	2	2	
<i>Ruellia angustifolia</i> Sw.		3m					2p		
Euphorbiaceae									
<i>Pachystroma longifolium</i> (Ness) IM Johnst.							1r	1r	
<i>Endlicheria paniculata</i> (Spr.) Macbr.							2p		
<i>Nectandra megapota mica</i> (Sprengel) Mez							2p		
Leguminosae - Faboideae									
<i>Myrocarpus frondosus</i> Fr. Allem.							2p		
Leguminosae - Mimosoideae									
<i>Inga marginata</i> Willd.			2p				1p	1r	
Malvaceae									
<i>Byrthneria australis</i> St. - Hil. (Malvaceae sens. lat.)							2a		
Melastomataceae									
<i>Miconia cinerascens</i> Miq.									
<i>Miconia hiemalis</i> St. Hil.									
Meliaceae									
<i>Cabralea canjerana</i> Sald.		1p							
<i>Guarea macrophylla</i> Vahl.		2p							
<i>Trichilia clausseii</i> C. DC.		2p							

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Oxalidaceae									
<i>Oxalis</i> sp.					1p				
Passifloraceae									
<i>Passiflora</i> sp.				1r					
Piperaceae									
<i>Piper</i> sp.									
Proteaceae									
<i>Roupala</i> sp.									
Sapindaceae									
<i>Cupania verrallii</i> Camb.						1r		1p	
Urticaceae									
<i>Urena baccifera</i> (L.) Gaud.							1p		
Peridophyta									
Aspleniaceae									
<i>Asplenium raddianum</i> Gaud.		1p							
<i>Asplenium</i> sp. L.									
Blechnaceae									
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.							3a		
Dennstaedtiaceae									
<i>Lastreopsis amplissima</i> (Presl) Tindale							3a	5m	
Pteridaceae									
<i>Pteridophyta</i>		1p							
Epiphytes									
Angiosperms - Dicotyledons									
Cactaceae									
<i>Rhipsalis</i> sp.			p		p				
Angiosperms - Monokotyledons									
Orchidaceae									
<i>Oncidium hookeri</i> Rolfe.									
<i>Oncidium</i> sp.							p		
Orchidaceae									
<i>Pleurothallis hygrophila</i> Barb. Rodr.			p		p				

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

	Zone 2	Zone 1
Specimen number	15	45
Date of record	25.6.02	25.8.02
1. Tree section		
Altitude (in m)	10	9
Angiosperms - Dicotyledons		
Bignoniaceae		
<i>Jacaranda cf. micrantha</i> Cham.		
Euphorbiaceae		
<i>Alchorrea triplinervia</i> (Spreng.) Muell. Arg.		
Myrtaceae		
<i>Eugenia neomyrtifolia</i> Sobral		
Myrtaceae	3a	
Meliaceae		
<i>Cedrela fissilis</i> Vell.		
Mimosaceae		
Moraceae		
<i>Ficus</i> sp.		
Sapindaceae		
<i>Matayba elaeagnoides</i> Radlk.		
Gymnosperms		
Araucariaceae		
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p	
2. Tree section		
Altitude (in m)	8	6
Angiosperms - Dicotyledons		
Araliaceae		
Bombacaceae		

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Bignoniaceae		
<i>Jacaranda cf. micrantha</i> Cham.		
Euphorbiaceae		
<i>Gymnanthes concolor</i> Spreng.		
Lauraceae		
<i>Nectandra megapota mica (Sprengel) Mez</i>		
Leguminosae - Mimosoideae		
<i>Inga marginata</i> Willd.		2p
Leguminosae (general)		
Meliaceae		
<i>Cabralea canjerana</i> Sald.		
<i>Trichilia clausenii</i> C. DC.		
Moraceae		
<i>Sorocea bonplandii</i> (Baill.) Burger		
Myrsinaceae		
<i>Myrsine</i> sp.		
Myrtaceae		
<i>Calyptanthus grandifolia</i> Berg.		2p
Rubiaceae		
<i>Psychothria leiocarpa</i> Cham. et Schlecht.		
Sapindaceae		
<i>Cupania vernalis</i> Camb.		
Tiliaceae		
<i>Luehea divaricata</i> Mart.		
Angiosperms - Monocotyledons		
Palmae (Arecaceae)		
<i>Euterpe edulis</i> Mart.		
3. Tree section		
Altitude (in m)		6
Angiosperms - Dicotyledons		
Annonaceae		
<i>Rollinia rugulosa</i> Schtdl.		

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Bignoniaceae			
<i>Tabebuia</i> sp.			
Euphorbiaceae			
<i>Gymnanthes concolor</i> Spreng.			
Flacourtiaceae			
<i>Banara tomentosa</i> Clos.			
Lauraceae			
<i>Ocotea pulchella</i> Mart.			
Leguminosae - Faboideae			
<i>Leguminosae (general)</i>			
<i>Myrocarpus frondosus</i> Fr. Allem.			
Meliaceae			
<i>Cabralea canjerana</i> Sald.			
<i>Guarea macrophylla</i> Vahl.			
<i>Trichilia clausenii</i> C. DC.			
Monimiaceae			
<i>Mollinedia cf. schottiana</i> (Spr.) Perk.			
Myrsinaceae			
<i>Myrsine</i> sp.			
Myrtaceae			
<i>Calyptanthus concinna</i> DC.			
<i>Campomanesia xanthocarpa</i> O. Berg			
<i>Eugenia schuechiana</i> O. Berg		2p	
<i>Myrcia oligantha</i> O. Berg		2p	
<i>Myrcianthes pungens</i> (Berg.) Legrand			
Rubiaceae			
<i>Rudgea parquioides</i> (Cham.) Müll. Arg.			
Rutaceae			
<i>Zanthoxylum cf. hyemale</i> A. St.-Hil.			
Sapindaceae			
<i>Allophylus edulis</i> (St. Hil.) Radlk.			
<i>Cupania vernalis</i> Camb.			

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Tiliaceae			
<i>Luehea divaricata</i> Mart.			
Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p		
Lianes			
<i>Arctolochia</i> sp.			
<i>Smilax</i> sp.			
<i>Dioscorea multiflora</i> Mart.			
Shrub section			
Altitude (in m)	3	3	3
Acanthaceae			
<i>Ruellia angustifolia</i> Sw.			2p
Euphorbiaceae			
<i>Pachystroma longifolium</i> (Ness) IM Johnst.			
<i>Endlicheria paniculata</i> (Spr.) Macbr.			
<i>Nectandra megapotamica</i> (Sprengel) Mez			
Leguminosae - Faboideae			
<i>Myrocarpus frondosus</i> Fr. Allem.			
Leguminosae - Mimosoideae			
<i>Inga marginata</i> Willd.	2p	2p	2p
Malvaceae			
<i>Bythneria australis</i> St. - Hil. (Malvaceae sens. lat.)			
Melastomataceae			
<i>Miconia cinerascens</i> Miq.	1r		
<i>Miconia hiemalis</i> St. Hil.	1r		
Meliaceae			
<i>Cabralea canjerana</i> Sald.			
<i>Guarea macrophylla</i> Vahl.			
<i>Trichilia clausenii</i> C. DC.			

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Myrtaceae		
<i>Calytranthès grandifolia</i> Berg	2p	
<i>Campomanesia xanthocarpa</i> (Mart.) O. Berg		
<i>Myrcianthes pungens</i> (Berg.) Legrand		
<i>Siphoneugenia reitzii</i> Legr.	2p	
Piperaceae		
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.		
<i>Piper cf. mikanianum</i> (Kunth) Steud.		2p
Rubiaceae		
<i>Couatara hexandra</i> (Jacq.) K. Schum.		
<i>Psychotria kleinii</i> Smith & Down		
<i>Rudgea parquiioides</i> (Cham.) Müll. Arg.		
Sapindaceae		
<i>Allophylus edulis</i> (St. Hil.) Radlk.		
<i>Cupania vernalis</i> Camb.		
Angiosperms - Monocotyledons		
Poaceae		
<i>Bambus</i> sp.		2p
Herb section		
Angiosperms - Dicotyledons		
Acanthaceae		
<i>Ruellia angustifolia</i> Sw.		
Leguminosae - Mimosoidae		
<i>Inga marginata</i> Willd.		
<i>Inga cf. marginata</i> Willd.	1p	
Melastomataceae		
<i>Leandra</i> sp.		
Meliaceae		
<i>Trichilia clausenii</i> C. DC.		
Monimiacae		
<i>Mollinedia elegans</i> Tul.		

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Oxalidaceae			
<i>Oxalis</i> sp.			1r
Passifloraceae			
<i>Passiflora</i> sp.			
Piperaceae			
<i>Piper</i> sp.			
Proteaceae			
<i>Roupala</i> sp.		1p	
Sapindaceae			
<i>Cupania vernalis</i> Camb.			
Urticaceae			
<i>Urtica baccifera</i> (L.) Gaud.			
Pteridophyta			
Aspleniaceae			
<i>Asplenium raddianum</i> Gaud.			
<i>Asplenium</i> sp.L.			1r
Blechnaceae			
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.			
Dennstaedtiaceae			
<i>Lastreopsis amplissima</i> (Presl) Tindale		5m	
Pteridaceae			
<i>Pteridophyta</i>			
Epiphytes			
Angiosperms - Dicotyledons			
Cactaceae			
<i>Rhipsalis</i> sp.			
Angiosperms - Monokotyledons			
Orchidiaceae			
<i>Oncidium hookeri</i> Rolfe.		p	
<i>Oncidium</i> sp.			
Orchidaceae			
<i>Pleurothallis hygrophila</i> Barb.Rodr.		p	

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Peridophyta			
<i>Antigramma brasiliensis</i> (Sw.) Moore			
<i>Microgramma</i> sp.	m		
Bryopsida			
moss	a		

Table III. 1: Alphabetic order of identified plants in semi-deciduous and deciduous forests (vegetation record according to Braun-Blanquet)

Zone 3

Specimen number	26	52	55	58	59	60	61	63
Date of record	18.7.02	21.12.02	23.1.04	23.1.04	24.1.04	24.1.04	25.1.04	25.1.04
1. Tree section								
Altitude (in m)	13	10	15		8	12	11	16
Angiosperms - Dicotyledons								
Asteraceae								
<i>Vernonia discolor</i> (Spreng.) Less					2p			
Myrtaceae								
<i>Eugenia multifcostata</i> Legr.	2p							
<i>Myrcia arborescens</i> Berg								
Myrtaceae								
Solanaceae								
Solanaceae (general)								
Gymnosperms								
Araucariaceae								
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	3p	2p	3p	3p	3a	3p	3p	4m
Podocarpaceae								
<i>Podocarpus lambertii</i> Klotzsch ex Endl.					2p			
2. Tree section								
Altitude (in m)	6	7	11			10	9	10
Angiosperms - Dicotyledons								
Anacardiaceae								
<i>Lithraea brasiliensis</i> (L.) March.								
Aquifoliaceae								
<i>Ilex brevicuspis</i> Reissek. Sin.						2p		
<i>Ilex cf. microdonta</i> Reissek								
<i>Ilex paraguayensis</i> St. Hil.					2p			
Bombacaceae								
Bombacaceae (geral)								

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Lauraceae									
<i>Nectandra megapotamica</i> (Sprengel) Mez									
<i>Ocotea pulchella</i> Mart.		2p		3p		2p			
Leguminosae - Mimosoidae									
<i>Leguminosae</i> (general)						2r			
Moraceae									
<i>Ficus organensis</i> (Miq.) Miq.				3r					
<i>Sorocea bonplandii</i> (Baill.) Burger		2p							
Myrsinaceae									
<i>Myrsine</i> sp.									
Myrtaceae									
<i>Calyptanthes grandifolia</i> Berg.		2p							
<i>Siphoneugenia reitzii</i> Legr.									
<i>Myrtaceae</i> (general)									
Sapindaceae									
<i>Cupania oblongifolia</i> Cambess.		2p							
<i>Cupania vernalis</i> Camb.		2p		3a					
<i>Matayba elaeagnoides</i> Radlk.								3p	
Styracaceae									
<i>Styrax cf. acuminatus</i> Pohl									
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze									
Podocarpaceae									
<i>Podocarpus lambertii</i> Klotzsch ex Endl.							2p		
3. Tree section									
Altitude (in m)		4	5	5		5	8	7	6
Angiosperms - Dicotyledons									
Annonaceae									
<i>Rollinia cf. sylvatica</i> (A. St.-Hil.) Mart.									2p
<i>Rollinia</i> sp.								2p	

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Thymelaeaceae									
<i>Daphnopsis fasciculata</i> (Meissner) Nevl.						2p			
<i>Daphnopsis racemosa</i> Gris.									3m
Winteraceae									
<i>Drymis brasiliensis</i> Miers		2p							
Angiosperms - Monocotyledons									
<i>Bambus</i> sp.									
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze						2p			2p
Podocarpaceae									
<i>Podocarpus lamberti</i> Klotzsch ex Endl.							2p		
Pteridophyta									
Dicksoniaceae									
<i>Dicksonia sellowiana</i> (Presl.) Hook.		2p							
<i>Alsophila</i> sp.									
Herb section									
Angiosperms - Dicotyledons									
Acanthaceae									
<i>Ruellia angustifolia</i> Sw.			1p						
Aquifoliaceae									
<i>Ilex cf. brevicuspis</i> Reissek. Sin.									
Asteraceae									
<i>Baccharis uncinella</i> DC.									
<i>Dasiphylum</i> sp.									
Euphorbiaceae									
<i>Sapium glandulatum</i> (Vell.) Pax								1r	
Leguminosae - Caesalpinioideae									
<i>Bauhinia</i> sp.							1r		
Leguminosae - Mimosoidae									
<i>Inga marginata</i> Willd.			1p						

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

		Zone 3				Zone 2				
Specimen number		64	65	73	76	5	33	39	68	70
Date of record		25.1.04	25.1.04	1.2.04	1.2.04	22.6.02	22.7.02	11.8.02	31.1.04	31.1.04
1. Tree section										
Altitude (in m)		16	16	10	9	12	12	11	12	12
Angiosperms - Dicotyledons										
Asteraceae										
<i>Vernonia discolor</i> (Spreng.) Less										
Myrtaceae										
<i>Eugenia multicostata</i> Legr.										
<i>Myrcia arborescens</i> Berg							3r			
Myrtaceae						2p				
Solanaceae										
<i>Solanaceae</i> (general)									2p	
Gymnosperms										
Araucariaceae										
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		4a	3a	4a	3a			3p	4m	3p
Podocarpaceae										
<i>Podocarpus lamberti</i> Klotzsch ex Endl.					3a					
2. Tree section										
Altitude (in m)		12	12	8		8	10	9	10	10
Angiosperms - Dicotyledons										
Anacardiaceae										
<i>Litorea brasiliensis</i> (L.) March.										
Aquifoliaceae										
<i>Ilex brevicuspis</i> Reissek. Sin.				3p						
<i>Ilex cf. microdonta</i> Reissek									2p	
<i>Ilex paraguayensis</i> St. Hil.										
Bombacaceae										
<i>Bombacaceae</i> (geral)								2p		

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Anacardiaceae									
<i>Schinus cf. lentiscifolius</i> L.		2p							
Annonaceae									
<i>Rollinia</i> sp.									2p
Aquifoliaceae									
<i>Ilex brevicuspis</i> Reissek. Sin.									
<i>Ilex paraguayensis</i> St. Hil.	2p	2p							
Asteraceae									
<i>Dasiphylum spinescens</i> (Less.) Cabr.						1r			
Bombacaceae									
Bombacaceae									
Celastraceae									
<i>Maytenus ilicifolia</i> Mart. Reiss									
Cunoniaceae									
<i>Weinmannia paulliniifolia</i> Pohl ex Ser.							2p		
Elaeocarpaceae									
<i>Sloanea monosperma</i> Vell.									
Euphorbiaceae									
<i>Sapum glandulatum</i> (Vell.) Pax									
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs									
Flacourtiaceae									
<i>Casearia decandra</i> Jacq.	2p	2p		3a					
<i>Casearia sylvestris</i> Sw.									2p
Icacinaceae									
<i>Citronella gongonha</i> (Mart.) Howard									
Lauraceae									
<i>Nectandra megaphylla</i> Hassl.	2p	2p							
<i>Nectandra megapotamica</i> (Sprengel) Mez									
<i>Nectandra</i> sp.									
Leguminosae - Caesalpinoideae									
<i>Bauhinia</i> sp.	1p								

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

<i>Myrciaria delicatula</i> (DC.) O. Berg									
<i>Psidium cattleianum</i> Sabine								2p	
<i>Siphoneugenia reitzii</i> Legr.				2p				2p	2p
Onagraceae									
<i>Fuchsia regia</i> (Vell.) Munz									
Piperaceae									
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.									
Proteaceae									
<i>Roupala brasiliensis</i> Klotzsch							1r		
Rosaceae									
<i>Prunus sellowii</i> Koehne									
Rubiaceae									
<i>Coutarea hexandra</i> (Jacq.) K. Schum.			2p						
<i>Psychotria kleinii</i> Smith & Down								2p	
<i>Psychotria</i> sp.				2p					
<i>Rudgea parquoides</i> (Cham.) Müll. Arg.				3a				2p	
Rutaceae									
<i>Citrus reticulata</i> (L.) Osbeck			1r						
<i>Zanthoxylum rhoifolium</i> Lam.									
<i>Zanthoxylum</i> sp.									
Salicaceae									
<i>Xylosma</i> sp. 1									
<i>Xylosma</i> sp. 2									1p
Sapindaceae									
<i>Allophylus edulis</i> (St. Hil.) Radlk.							2p		
<i>Cupania oblongifolia</i> Camb.									
<i>Cupania verralis</i> Camb.									
Solanaceae									
<i>Brunfelsia</i> sp.									

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Orchidiaceae																			
<i>Maxillaria picta</i> Hooker																			
<i>Oncidium concolor</i> Hooker																			
<i>Oncidium hookeri</i> Rolfe.																			
<i>Oncidium</i> sp.																			
<i>Pleurothallis grobyi</i> Bateman ex Lindl.																			
<i>Pleurothallis hygrophila</i> Barb. Rodr.																			
<i>Pleurothallis</i> cf. <i>linearifolia</i> Cogn.																			
<i>Pleurothallis trado</i>																			
Pteridophyta																			
<i>Microgramma</i> sp.																			
Bryopsida																			
moss																			

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

		Zone 2				Zone 1			
Specimen number		75	79	80	20	21	44		
Date of record		1.2.04	2.2.04	2.2.04	6.7.02	6.7.02	25.8.02		
1. Tree section									
Altitude (in m)		10	12	10	10	11	8		
Angiosperms - Dicotyledons									
Asteraceae									
<i>Vernonia discolor</i> (Spreng.) Less									
Myrtaceae									
<i>Eugenia multicosata</i> Legr.									
<i>Myrcia arborescens</i> Berg									
Myrtaceae									
Solanaceae									
<i>Solanaceae</i> (general)									
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		2p	4m	3a	2p	3p	2p		
Podocarpaceae									
<i>Podocarpus lambertii</i> Klotzsch ex Endl.									
2. Tree section									
Altitude (in m)		8	6	8	7	9	5		
Angiosperms - Dicotyledons									
Anacardiaceae									
<i>Lithraea brasiliensis</i> (L.) March.				2p					
Aquifoliaceae									
<i>Ilex brevicuspis</i> Reissek. Sin.									
<i>Ilex cf. microdonta</i> Reissek									
<i>Ilex paraguayensis</i> St. Hil.									
Bombacaceae									
<i>Bombacaceae</i> (geral)									

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Lauraceae							
<i>Nectandra megapotamica</i> (Sprengel) Mez							
<i>Ocotea pulchella</i> Mart.				3p			
Leguminosae - Mimosoidae							
<i>Leguminosae</i> (general)							
Moraceae							
<i>Ficus organensis</i> (Miq.) Miq.							
<i>Sorocea bonplandii</i> (Baill.) Burger							
Myrsinaceae							
<i>Myrsine</i> sp.							2p
Myrtaceae							
<i>Calytranthos grandifolia</i> Berg.							
<i>Siphoneugenia reitzii</i> Legr.						2p	
<i>Myrtaceae</i> (general)							2p
Sapindaceae							
<i>Cupania oblongifolia</i> Cambess.							
<i>Cupania vernalis</i> Camb.							
<i>Matayba elaeagnoides</i> Radlk.							
Styracaceae							
<i>Styrax cf. acuminatus</i> Pohl							
Gymnosperms							
Araucariaceae							
<i>Araucaria angustifolia</i> (Bertol.) Kuntze				2a			2p
Podocarpaceae							
<i>Podocarpus lambertii</i> Klotzsch ex Endl.					2p		2p
3. Tree section							
Altitude (in m)		6	4		6	5	
Angiosperms - Dicotyledons							
Annonaceae							
<i>Rollinia cf. sylvatica</i> (A. St.-Hil.) Mart.							
<i>Rollinia</i> sp.		2p					

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Aquifoliaceae									
<i>Ilex brevicuspis</i> Reissek. Sin.									
<i>Ilex paraguayensis</i> St. Hil.									
Asteraceae									
<i>Dasiphylum spinescens</i> (Less.) Cabr.									
Berberidaceae									
<i>Berberis laurina</i> Billb.				2p					
Bombacaceae									
Cunoniaceae									
<i>Weinmannia paulliniifolia</i> Pohl ex Ser.								1r	
Elaeocarpaceae									
<i>Sloanea monosperma</i> Vell.									
Euphorbiaceae									
<i>Gymnanthes concolor</i> Spreng.									
<i>Sapium glandulatum</i> (Vell.) Pax				2p					
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs									
<i>Stillingia oppositifolia</i> Baill.									
Flacourtiaceae									
<i>Banara tomentosa</i> Clos.									
<i>Casearia sylvestris</i> Sw.									
Lauraceae									
<i>Nectandra megapotamica</i> (Sprengel) Mez									
<i>Nectandra</i> sp.									
Melastomataceae									
<i>Miconia hiemalis</i> St. Hil.									
Meliaceae									
<i>Trichilia catigua</i> A. Juss.									
Monimiaceae									
<i>Mollinedia elegans</i> Tul.									
Moraceae									
<i>Sorocea bonplandii</i> (Baill.) Burger									

Table III.2: Alphabetic order of identified plants in Araucaria forests in vegetation record according to Braun-Blanquet

Myrsinaceae							
<i>Myrsine</i> sp.							
Myrtaceae							
<i>Acca sellowiana</i> (O. Berg) Burret			2p				
<i>Calyptanthus concinna</i> DC.							
<i>Eugenia uniflora</i> L.							
<i>Myrtaceae</i> (general)					3a		
<i>Siphonoeugenia reitzii</i> Legr.							
Phytolaccaceae							
<i>SeQUIERIA aculeata</i> L.							
Rubiaceae							
Sapindaceae							
<i>Cupania verralis</i> Camb.							
<i>Matayba elaeagnoides</i> Radlk.							
Symplocaceae							
<i>Symplocos uniflora</i> (Pohl) Benth.							
Thymelaeaceae							
<i>Daphnopsis fasciculata</i> (Meissner) Nevl.							
<i>Daphnopsis racemosa</i> Griseb.							
Winteraceae							
<i>Drymis brasiliensis</i> Miq.		2p					
Angiosperms - Monocotyledons							
<i>Bambusa</i> sp.							
Gymnosperms							
Araucariaceae							
<i>Araucaria angustifolia</i> (Bertol.) Kuntze							
Podocarpaceae							
<i>Podocarpus lamBERTII</i> Klotzsch ex Endl.							
Shrub section							
Altitude (in m)		2	2,5				3
Acanthaceae							
<i>Ruellia angustifolia</i> Sw.							

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Anacardiaceae									
<i>Schinus cf. lentiscifolius</i> L.									
Annonaceae									
<i>Rollinia</i> sp.									
Aquifoliaceae									
<i>Ilex brevicuspis</i> Reissek. Sin.									
<i>Ilex paraguariensis</i> St. Hil.									
Asteraceae									
<i>Dasyphyllum spinescens</i> (Less.) Cabr.									
Bombacaceae									
Bombacaceae									
Celastraceae									
<i>Maytenus ilicifolia</i> Mart. Reiss			1r						
Cunoniaceae									
<i>Weinmannia paulliniifolia</i> Pohl ex Ser.									
Elaeocarpaceae									
<i>Sloanea monosperma</i> Vell.									
Euphorbiaceae									
<i>Sapium glandulatum</i> (Vell.) Pax				2p					
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs									
Flacourtiaceae									
<i>Casearia decandra</i> Jacq.					3a				
<i>Casearia sylvestris</i> Sw.									
Icacinaceae									
<i>Citronella gongonha</i> (Mart.) Howard					2p				
Lauraceae									
<i>Nectandra megaphylla</i> Hassl.									
<i>Nectandra megapotamica</i> (Sprengel) Mez									
<i>Nectandra</i> sp.									1p
Leguminosae - Caesalpinioideae									
<i>Bauhinia</i> sp.									

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Leguminosae - Mimosoidae									
<i>Mimosa scabrella</i> Benth.									
Leguminosae (general)									
Melastomataceae									
<i>Leandra cf regnellii</i> Cogn.									
<i>Leandra</i> sp.	2p				1p		1p		
<i>Miconia cinerascens</i> Miq.	2p				2p			2p	
<i>Miconia hiemalis</i> St. Hil.	2p	2m			2p			2p	
Meliaceae									
<i>Cabralea canjerana</i> Sald.									
Monimiaceae									
<i>Mollinedia elegans</i> Tul.									
Myrsinaceae									
<i>Myrsine</i> sp.									
Myrtaceae									
<i>Acca sellowiana</i> (O. Berg) Burret									
<i>Eugenia pyriformis</i> Cambess.									
<i>Eugenia ramboi</i> D. Legrand									
<i>Eugenia uniflora</i> L.									
<i>Calyptanthes concinna</i> DC.									
<i>Campomanesia xanthocarpa</i> (Mart.) O. Berg									
<i>Eugenia psidiifolia</i>									
<i>Eugenia multicosata</i> D. Legrand									
<i>Myrcougenia glaucescens</i> Cam.) D. Legrand et Kausel									
<i>Myrcougenia miersiana</i> (Gardner) D. Legrand et Kausel									
<i>Myrcougenia mesomischia</i> (Burret) D. Legrand et Kausel									
<i>Myrcougenia myrcioides</i> (Cambess.) Berg.									
<i>Myrcougenia oxypala</i> (Burret) D. Legrand et Kausel									
<i>Myrcia guianensis</i> (Aubl.) DC. = <i>M. obtecta</i> (Berg) Kiaerskov									
<i>Myrcianthes pungens</i> (Berg.) Legrand									

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

<i>Myrciaria delicatula</i> (DC.) O. Berg									
<i>Psidium cattleianum</i> Sabine									
<i>Siphoneugenia reitzii</i> Legr.									
Onagraceae									
<i>Fuchsia regia</i> (Vell.) Munz				2p					
Piperaceae									
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.									
Proteaceae									
<i>Roupala brasiliensis</i> Klotzsch									
Rosaceae									
<i>Prunus sellowii</i> Koehne									
Rubiaceae									
<i>Coutarea hexandra</i> (Jacq.) K. Schum.									
<i>Psychotria kleinii</i> Smith & Down									
<i>Psychotria</i> sp.									
<i>Rudgea parquiioides</i> (Cham.) Müll. Arg.									
Rutaceae									
<i>Citrus reticulata</i> (L.) Osbeck									
<i>Zanthoxylum rhoifolium</i> Lam.									
<i>Zanthoxylum</i> sp.				1r					
Salicaceae									
<i>Xylosma</i> sp. 1								1r	
<i>Xylosma</i> sp. 2									
Sapindaceae									
<i>Allophylus edulis</i> (St. Hil.) Radlk.									
<i>Cupania oblongifolia</i> Camb.									
<i>Cupania vernalis</i> Camb.									
Solanaceae									
<i>Brunfelsia</i> sp.									

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Thymelaeaceae								
<i>Daphnopsis fasciculata</i> (Meissner) Nevl.		2p						
<i>Daphnopsis racemosa</i> Gris.		2m	3a					
Winteraceae								
<i>Drymis brasiliensis</i> Miers	2p	3p						
Angiosperms - Monocotyledons								
<i>Bambus</i> sp.								
Gymnosperms								
Araucariaceae								
<i>Araucaria angustifolia</i> (Bertol.) Kuntze								
Podocarpaceae								
<i>Podocarpus lamberti</i> Klotzsch ex Endl.								
Pteridophyta								
Dicksoniaceae								
<i>Dicksonia sellowiana</i> (Presl.) Hook.	2p					1p		
<i>Alsophila</i> sp.								
Herb section								
Angiosperms - Dicotyledons								
Acanthaceae								
<i>Ruellia angustifolia</i> Sw.								
Aquifoliaceae								
<i>Ilex cf. brevicuspis</i> Reissek. Sin.								
Asteraceae								
<i>Baccharis uncinella</i> DC.								1p
<i>Dasiphylum</i> sp.	1r							
Euphorbiaceae								
<i>Sapium glandulatum</i> (Vell.) Pax		1p						
Leguminosae - Caesalpinioideae								
<i>Bauhinia</i> sp.								
Leguminosae - Mimosoidae								
<i>Inga marginata</i> Willd.								

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Melastomataceae									
<i>Leandra</i> sp.									
<i>Miconia cinerascens</i> Miq.									2p
<i>Miconia hiemalis</i> St. Hil.									2p
Monimiaceae									
<i>Mollinedia elegans</i> Tul.									
Myrsinaceae									
<i>Myrsine</i> sp.							1p		
Myrtaceae									
<i>Myrciaria delicatula</i> (DC.) O. Berg									
Passifloraceae									
<i>Passiflora</i> sp.									
Phytolaccaceae									
<i>Seguieria aculeata</i> L.									
Plantagaceae									
<i>Plantago</i> sp.				2m					
Polygonaceae									
<i>Polygonum</i> sp.									
Piperaceae									
<i>Piper</i> sp.									
Rubiaceae									
<i>Coutarea hexandra</i> (Jacq.) K. Schum.									
Sapindaceae									
<i>Allophylus edulis</i> (St. Hil.) Radlk.									
<i>Cupania vernalis</i> Camb.									
<i>Matayba elaeagnoides</i> Radlk.									
Solanaceae									
<i>Brunfelsia</i> sp.									
<i>Solanum montegazzianum</i> Somm. et Lev.									
Thymeleaceae									
<i>Daphnopsis racemosa</i> Gris.									
Winteraceae									
<i>Drymis brasiliensis</i> Miers.						1p			

Table III. 2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Angiosperms - Monokotyledons								
Poaceae								
<i>Agrostis</i> sp.								
Poaceae (general)	3m	3m	5m					
Cyperaceae								
<i>Cyperus</i> sp.								
<i>Rhynchospora</i> sp.					4a			
Gymnosperme								
Araucariaceae								
<i>Araucaria angustifolia</i> (Bertol.) Kuntze								
Pinaceae								
<i>Pinus</i> sp. L.		2a						
Pteridophyta								
Blechnaceae								
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.								
Gleicheniaceae								
<i>Gleichenia</i> sp.								
<i>Pteridophyta</i>								
Epiphytes								
Angiosperms - Dicotyledons								
Cactaceae								
<i>Rhipsalis</i> sp.								
Loranthaceae - parasite								
Angiosperms - Monokotyledons								
Bromeliaceae								
<i>Bromelia</i> sp.					a			
<i>Tillandsia usneoides</i> (L.) L.		p	p				p	
<i>Tillandsia</i> with red blossom								
<i>Vriesea platynema</i> Gaud.		p						

Table III.2: Alphabetic order of identified plants in Araucaria forests (vegetation record according to Braun-Blanquet)

Orchidiaceae									
<i>Maxillaria picta</i> Hooker									
<i>Oncidium concolor</i> Hooker									p
<i>Oncidium hookeri</i> Rolfe.									
<i>Oncidium</i> sp.									
<i>Pleurothallis grobyi</i> Bateman ex Lindl.									
<i>Pleurothallis hygrophila</i> Barb.Rodr.									
<i>Pleurothallis cf. linearifolia</i> Cogn.									
<i>Pleurothallis triado</i>									
Pteridophyta									
<i>Microgramma</i> sp.									a
Bryopsida									
moss								a	

Table III.2: Alphabetic order of identified plants in Araucaria forests in Araucaria forests (vegetation record according to Braun-Blanquet)

Specimen number	Zone 3				Zone 2			
	27	28	57	74	6	13	40	49
Date of record	18.7.02	20.7.02	23.1.04	1.2.04	22.6.02	23.6.02	11.8.02	21.12.02
1. Tree section								
Altitude (in m)	10	12	12	9	13	12	10	10
Angiosperms - Dicotyledons								
Asteraceae								
<i>Vernonia discolor</i> (Spreng.) Less								
Myrtaceae								
<i>Myrtaceae</i>	3a				2a			
Gymnosperms								
Araucariaceae								
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p	3p	3p	3a	3p	3a	2p	2p
2. Tree section								
Altitude (in m)	6	9	9	7	10	8	8	7
Angiosperms - Dicotyledons								
Anacardiaceae								
<i>Lithraea brasiliensis</i> (L.) March.								
Aquifoliaceae								
<i>Ilex brevicuspis</i> Reissek. Sin.								
Asteraceae								
<i>Dasyphyllum spinescens</i> (Less.) Cabr.								
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.	1r							
<i>Vernonia discolor</i> (Spreng.) Less								
Bombacaceae								
Euphorbiaceae								
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs								2p
Lauraceae								
<i>Ocotea pulchella</i> Mart.								

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Asteraceae																			
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.		2p																	
Bombacaceae																			
Bombacaceae (general)																			
Euphorbiaceae																			
<i>Stillingia oppositifolia</i> Baill.					2p														
Flacourtiaceae																			
<i>Casearia decandra</i> Jacq.						2p													
Melastomataceae																			
<i>Miconia cinerascens</i> Miq.		2p																	
<i>Miconia hiemalis</i> St. Hil.		2p																	
Myrtaceae																			
<i>Acacia sellowiana</i> (O. Berg) Burret																			
<i>Calyptanthus concinna</i> DC.				2p															
<i>Campomanesia xanthocarpa</i> O. Berg									2p										
<i>Eugenia neomyrtifolia</i> Sobral																			2a
<i>Hexachlamys etataiensis</i> Mattos									2p										
<i>Myrceugenia alpigena</i> (DC.) Landrum																			2p
<i>Myrrhinium atropurpureum</i> Schott																			2p
Myrtaceae (general)																			
<i>Siphoneugenia reitzii</i> Legr.																			2p
Rubiaceae																			
<i>Rudgea parquiioides</i> (Cham.) Müll. Arg.				2p															
Rutaceae																			
<i>Zanthoxylum</i> sp.																			
Sapindaceae																			
<i>Allophylus edulis</i> (St. Hil.) Radlk.																			
Solanaceae																			
<i>Solanum namulosum</i> Sendtner											2p								
Thymelaeaceae																			
<i>Daphnopsis racemosa</i> Gris.																			2a
Winteraceae																			
<i>Drymis brasiliensis</i> Miars.																			2p

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Leguminosae - Mimosoidae									
<i>Mimosa scabrella</i> Benth.									
Melastomataceae									
<i>Leandra cf regnellii</i>									
<i>Leandra sublanata</i> Cogn.	2p								
<i>Leandra</i> sp.									
<i>Miconia cinerascens</i> Miq.	2p					2a	2p	2p	2p
<i>Miconia hiemalis</i> St. Hil.						2p		2p	2p
Myrsinaceae									
<i>Myrsine coriacea</i> (Sw.) R. Br.									
<i>Myrsine lorenzianum</i>									
Myrtaceae									
<i>Acca sellowiana</i> (O. Berg) Burret									
<i>Calyptanthus concinna</i> DC.		2p							2p
<i>Eugenia pyriformis</i> Cambess.		2p							
<i>Eugenia cf. oeidocarpa</i> O.Berg									
<i>Eugenia uruguayensis</i> Cambess.									
<i>Hexachlamys etataiensis</i> Mattos				2p					
<i>Myrceugenia alpigena</i> DC.) Landrum									
<i>Myrceugenia mesomischia</i> (Burret) D.Legrand et Kausel					2p				
<i>Myrceugenia miersiana</i> (Gardner) D. Legrand et Kausel									
<i>Myrceugenia myrcioides</i> (Cambess.) Berg.	2p								
<i>Myrceugenia ovata</i> (Hook. & Arn.) O. Berg									
Myrtaceae (general)								3a	
<i>Siphoneugenia reitzii</i> Legr.									
Onagraceae									
<i>Fuchsia regia</i> (Vell.) Munz									
Piperaceae									
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.	2a								
Rhamnaceae									
<i>Rhamnus sphaerosperma</i> Sw.					2p				

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Specimen number	Zone 2		Zone 1					
	67	77	17	18	23	36	46	47
Date of record	31.1.04	1.2.04	25.6.02	6.7.02	7.7.02	10.8.02	25.8.02	21.12.02
1. Tree section								
Altitude (in m)	10	12	11	10	8	12	10	9
Angiosperms - Dicotyledons								
Asteraceae								
<i>Vernonia discolor</i> (Spreng.) Less	2p							
Myrtaceae								
<i>Myrtaceae</i>				3a				
Gymnosperms								
Araucariaceae								
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	3p	3p	2p	2p	2p	2p	2p	2p
2. Tree section								
Altitude (in m)	8	8	7	7	6	8	8	6
Angiosperms - Dicotyledons								
Anacardiaceae								
<i>Lithraea brasiliensis</i> (L.) March.		3p						
Aquifoliaceae								
<i>Ilex brevicuspis</i> Reissek. Sin.								
Asteraceae								
<i>Dasyphyllum spinescens</i> (Less.) Cabr.			2p					
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.								
<i>Vernonia discolor</i> (Spreng.) Less				2p				
Bombacaceae								
Euphorbiaceae							2p	
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs								
Lauraceae								
<i>Ocotea pulchella</i> Mart.								

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Asteraceae									
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.									
Bombacaceae									
Bombacaceae (general)					1r				
Euphorbiaceae									
<i>Stillingia oppositifolia</i> Baill.									
Flacourtiaceae									
<i>Casearia decandra</i> Jacq.									
Melastomataceae									
<i>Miconia cinerascens</i> Miq.									
<i>Miconia hiemalis</i> St. Hil.									
Myrtaceae									
<i>Acacia sellowiana</i> (O. Berg) Burret									
<i>Calytranthus concinna</i> DC.									
<i>Campomanesia xanthocarpa</i> O. Berg									
<i>Eugenia neomyrtifolia</i> Sobral									
<i>Hexachlamys etataiensis</i> Mattos									
<i>Myrceugenia alpigena</i> (DC.) Landrum	2p								
<i>Myrrhinium atropurpureum</i> Schott									
Myrtaceae (general)									
<i>Siphoneugenia reitzii</i> Legr.			2p						
Rubiaceae									
<i>Rudgea parquiioides</i> (Cham.) Müll. Arg.									
Rutaceae									
<i>Zanthoxylum</i> sp.									
Sapindaceae									
<i>Allophylus edulis</i> (St. Hil.) Radlk.									
Solanaceae									
<i>Solanum namulosum</i> Sendtner									
Thymeleaceae									
<i>Daphnopsis racemosa</i> Gris.					2p				
Winteraceae									
<i>Drymis brasiliensis</i> Miers.			2r						2p

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Leguminosae - Mimosoidae																			
<i>Mimosa scabrella</i> Benth.									1p										
Melastomataceae																			
<i>Leandra cf regnellii</i>		2p																	
<i>Leandra sublanata</i> Cogn.											2p								
<i>Leandra</i> sp.										2p									
<i>Miconia cinerascens</i> Miq.			3a							2p									
<i>Miconia hiemalis</i> St. Hil.			3a	1p						2p	2p								2p
Myrsinaceae																			
<i>Myrsine coriacea</i> (Sw.) R. Br.		1p																	
<i>Myrsine lorenzianum</i>											1p								
Myrtaceae																			
<i>Acca sellowiana</i> (O. Berg) Burret																			2p
<i>Calyptanthes concinna</i> DC.		2p																	
<i>Eugenia pyriformis</i> Cambess.																			
<i>Eugenia cf. oeidocarpa</i> O.Berg		2p																	
<i>Eugenia uruguayensis</i> Cambess.									1p										
<i>Hexachlamys etataiensis</i> Mattos																			
<i>Myrceugenia alpigena</i> DC.) Landrum		2p																	
<i>Myrceugenia mesomischia</i> (Burret) D.Legrand et Kausel																			
<i>Myrceugenia miersiana</i> (Gardner) D. Legrand et Kausel		2p																	
<i>Myrceugenia myrcioides</i> (Cambess.) Berg.																			
<i>Myrceugenia ovata</i> (Hook. & Arn.) O. Berg		2p																	2p
<i>Myrtaceae</i> (general)				3a						3a									
<i>Siphoneugenia reitzii</i> Legr.		2p		3p						2p									2p
Onagraceae																			
<i>Fuchsia regia</i> (Vell.) Munz																			
Piperaceae																			
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.																			
Rhamnaceae																			
<i>Rhamnus sphaerosperma</i> Sw.																			

Table III. 3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Thymelaeaceae																				
<i>Daphnopsis racemosa</i> Gris.																				
Urticaceae																				
<i>Urea baccifera</i> (L.) Gaud.																				
Commelinaceae																				
<i>Commelina</i> sp.						4m										4m				
Poaceae																				
<i>Poaceae</i> (general)																				
Gymnosperme																				
Araucariaceae																				
<i>Araucaria angustifolia</i> (Bertol.) Kuntze										2p										
Pinaceae																				
Podocarpaceae																				
<i>Podocarpus lamBERTii</i> Klotzsch ex Endl.										2p										
Pteridophyta																				
Blechnaceae																				
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.										2a										
Epiphytes																				
Angiosperms - Dicotyledons																				
Cactaceae																				
<i>Rhipsalis</i> sp.																				
Angiosperms - Monokotyledons																				
Bromeliaceae																				
<i>Bromelia</i> sp.																				
<i>Tillandsia usneoides</i> (L.) L.										p						m				
<i>Tillandsia</i> with red blossom																				
<i>Vriesea platyneura</i> Gaud.										p										
Orchidiaceae																				
<i>Bulbophyllum</i> cf. <i>glutinatum</i> (Barb. Rodr.) Cogn.																				
<i>Maxillaria picta</i> Hooker																				
<i>Ocymella</i> sp.																				
<i>Oncidium concolor</i> Hooker																				
<i>Pleurothallis hygrophila</i> Barb. Rodr.																				
<i>Pleurothallis sonderana</i> Reichenbach																				

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Specimen number	83	84	85
Date of record	28.2.04	28.2.04	29.2.04
1. Tree section			
Altitude (in m)	12	10	14
Angiosperms - Dicotyledons			
Asteraceae			
<i>Vernonia discolor</i> (Spreng.) Less			
Myrtaceae			
Myrtaceae			
Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	3p	2p	3a
2. Tree section			
Altitude (in m)	9	7	8
Angiosperms - Dicotyledons			
Anacardiaceae			
<i>Lithraea brasiliensis</i> (L.) March.			
Aquifoliaceae			
<i>Ilex brevicuspis</i> Reissek. Sin.		2r	
Asteraceae			
<i>Dasyphyllum spinescens</i> (Less.) Cabr.			
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.			
<i>Vernonia discolor</i> (Spreng.) Less			
Bombacaceae			
Euphorbiaceae			
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs			
Lauraceae			
<i>Ocotea pulchella</i> Mart.			3p

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Myrsinaceae				
<i>Myrsine umbellata</i> Mart.				
Myrtaceae				
<i>Acca sellowiana</i> (O. Berg) Burret				
<i>Campomanesia aurea</i> O. Berg.				
<i>Campomanesia xanthocarpa</i> O. Berg				3a
<i>Calyptanthes concinna</i> DC.				
<i>Eugenia multicostata</i> D. Legrand		2p		
<i>Myrceugenia oxypala</i> (Burret) D. Legrand et Kausel				
<i>Myrcia arborescens</i> Berg		3r		
<i>Siphoneugenia reitzii</i> Legr.				
Myrtaceae (general)				3a
Rhamnaceae				
<i>Rhamnus sphaerosperma</i> Sw.				
Thymeleaceae				
<i>Daphnopsis racemosa</i> Gris.				
Winteraceae				
<i>Drymis brasiliensis</i> Miers.				
Gymnosperms				
Araucariaceae				
<i>Araucaria angustifolia</i> (Bertol.) Kuntze			2p	
Podocarpaceae				
<i>Podocarpus lambertii</i> Klotzsch ex Endl.			2p	
3. Tree section				
Altitude (in m)		4	5	6
Angiosperms - Dicotyledons				
Annonaceae				
<i>Rollinia rugulosa</i> Schtdl.				
<i>Rollinia</i> cf. <i>rugulosa</i> Schtdl.				
Aquifoliaceae				
<i>Ilex paraguariensis</i> St. Hil.				

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Asteraceae				
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.				
Bombacaceae				
Bombacaceae (general)				
Euphorbiaceae				
<i>Stillingia oppositifolia</i> Baill.				
Flacourtiaceae				
<i>Casearia decandra</i> Jacq.				
Melastomataceae				
<i>Miconia cinerascens</i> Miq.				
<i>Miconia hiemalis</i> St. Hil.				
Myrtaceae				
<i>Acca sellowiana</i> (O. Berg) Burret				2p
<i>Calyptanthus concinna</i> DC.				
<i>Campomanesia xanthocarpa</i> O. Berg				
<i>Eugenia neomyrtifolia</i> Sobral				
<i>Hexachlamys etataiensis</i> Mattos				
<i>Myrcogenia alpigena</i> (DC.) Landrum				
<i>Myrrhinium atropurpureum</i> Schott				
Myrtaceae (general)	2a			
<i>Siphoneugenia reitzii</i> Legr.				
Rubiaceae				
<i>Rudgea parquiioides</i> (Cham.) Müll. Arg.				
Rutaceae				
<i>Zanthoxylum</i> sp.			1p	
Sapindaceae				
<i>Allophylus edulis</i> (St. Hil.) Radlk.			2p	
Solanaceae				
<i>Solanum namulosum</i> Sendtner				
Thymeleaceae				
<i>Daphnopsis racemosa</i> Gris.				
Winteraceae				
<i>Drymis brasiliensis</i> Miars.			2a	

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	1p	2p	
Podocarpaceae			
<i>Podocarpus lambertii</i> Klotzsch ex Endl.		2p	
Shrub section			
Altitude (in m)			
Anacardiaceae			
<i>Lithraea brasiliensis</i> (L.) March.			
Annonaceae			
<i>Rollinia cf. rugulosa</i> Schtdl.			
<i>Rollinia</i> sp.			
Aquifoliaceae			
<i>Ilex brevicuspis</i> Reissek. Sin.			
<i>Ilex paraguayensis</i> St. Hll.			
Asteraceae			
<i>Dasyphyllum spinescens</i> (Less.) Cabr.			
Berberidaceae			
<i>Berberis laurina</i> Billb.		2p	
Celastraceae			
<i>Maytenus ilicifolia</i> Mart. Reiss		1r	
Euphorbiaceae			
<i>Sapium glandulatum</i> (Vell.) Pax			
<i>Stillingia oppositifolia</i>			
Flacourtiaceae			
<i>Casearia decandra</i> Jacq.			
<i>Casearia sylvestris</i> Sw.			
Icacinaceae			
<i>Citronella gongonha</i> (Mart.) Howard		1r	
Lauraceae			
<i>Nectandra</i> sp.			1p

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Leguminosae - Mimosoidae			
<i>Mimosa scabrella</i> Benth.			
Melastomataceae			
<i>Leandra cf regnellii</i>			
<i>Leandra sublanata</i> Cogn.			
<i>Leandra</i> sp.			
<i>Miconia cinerascens</i> Miq.		1p	
<i>Miconia hiemalis</i> St. Hil.	2p	1p	
Myrsinaceae			
<i>Myrsine coriacea</i> (Sw.) R. Br.			
<i>Myrsine lorenzianum</i>			
Myrtaceae			
<i>Acca sellowiana</i> (O. Berg) Burret		1r	
<i>Calyptanthes concinna</i> DC.			
<i>Eugenia pyriformis</i> Cambess.			
<i>Eugenia cf. oeidocarpa</i> O.Berg			
<i>Eugenia uruguayensis</i> Cambess.			
<i>Hexachlamys etataiensis</i> Mattos			
<i>Myrceugenia alpigena</i> DC.) Landrum			
<i>Myrceugenia mesomischia</i> (Burret) D.Legrand et Kausel			
<i>Myrceugenia miersiana</i> (Gardner) D. Legrand et Kausel			
<i>Myrceugenia myrcioides</i> (Cambess.) Berg.			
<i>Myrceugenia ovata</i> (Hook. & Arn.) O. Berg			
Myrtaceae (general)			
<i>Siphoneugenia reitzii</i> Legr.			
Onagraceae			
<i>Fuchsia regia</i> (Vell.) Munz		1p	
Piperaceae			
<i>Piper gaudichaudianum</i> (Kunth) Kunth ex C. DC.			
Rhamnaceae			
<i>Rhamnus sphaerosperma</i> Sw.			

Table III. 3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Rubiaceae				
<i>Rudgea parquioides</i> (Cham.) Müll. Arg.				
Symplocaceae				
<i>Symplocos uniflora</i> (Pohl) Benth.				
Thymeleaceae				
<i>Daphnopsis racemosa</i> Gris.	3a	2p		
Winteraceae				
<i>Drymis brasiliensis</i> Miers		2p		
Gymnosperms				
Araucariaceae				
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	1p	2p	1p	
Podocarpaceae				
<i>Podocarpus lambertii</i> Klotzsch ex Endl.		3a		
Pteridophyta				
Dicksoniaceae				
<i>Dicksonia sellowiana</i> (Presl.) Hook.				
Herb section				
Angiosperms - Dicotyledons				
Annonaceae				
<i>Rollinia cf. rugulosa</i> Schitdl.				
Apiaceae				
<i>Eryngium horridum</i> Malme				
Asteraceae				
<i>Conyza notobellidiastrum</i> Griseb.				
Melastomataceae				
<i>Leandra</i> sp.				
<i>Miconia cinerascens</i> Miq.		1p		
<i>Miconia hiemalis</i> St. Hil.		1p		
Solanaceae				
<i>Solanum variabile</i> Mart.				
<i>Solanum montegazzianum</i> Somm.et Lev.				
<i>Solanum namulosum</i> Sendtner				
<i>Solanum</i> sp.				

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Thymelaeaceae			
<i>Daphnopsis racemosa</i> Gris.		1a	
Urticaceae			
<i>Urea baccifera</i> (L.) Gaud.			
Commelinaceae			
<i>Commelina</i> sp.			
Poaceae			
<i>Poaceae</i> (general)	5m	5m	
Gymnosperme			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze			
Pinaceae			
Podocarpaceae			
<i>Podocarpus lamberti</i> Klotzsch ex Endl.			
Pteridophyta			
Blechnaceae			
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.		2a	
Epiphytes			
Angiosperms - Dicotyledons			
Cactaceae			
<i>Rhipsalis</i> sp.		p	
Angiosperms - Monokotyledons			
Bromeliaceae			
<i>Bromelia</i> sp.			
<i>Tillandsia usneoides</i> (L.) L.	m	m	p
<i>Tillandsia</i> with red blossom			
<i>Vriesea platynema</i> Gaud.			a
Orchidiaceae			
<i>Bulbophyllum</i> cf. <i>glutinosum</i> (Barb. Rodr.) Cogn.			
<i>Maxillaria picta</i> Hooker			
<i>Ocymella</i> sp.			
<i>Oncidium concolor</i> Hooker			
<i>Pleurothallis hygrophila</i> Barb. Rodr.			
<i>Pleurothallis sonderana</i> Reichenbach			

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

Pteridophyta				
<i>Microgramm</i> a sp.				
Bryopsida				
moss			a	
lichens				

Table III.3: Alphabetic order of identified plants in Capões (vegetation record according to Braun-Blanquet)

	Zone 3	Zone 2	Zone 1
Specimen number	50	9	82
Date of record	21.12.02	23.6.02	31.1.04
1. Tree section			
Altitude (in m)	8	11	15
Angiosperms - Dicotyledons			
Euphorbiaceae			
<i>Sebastiania commersoniana</i> (Ball.) Smith & Downs	2p		
Myrtaceae			
<i>Calyptanthus concinna</i> DC.	2p		
<i>Myrtaceae</i>	3p	3a	
Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		4a	2p
2. Tree section			
Altitude (in m)	6	7	
Angiosperms - Dicotyledons			
Lauraceae			
<i>Ocotea pulchella</i> Mart.			3p
Myrtaceae			
<i>Calyptanthus concinna</i> DC.	2p		
<i>Siphoneugenia reitzii</i> Legr.		2p	
3. Tree section			
Altitude (in m)	5		8
Angiosperms - Dicotyledons			
Myrtaceae			
<i>Myrcia lajeana</i> D. Legrand	2p		

Table III.4: Alphabetic order of identified plants in gallery forest (vegetation record according to Braun-Blanquet)

Gymnosperms					
Podocarpaceae					
<i>Podocarpus lambertii</i> Klotzsch ex Endl.					2p
Shrub section					
Altitude (in m)		2	2	2,5	2,5
Aquifoliaceae					
<i>Ilex</i> sp.		2p			
Asteraceae					
<i>Eupatorium serratum</i>				4m	
Euphorbiaceae					
<i>Sebastiania</i> cf. <i>brasiliensis</i> Spreng.				2p	
Melastomataceae					
<i>Leandra</i> sp.			1p		
<i>Miconia cinerascens</i> Miq.		2p	1p		3a
<i>Miconia hiemalis</i> St. Hil.					3a
Myrtaceae					
<i>Eugenia schuechiana</i> O. Berg			2p		
<i>Eugenia</i> sp.			2p		
<i>Myrcia richardiana</i> (O. Berg) Kiaersk.			2p		
<i>Siphoneugenia reitzii</i> Legr.			2p		2a
Sapindaceae					
<i>Allophylus guaraniticus</i> (St. Hil.) Radlk.				2a	
Thymeleaceae					
<i>Daphnopsis racemosa</i> Gris.					2p
Winteraceae					
<i>Drymis brasiliensis</i> Miers		2p			
Gymnosperms					
Podocarpaceae					
<i>Podocarpus lambertii</i> Klotzsch ex Endl.					1p
Herb section					
Angiosperms - Dicotyledons					
Amaranthaceae					
<i>Iresine diffusa</i> Humb. & Bonpl. ex Willd.		1p			

Table III.4: Alphabetic order of identified plants in gallery forest (vegetation record according to Braun-Blanquet)

Apiaceae					
<i>Eryngium horridum</i> Malme				3m	
Malvaceae					
<i>Sida rhombifolia</i> L.	1r				
<i>Sida spinosa</i> L.	1p				
Melastomataceae					
<i>Miconia hiemalis</i> St. Hil.					2a
Polygonaceae					
<i>Polygonum hydropper</i> L.	2p				
Rubiaceae					
<i>Relbunium cf. gracilium</i>		1r			
Sapindaceae					
<i>Allophylus guaraniticus</i> (St. Hil.) Radlk.			1p		
<i>Solanum</i> sp.	2p				
Verbenaceae					
<i>Verbena</i> cf. <i>littoralis</i> Kunth.	1r				
Angiosperms - Monokotyledons					
Poaceae					
<i>Paspalum purnilum</i> Nees	2a				
Poaceae (general)					2p
Cyperaceae					
<i>Cyperus</i> sp.	1r				
Gymnosperme					
Araucariaceae					
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		1p			
Podocarpaceae					
<i>Podocarpus lamberti</i> Klotzsch ex Endl.					1p
Pteridophyta					
Dennstaedtiaceae					
<i>Pteridium aquilinum</i> (L.) Kuhn.		1p			

Table III. 4: Alphabetic order of identified plants in gallery forest (vegetation record according to Braun-Blanquet)

Epiphytes					
Angiosperms - Dicotyledons					
Cactaceae					
<i>Rhipsalis</i> sp.	p				
Angiosperms - Monokotyledons					
Bromeliaceae					
<i>Tillandsia usneoides</i> (L.) L.	a	m			m
Pteridophyta					
<i>Microgramma</i> sp.			m		
Bryopsida					
moss					m

Table III.4: Alphabetic order of identified plants in gallery forest (vegetation record according to Braun-Blanquet)

	Zone 3	Zone 2	Zone 1	
Specimen number	51	8	66	43
Date of record	21.12.02	22.6.02	25.1.04	25.8.02
3. Tree section				
Altitude (in m)		4		
Angiosperms - Dicotyledons				
Gymnosperms				
Araucariaceae				
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		2r		
Shrub section				
Altitude (in m)	2,5	2	1,5	2,5
Anacardiaceae				
<i>Schinus polygamus</i> (Cav.) Cabrera.	2r			
Asteraceae				
<i>Baccharis uncinella</i> DC.	3a	4a	3m	4a
<i>Eupatorium serratum</i>			3m	
Berberidaceae				
<i>Berberis laurina</i> Billb.	2p	1p		
Erythroxylaceae				
<i>Erythroxylum</i> sp.			2p	
Melastomataceae				
<i>Miconia cinerascens</i> Miq.		1p		
Myrtaceae				
<i>Psidium cattleianum</i> Sabine				2p
Rubiaceae				
<i>Psychotria</i> sp.				2p
Thymeleaceae				
<i>Daphnopsis racemosa</i> Gris.	2p			

Table III. 5: Alphabetic order of identified plants in Capoeira vegetation (vegetation record according to Braun-Blanquet)

Gymnosperms					
Araucariaceae					
<i>Araucaria angustifolia</i> (Bertol.) Kuntze		1r		1r	
Herb section					
Angiosperms - Dicotyledons					
Apiaceae					2a
<i>Eryngium horridum</i> Malme					
Asteraceae					
<i>Baccharis articulata</i> (Lam.) Pers.		2a			
<i>Senecio brasiliensis</i> Less.				1p	
<i>Trichocline catharinensis</i> Cabr.				1p	
Erythroxylaceae					
<i>Erythroxylum</i> sp.					2r
Angiosperms - Monokotyledons					
Poaceae					
<i>Andropogon lateralis</i> Ness					5m
<i>Cortaderia selloana</i> (Schultes)Ascherson et Graebner					4m
<i>Holcus lanatus</i> L.					1p
<i>Poa</i> cf. <i>anua</i> L.		5m			
Poaceae (general)				5m	
Cyperaceae					
<i>Rhynchospora</i> sp.				1p	
Pteridophyta					
Dennstaedtiaceae					
<i>Pteridium aquilinum</i> (L.) Kuhn.		1p			
Pteridophyta					
Lycopsidea - Lycopodiaceae				2a	
<i>Lycopodium</i> sp.					1p

Table III.5: Alphabetic order of identified plants in Capoeira vegetation (vegetation record according to Braun-Blanquet)

Specimen number	Zone 3			Zone 2			
	29	53	62	7	10	12	38
Date of record	20.7.02	22.12.02	25.1.04	22.6.02	23.6.02	23.6.02	11.8.02
3. Tree section							
Altitude (in m)							
Gymnosperms							
Araucariaceae							
<i>Araucaria angustifolia</i> (Bertol.) Kuntze							
Shrub section							
Altitude (in m)	4						
Asteraceae							
<i>Baccharis uncinella</i> DC.							
Berberidaceae							
<i>Berberis laurina</i> Billb.							
Flacourtiaceae							
<i>Casearia decandra</i> Jacq.	2p						
Loganiaceae							
<i>Buddleja reitzii</i> EM Norman et LB Sm.	1p						
Melastomataceae							
<i>Miconia cinerascens</i> Miq.							
Myrtaceae							
<i>Acca sellowiana</i> (O. Berg) Burret							
<i>Calyptranthes concinna</i> DC.	1p						
Winteraceae							
<i>Drymis brasiliensis</i> Miers	2p						
Gymnosperms							
Araucariaceae							
<i>Araucaria angustifolia</i> (Bertol.) Kuntze							

Table III. 6:Alphabetic order of identified plants in campo vegetation (Vegetation record according to Braun-Blanquet)

Lamiaceae									
<i>Hyptis muelleri</i> Briquet									
Leguminosae - Faboidae									
<i>Aeschynomene</i> cf. <i>microphylla</i> Chodat et Hassl.									
<i>Agmella grissi</i>									
Leguminosae - Mimosoidae									
<i>Mimosa scabrella</i> Benth.									
Lentibulariaceae									
<i>Utricularia</i> sp. L.									
Lythraceae									
<i>Heimia myrtifolia</i> Cham. et Schlecht.									
Melastomataceae									
<i>Tibouchina gracilis</i> (Kunth) Cogn.				1p		1p			
Meliaceae									
<i>Trichilia clausenii</i> C. DC.									
Oxalidaceae									
<i>Oxalis</i> sp.				1p					
Polygalaceae									
<i>Polygala campestris</i> Gardn.				1p		1r			1p
Polygonaceae									
<i>Polygonum</i> sp.									
Rubiaceae									
<i>Borreria fastigata</i> (Griseb.) K. Schum.						1p			
<i>Borreria verticillata</i> var. <i>sulcata</i> Bacig.						1p			
Verbenaceae									
<i>Verbena bonariensis</i> L.									
<i>Verbena</i> sp. 1									
<i>Verbena</i> sp. 2									
Angiosperms - Monokotyledons									
Iridaceae									
<i>Sisyrinchium foliosum</i> IM Johnst.									
<i>Sisyrinchium laxum</i> Sims									
<i>Sisyrinchium vaginatum</i> Spreng.									

Table III. 6:Alphabetic order of identified plants in campo vegetation (vegetation record according to Braun-Blanquet)

Specimen number	Zone 2						Zone 1		
	41	48	69	71	78	19	22	37	
Date of record	11.8.02	21.12.02	31.1.04	31.1.04	2.2.04	6.7.02	7.7.02	10.8.02	
3. Tree section									
Altitude (in m)									
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze									
Shrub section									
Altitude (in m)									
Asteraceae									
<i>Baccharis uncinella</i> DC.									
Berberidaceae									
<i>Berberis laurina</i> Billb.									
Flacourtiaceae									
<i>Casearia decandra</i> Jacq.									
Loganiaceae									
<i>Buddleja reitzii</i> EM Norman et LB Sm.									
Melastomataceae									
<i>Miconia cinerascens</i> Miq.									
Myrtaceae									
<i>Acca sellowiana</i> (O. Berg) Burret									
<i>Calyptranthes concinna</i> DC.									
Winteraceae									
<i>Drymis brasiliensis</i> Miers									
Gymnosperms									
Araucariaceae									
<i>Araucaria angustifolia</i> (Bertol.) Kuntze									

Table III. 6:Alphabetic order of identified plants in campo vegetation (Vegetation record according to Braun-Blanquet)

Lamiaceae																			
<i>Hyptis muelleri</i> Briquet				1p			1p												
Leguminosae - Faboidae																			
<i>Aeschynomene</i> cf. <i>microphylla</i> Chodat et Hassl.			1r																
<i>Agmella grissi</i>	1p																		
Leguminosae - Mimosoidae																			
<i>Mimosa scabrella</i> Benth.																			
Lentibulariaceae																			
<i>Utricularia</i> sp. L.				1p															
Lythraceae																			
<i>Heimia myrtifolia</i> Cham. et Schlecht.												2p							
Melastomataceae																			
<i>Tibouchina gracilis</i> (Kunth) Cogn.				1p	1p	1p	1p	1p											
Meliaceae																			
<i>Trichilia clausenii</i> C. DC.																			
Oxalidaceae																			
<i>Oxalis</i> sp.	1p																		
Polygalaceae																			
<i>Polygala campestris</i> Gardn.																			1p
Polygonaceae																			
<i>Polygonum</i> sp.																			1p
Rubiaceae																			
<i>Borreria fastigata</i> (Griseb.) K. Schum.													1p						
<i>Borreria verticillata</i> var. <i>sulcata</i> Bacig.																			
Verbenaceae																			
<i>Verberna bonariensis</i> L.							1r												
<i>Verberna</i> sp. 1	1p																		
<i>Verberna</i> sp. 2	1r																		
Angiosperms - Monokotyledons																			
Iridaceae																			
<i>Sisyrinchium foliosum</i> IM Johnst.	1r																		
<i>Sisyrinchium laxum</i> Sims	1r																		
<i>Sisyrinchium vaginatum</i> Spreng.							1r												

Table III. 6:Alphabetic order of identified plants in campo vegetation (Vegetation record according to Braun-Blanquet)

		Zone 1	
Specimen number		42	81
Date of record		12.8.02	2.2.04
3. Tree section			
Altitude (in m)			
Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze			
Shrub section			
Altitude (in m)			
Asteraceae			
<i>Baccharis uncinella</i> DC.			
Berberidaceae			
<i>Berberis laurina</i> Billb.			
Flacourtiaceae			
<i>Casearia decandra</i> Jacq.			
Loganiaceae			
<i>Buddleja reitzii</i> EM Norman et LB Sm.			
Melastomataceae			
<i>Miconia cinerascens</i> Miq.			
Myrtaceae			
<i>Acca sellowiana</i> (O. Berg) Burret			
<i>Calyptranthes concinna</i> DC.			
Winteraceae			
<i>Drymis brasiliensis</i> Miers			
Gymnosperms			
Araucariaceae			
<i>Araucaria angustifolia</i> (Bertol.) Kuntze			

Table III. 6:Alphabetic order of identified plants in campo vegetation (Vegetation record according to Braun-Blanquet)

Herb section			
Angiosperms - Dicotyledons			
Amaranthaceae			
<i>Pfiafia cf. gnaphaloides (L.f) Mart.</i>			
Apiaceae			
<i>Eryngium cf. ebracteatum Lam.</i>			
<i>Eryngium horridum Malme</i>			
<i>Eryngium pandanifolium Cham. et Schltdl.</i>			4m
Aquifoliaceae			
Asteraceae			
<i>Achyrocline saturoides (Lam.) DC</i>			
<i>Aspilia montevidensis (Spreng.) Kuntze</i>			
<i>Baccharis articulata (Lam.) Pers.</i>			1p
<i>Baccharis axillaris DC.</i>			1p
<i>Baccharis cultrata Baker</i>			1p
<i>Baccharis microcephala Baker</i>			
<i>Baccharis cf. milleflora (Less.)</i>			
<i>Baccharis pseudovillosa Teodoro</i>			
<i>Baccharis spicata (Lam.) Baill.</i>			
<i>Baccharis tridentata Vahl.</i>			
<i>Baccharis uncinella DC.</i>			
<i>Conyza chilensis Spreng.</i>			
<i>Erigeron catharinensis Cabr.</i>			
<i>Erigeron tweediei Hook et Arn.</i>			
<i>Eupatorium betoniciforme (DC.) Bak. var. hastatum</i>			
<i>Eupatorium bupleurifolium DC.</i>			
<i>Eupatorium catharinensis</i>			
<i>Senecio brasiliensis Less.</i>			
<i>Senecio conizifolius Bak.</i>			
<i>Stevia lundiana DC.</i>			
<i>Trichocline catharinensis Cabr.</i>			
<i>Vernonia brasiliensis Less.</i>			
<i>Vernonia flexuosa Sims</i>			

Table III. 6. Alfabetic order of identified plants in campo vegetation (vegetation record according to Braun-Blanquet)

Lamiaceae			
<i>Hyptis muelleri</i> Briquet			
Leguminosae - Faboidae			
<i>Aeschynomene</i> cf. <i>microphylla</i> Chodat et Hassl.			
<i>Agmella grissi</i>			
Leguminosae - Mimosoidae			
<i>Mimosa scabrella</i> Benth.	1r		
Lentibulariaceae			
<i>Utricularia</i> sp. L.			
Lythraceae			
<i>Heimia myrtifolia</i> Cham. et Schlecht.		2p	
Melastomataceae			
<i>Tibouchina gracilis</i> (Kunth) Cogn.		1p	
Meliaceae			
<i>Trichilia clausenii</i> C. DC.			
Oxalidaceae			
<i>Oxalis</i> sp.			
Polygalaceae			
<i>Polygala campestris</i> Gardn.		1p	
Polygonaceae			
<i>Polygonum</i> sp.			
Rubiaceae			
<i>Borreria fastigata</i> (Griseb.) K. Schum.			
<i>Borreria verticillata</i> var. <i>sulcata</i> Bacig.			
Verbenaceae			
<i>Verbena bonariensis</i> L.			
<i>Verbena</i> sp. 1			
<i>Verbena</i> sp. 2			
Angiosperms - Monokotyledons			
Iridaceae			
<i>Sisyrinchium foliosum</i> IM Johnst.			
<i>Sisyrinchium laxum</i> Sims			
<i>Sisyrinchium vaginatum</i> Spreng.			

Table III. 6:Alphabetic order of identified plants in campo vegetation (Vegetation record according to Braun-Blanquet)

Poaceae			
<i>Andropogon lateralis</i> Ness		5a	
<i>Calamagrostis viridiflavescens</i> (Poir.) Steud.			
<i>Eriochrysis cayanensis</i> P. Beauvois			
<i>Paspalum pumilum</i> Nees		4m	
<i>Poa cf. anua</i> L.			
Poaceae (general)			5m
<i>Sacciolepis campestris</i> (Nees) L. R. Parodi			
Cyperaceae			
<i>Cyperus cf. eragrostis</i> Lam.			
<i>Cyperus</i> sp.			
<i>Pycurus uniloides</i> (R. BR.) Urban			
<i>Rhynchospora cf. corymbosa</i> (L.) Britton			
<i>Rhynchospora marisculus</i> Nees ex Lindl.			
<i>Rhynchospora</i> sp.			
Pteridophyta			
Blechnaceae			
<i>Blechnum imperiale</i> (Fée et Glaz.) C. Chr.			
Dennstaedtiaceae			
<i>Pteridium aquilinum</i> (L.) Kuhn.			
<i>Pteridophyta</i>			
Bryopsida - Sphagnopsida			
<i>Sphagnum</i> sp.			2a

Table III. 6. Alphabetic order of identified plants in campo vegetation (vegetation record according to Braun-Blanquet)

	Zone 3			Zone 2		Zone 1
Specimen number	1	3	30	14	16	
Date of record	4.5.02	5.5.02	21.7.02	25.6.02	25.6.02	
Vegetation type	E	Secyf	Secf	AM	P	
1. Tree section						
Altitude (in m)	15		11	12	8	
Angiosperms - Dicotyledons						
Asteraceae						
<i>Vernonia discolor</i> (Spreng.) Less						
<i>Vernonia</i> sp.			2p			
Myrtaceae						
<i>Eucalyptus</i> sp.						
Gymnosperms	3a					
Araucariaceae						
<i>Araucaria angustifolia</i> (Bertol.) Kuntze			2p	3p		
Pinaceae						
<i>Pinus taeda</i> L.					4m	
Podocarpaceae						
<i>Podocarpus lambertii</i> Klotzsch ex Endl.						
2. Tree section						
Altitude (in m)						
Angiosperms - Dicotyledons	9		8			
<i>Leguminosae</i> (general)			2p			
Myrtaceae						
<i>Eucalyptus</i> sp.		2a				
Gymnosperms						
Araucariaceae						
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p					

Table III. 7: Alphabetic order of identified plants in anthropogenic affected areas (vegetation record according to Braun-Blanquet)

3. Tree section					
Altitude (in m)	5			5	
Angiosperms - Dicotyledons					
Asteraceae					
<i>Dasyphyllum tomentosum</i> (Spreng.) Cabr.				2p	
<i>Vernonia</i> sp.				2p	
Moraceae					
<i>Sorocea bonplandii</i> (Baill.) Burger				2p	
Myrtaceae					
<i>Eucalyptus</i> sp.	2a				
Myrtaceae (general)				2p	
Gymnosperms					
Araucariaceae					
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p				
Podocarpaceae					
<i>Podocarpus lambertii</i> Klotzsch ex Endl.	2p				
Shrub section					
Altitude (in m)	3			3	
Moraceae					
<i>Sorocea bonplandii</i> (Baill.) WC Burger				1p	
Myrsinaceae					
<i>Myrsine</i> sp.				2p	
Myrtaceae					
<i>Eucalyptus</i> sp.	2p				
Angiosperms - Monocotyledons					
Poaceae					
<i>Bambus</i> sp.				4a	
Gymnosperms					
Araucariaceae					
<i>Araucaria angustifolia</i> (Bertol.) Kuntze	2p				
Podocarpaceae					
<i>Podocarpus lambertii</i> Klotzsch ex Endl.	2a			2p	

Table III. 7: Alphabetic order of identified plants in anthropogenic affected areas (vegetation record according to Braun-Blanquet)

Herb section							
Angiosperms - Dicotyledons							
Asteraceae							
<i>Hypochoeris</i> sp. L.							1r
<i>Solidago chilensis</i> Meyen.							1r
Melastomataceae							
<i>Leandra</i> sp.			1p				
<i>Miconia cinerascens</i> Miq.			1p				1p
<i>Miconia hiemalis</i> St. Hil.					1p		1p
Myrsinaceae							
<i>Myrsine</i> sp.					1p		
Solanaceae							
<i>Solanum variabile</i> Mart.							1p
Angiosperms - Monokotyledons							
Poaceae							
<i>Poaceae (general)</i>			r				4m
Pteridophyta							
Dennstaedtiaceae							
<i>Pteridium aquilinum</i> (L.) Kuhn.			1p				
Angiosperms - Dicotyledons							
Cactaceae							
<i>Rhipsalis</i> sp.							r
Angiosperms - Monokotyledons							
Bromeliaceae							
<i>Bromelia</i> sp.							p
<i>Tillandsia usneoides</i> (L.) L.							p
<i>Oncidium concolor</i> Hooker					p		p
<i>Pleurothallis hygrophila</i> Barb. Rodr.					p		

Table III. 7: Alphabetic order of identified plants in anthropogenic affected areas (vegetation record according to Braun-Blanquet)

Appendix IV

Table with data of the geographical position (GPS), altitude and vegetation types of the sample points

	Lat - South	Long - West	
specimen number	Deg Min Sec	Deg Min Sec	Altitude (in m) Space shuttle
1	29° 18' 59,40"	50° 50' 33,60"	770
2	29° 18' 50,04"	50° 54' 36,96"	528
3	29° 18' 55,74"	50° 51' 37,20"	752
4	29° 19' 14,70"	50° 53' 10,62"	612
5	29° 21' 52,68"	50° 47' 29,82"	832
6	29° 13' 34,50"	50° 44' 29,82"	847
7	29° 13' 34,74"	50° 44' 23,70"	846
8	29° 08' 42,54"	50° 48' 32,76"	901
9	29° 04' 48,48"	50° 37' 58,20"	852
10	29° 04' 46,68"	50° 38' 07,08"	848
11	29° 04' 38,40"	50° 38' 01,20"	826
12	29° 18' 29,70"	50° 29' 42,42"	934
13	29° 19' 48,12"	50° 30' 30,06"	892
14	29° 20' 19,86"	50° 40' 23,70"	877
15	29° 20' 24,66"	50° 40' 26,04"	882
16	29° 12' 18,54"	50° 14' 37,14"	937
17	29° 10' 43,44"	50° 16' 00,36"	961
18	29° 03' 24,72"	50° 07' 11,40"	1004
19	29° 07' 16,20"	50° 06' 07,62"	1006
20	29° 09' 38,64"	50° 04' 38,82"	922
21	29° 09' 38,10"	50° 05' 28,50"	935
22	29° 04' 07,68"	50° 01' 34,74"	1005
23	29° 11' 28,80"	50° 13' 37,14"	949
24	29° 20' 19,08"	51° 09' 20,94"	115
25	29° 19' 19,62"	51° 08' 30,30"	319
26	29° 15' 46,38"	51° 01' 59,94"	732
27	29° 12' 58,08"	51° 01' 24,36"	732
28	29° 12' 15,90"	51° 00' 28,50"	777
29	29° 12' 12,90"	51° 00' 28,80"	785
30	29° 10' 56,70"	51° 00' 26,70"	789
31	29° 12' 10,44"	51° 04' 09,42"	535
32	29° 13' 03,18"	51° 08' 49,44"	726
33	29° 23' 07,38"	50° 47' 01,62"	854
34	29° 19' 03,84"	50° 58' 23,64"	326
35	29° 19' 49,98"	50° 55' 16,44"	771
36	29° 15' 15,00"	50° 20' 10,50"	878
37	29° 15' 17,22"	50° 20' 13,74"	890

Table IV: Geographical position, altitude and vegetation type of sample points

38	29° 01' 12,30"	50° 33' 09,78"	942
39	28° 56' 34,86"	50° 28' 09,60"	841
40	29° 09' 13,26"	50° 39' 55,68"	920
41	29° 09' 14,64"	50° 39' 55,20"	920
42	29° 13' 18,36"	50° 28' 20,52"	949
43	29° 27' 19,50"	50° 20' 44,80"	925
44	29° 29' 50,20"	50° 21' 03,30"	882
45	29° 31' 10,70"	50° 19' 08,10"	307
46	29° 10' 56,50"	50° 12' 37,30"	1020
47	29° 15' 00,00"	50° 20' 55,26"	874
48	29° 15' 01,20"	50° 20' 59,28"	880
49	29° 09' 54,00"	50° 31' 24,42"	966
50	29° 06' 32,64"	50° 35' 28,20"	940
51	29° 07' 42,36"	50° 48' 03,90"	911
52	29° 07' 39,72"	50° 48' 11,04"	940
53	29° 09' 40,08"	50° 50' 08,40"	869
54	29° 25' 41,94"	51° 09' 53,94"	528
55	29° 24' 45,72"	51° 08' 05,70"	506
56	29° 26' 48,18"	51° 02' 50,28"	383
57	29° 22' 41,16"	51° 03' 27,84"	616
58	29° 22' 32,64"	50° 50' 06,78"	831
59	29° 21' 24,96"	51° 04' 22,50"	663
60	29° 06' 09,96"	51° 03' 36,18"	710
61	29° 06' 50,46"	51° 02' 48,18"	756
62	29° 09' 09,54"	50° 56' 52,98"	741
63	29° 10' 32,88"	50° 54' 57,00"	840
64	29° 13' 08,04"	50° 53' 32,88"	763
65	29° 13' 02,40"	50° 53' 36,24"	771
66	29° 13' 53,16"	50° 55' 33,18"	832
67	29° 26' 09,36"	50° 35' 55,92"	879
68	29° 23' 00,06"	50° 38' 22,20"	862
69	29° 17' 48,36"	50° 39' 52,74"	825
70	29° 16' 41,10"	50° 38' 12,90"	874
71	29° 14' 16,02"	50° 34' 36,72"	940
72	29° 09' 09,00"	50° 34' 17,16"	910
73	29° 05' 03,72"	50° 43' 57,78"	857
74	29° 04' 04,14"	50° 55' 12,72"	862
75	29° 06' 40,20"	50° 50' 36,30"	940
76	29° 08' 06,54"	50° 41' 46,92"	959
77	29° 09' 52,02"	50° 49' 33,48"	808
78	29° 10' 49,02"	50° 46' 51,54"	864

Table IV: Geographical position, altitude and vegetation type of sample points

79	29° 10' 14,70"	50° 31' 48,66"	918
80	29° 05' 11,28"	50° 26' 31,80"	941
81	29° 01' 43,26"	50° 25' 45,66"	959
82	29° 07' 47,04"	50° 26' 29,34"	941
83	29° 08' 45,70"	50° 18' 19,00"	872
84	29° 06' 10,70"	50° 19' 47,90"	899
85	29° 01' 53,70"	50° 07' 15,80"	1028

legend:

- Araucaria* forest
- Semi-deciduous and deciduous forest
- Capão
- Gallery forest
- Capoeira
- Grassland (Campo)
- Anthropogenic affected area

Table IV: Geographical position, altitude and vegetation type of sample points

Bildungsgang

Name: Kerstin Kriegel

Geburtstag: 12. Januar 1977

Geburtsort: Dresden

Schulbildung: 09/83 – 07/91 Polytechnische Oberschule „Alexander Puschkin“ in Dresden
09/91 – 07/92 Mittelschule „Alexander Puschkin“ in Dresden
09/92 – 06/95 Gymnasium Dresden - Großschachwitz

Schulabschluss: **Abitur**

Studium: 10/95 – 04/01 Studium der Biologie an der Technischen Universität Dresden

Studienabschluss: **Diplombiologin**

Promotion: 01/02 – 06/06 Anfertigung der Dissertation am Institut für Geowissenschaften, Lehrstuhl für Biogeologie und Angewandte Paläontologie an der Eberhard –Karls – Universität –

Thema der Dissertation:

”Recent palynological spectra from the Planalto region of the Serra Geral, Rio Grande do Sul/ Brazil: Reconstruction of contemporary vegetation – a case study”

Betreuer: Prof. Dr. Dr. h.c. V. Mosbrugger
Prof. Dr. em. H. Walther